A Combined Epidemiologic and Metabolomic Approach Improves CKD Prediction

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ABSTRACT

Metabolomic approaches have begun to catalog the metabolic disturbances that accompany CKD, but whether metabolite alterations can predict future CKD is unknown. We performed liquid chromatography/mass spectrometry–based metabolite profiling on plasma from 1434 participants in the Framingham Heart Study (FHS) who did not have CKD at baseline. During the following 8 years, 123 individuals developed CKD, defined by an estimated GFR of <60 ml/min per 1.73 m². Numerous metabolites were associated with incident CKD, including 16 that achieved the Bonferroni-adjusted significance threshold of \( P \approx 0.00023 \). To explore how the human kidney modulates these metabolites, we profiled arterial and renal venous plasma from nine individuals. Nine metabolites that predicted CKD in the FHS cohort decreased more than creatinine across the renal circulation, suggesting that they may reflect non–GFR-dependent functions, such as renal metabolism and secretion. Urine isotope dilution studies identified citrulline and choline as markers of renal metabolism and kynurenic acid as a marker of renal secretion. In turn, these analytes remained associated with incident CKD in the FHS cohort, even after adjustment for eGFR, age, sex, diabetes, hypertension, and proteinuria at baseline. Addition of a multi-marker metabolite panel to clinical variables significantly increased the \( c \)-statistic (0.77–0.83, \( P < 0.0001 \)); net reclassification improvement was 0.78 (95% confidence interval, 0.60 to 0.95; \( P < 0.0001 \)). Thus, the addition of metabolite profiling to clinical data may significantly improve the ability to predict whether an individual will develop CKD by identifying predictors of renal risk that are independent of estimated GFR.

Given the significant morbidity and mortality attributable to established CKD, early markers of CKD risk are needed. The kidneys can modulate circulating small-molecule levels through a variety of mechanisms, such as filtration, reabsorption, secretion, and metabolism (including both catabolism and anabolism). Currently available metrics of kidney function (serum creatinine and urea) primarily reflect relatively advanced impairment in renal filtration. In contrast, early and specific disease markers may reflect impairments along other axes of renal small-molecule handling.

Metabolite profiling technologies enable high-throughput, high-resolution metabolic phenotyping of human plasma. Applied to well-characterized human cohorts, these techniques have the potential to identify novel disease biomarkers and to highlight their underlying metabolic pathways. For example, we have previously applied liquid chromatography-mass spectrometry (LC-MS)–based metabolite profiling to identify numerous metabolite alterations that accompany ESRD. Similar findings have been observed using capillary electrophoresis-mass spectrometry (CE-MS)–based metabolite profiling of plasma obtained from individuals across a spectrum of extant kidney disease. Whether any of these metabolite perturbations presage the development of clinically overt kidney disease is unknown.

Here, we report the application of metabolite profiling to plasma obtained from participants in the Framingham Heart Study (FHS). Prior work in the FHS has highlighted branched-chain and aromatic amino acids as robust predictors of future type 2 diabetes. Because of access to archived plasma samples, detailed phenotyping, and longitudinal follow-up on clinical outcomes, this sample provides an ideal opportunity to identify novel markers of CKD risk. To explore how select CKD predictors are modulated by the human kidney, we also performed metabolite profiling of plasma and urine samples obtained from individuals undergoing aortic and renal vein catheterization. Taken together, these studies demonstrate the broad effect kidney function has on the plasma metabolome and show how this perspective can improve CKD prediction beyond estimated GFR (eGFR) and other established CKD risk factors.

### Results

#### Baseline Characteristics of the Epidemiologic Study Sample

We performed a prospective study of incident CKD in the FHS. Among all 1434 eligible participants with an eGFR ≥ 60 ml/min per 1.73 m² at baseline, we identified 123 individuals who developed new-onset CKD (eGFR < 60 ml/min per 1.73 m²) during an 8-year follow-up period (see Concise Methods). Baseline characteristics of the FHS study sample are shown in Table 1. The participants who subsequently developed CKD were older (61 versus 54 years), had a higher baseline prevalence of hypertension (62% versus 33%) and diabetes (17% versus 5%), and had a trend for a higher prevalence of proteinuria (33% versus 24%). Mean baseline eGFR was 86.5 ± 30.1 ml/min per 1.73 m² for those who subsequently developed CKD and 94.3 ± 25.7 ml/min per 1.73 m² for those who remained CKD-free during the study interval (P = 0.0017). At 8 years of follow-up, eGFR was 49.7 ± 8.8 ml/min per 1.73 m² for cases and 89.0 ± 17.1 ml/min per 1.73 m² for those who remained free of CKD (P < 0.0001).

### Select Metabolites Are Associated with Incident CKD in FHS

We performed metabolite profiling on fasting plasma obtained from the baseline examination. The three components of the metabolite profiling platform quantified 54 positively charged polar analytes, 59 negatively charged polar analytes, and 104 lipids. For each of the 217 metabolites, Figure 1A shows the mean ratio of each analyte in patients who went on to develop CKD (cases) versus those who did not, plotted against their corresponding P values (full results are shown in Supplementary Table 1). Sixteen metabolites achieved a conservative Bonferroni-adjusted significance threshold of P ≤ 0.00023 (i.e., 0.05/217 metabolites), with 13 of these metabolites higher in cases relative to persons who remained CKD-free (Table 2). Several metabolites derive from the same metabolic pathway (e.g., tryptophan metabolites [kynurenic acid, kynurenine, 5-hydroxyindoleacetic acid, quinolinolate], choline derivatives [choline, trimethylamine-N-oxide], citric acid cycle intermediates [aconitate, isocitrate], and purine metabolites [xanthosine, adenosine]). The only lipid analytes that achieved Bonferroni-adjusted significance, lysophosphatidylcholine 18:1 and lysophosphatidylcholine 18:2, were both lower in cases. Notably, the association between creatinine and case status did not reach Bonferroni-level significance.

For the 16 metabolites listed in Table 2, Figure 1B plots the metabolite ratio in cases relative to participants who remained CKD-free against each metabolite’s correlation with baseline eGFR across the study sample. As expected, creatinine was negatively correlated with baseline eGFR (R = −0.16); no other

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**Table 1. Baseline characteristics of the FHS study sample**

<table>
<thead>
<tr>
<th>Clinical Characteristics</th>
<th>Individuals Who Developed CKD (n=123)</th>
<th>Individuals Who Did Not Develop CKD (n=1311)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>61 ± 8b</td>
<td>54 ± 9</td>
</tr>
<tr>
<td>Women (%)</td>
<td>59b</td>
<td>50</td>
</tr>
<tr>
<td>Body mass index (kg/m²)</td>
<td>28.1 ± 4.4</td>
<td>27.6 ± 5.0</td>
</tr>
<tr>
<td>Hypertension (%)</td>
<td>62a</td>
<td>33</td>
</tr>
<tr>
<td>Diabetes (%)</td>
<td>17a</td>
<td>5</td>
</tr>
<tr>
<td>eGFR (ml/min per 1.73 m²)</td>
<td>86.5 ± 30.1b</td>
<td>94.3 ± 25.7</td>
</tr>
<tr>
<td>eGFR at follow-up (ml/min per 1.73 m²)</td>
<td>49.7 ± 8.8a</td>
<td>89.0 ± 17.1</td>
</tr>
<tr>
<td>Dipstick proteinuria (%)</td>
<td>33</td>
<td>24</td>
</tr>
</tbody>
</table>

Values expressed with plus/minus sign mean ± SD.  
aP < 0.0001.  
bP < 0.05.
Arteriovenous Plasma Sampling Demonstrates Heterogeneous Renal Metabolite Handling

To explore the various axes of renal metabolite handling (i.e., beyond renal filtration alone), we collected arterial and renal venous plasma from nine individuals undergoing invasive catheterization (see Concise Methods). These patients had a mean eGFR of 65.4 ± 8.7 ml/min per 1.73 m² (Table 3). As expected, creatinine levels decreased from the arterial to renal venous plasma sample in all nine individuals (Figure 2A); the mean ratio of venous to arterial creatinine level (V/A) was 0.84 (P < 0.0001). Figure 2B demonstrates the strong correlation between renal venous creatinine level as measured by the LC-MS platform and serum creatinine measured by the clinical laboratory on a peripheral venous sample drawn on the same day (R = 0.95).

Figure 2C depicts the range of arteriovenous gradients for 125 polar (left panel) and 102 lipid (right panel) analytes measured in all samples, ordered by mean V/A (full results are shown in Supplementary Table 2). Most polar analyte levels were decreased in the renal vein relative to the aorta, although several metabolite levels increased across the kidney. By contrast, lipid metabolite levels were not appreciably altered in transit from the arterial to the renal venous circulation. For any given metabolite, V/A < 1 is consistent with net uptake by the kidney, whether via filtration, secretion, and/or metabolism (catabolism), whereas V/A > 1 suggests net release by the kidney (anabolism). Thus, our physiologic data demonstrate how different metabolites could potentially serve as markers of different aspects of renal function.

Combined Epidemiologic and Physiologic Approach Nominates Markers of CKD Risk

For the 16 metabolites that achieved Bonferroni-level significance for incident CKD in FHS, the V/A values across all nine patients who underwent catheterization are shown in Table 4. Because creatinine excretion is primarily a function of glomerular filtration, we hypothesized that metabolites that decrease more than creatinine across the renal circulation are potential markers of non-GFR renal function (e.g., renal metabolism and secretion). Thus, we focused on metabolites that had a mean V/A less than that for creatinine (< 0.84) and demonstrated a decrease across the kidney in all nine individuals undergoing catheterization. As a result, we highlight nine polar metabolites as potential markers of CKD risk: citrulline (urea cycle intermediate); choline; kynurenic acid, kynurenine, and 5-hydroxyindoleacetic acid (tryptophan metabolites); aconitate and isocitrate (citric acid cycle intermediates); xanthosine (purine metabolism); and β-aminoisobutyric acid (pyrimidine metabolite).

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>Geometric Mean Ratio in FHS (95% CI)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Xanthosine</td>
<td>1.19 (1.13 to 1.27)</td>
<td>5.2 x 10^-9</td>
</tr>
<tr>
<td>Citrulline</td>
<td>1.13 (1.08 to 1.18)</td>
<td>2.0 x 10^-8</td>
</tr>
<tr>
<td>Isocitrinate</td>
<td>1.14 (1.09 to 1.20)</td>
<td>9.1 x 10^-8</td>
</tr>
<tr>
<td>Aconitate</td>
<td>1.13 (1.08 to 1.19)</td>
<td>9.4 x 10^-8</td>
</tr>
<tr>
<td>Choline</td>
<td>1.08 (1.05 to 1.12)</td>
<td>3.7 x 10^-7</td>
</tr>
<tr>
<td>Kynurenine</td>
<td>1.13 (1.08 to 1.19)</td>
<td>4.1 x 10^-7</td>
</tr>
<tr>
<td>β-aminobutyric acid</td>
<td>1.22 (1.13 to 1.34)</td>
<td>1.1 x 10^-6</td>
</tr>
<tr>
<td>Kynurenic acid</td>
<td>1.25 (1.14 to 1.38)</td>
<td>1.4 x 10^-6</td>
</tr>
<tr>
<td>Trimethylamine-N-oxide</td>
<td>1.33 (1.18 to 1.49)</td>
<td>1.6 x 10^-6</td>
</tr>
<tr>
<td>Adenosine</td>
<td>1.50 (1.27 to 1.77)</td>
<td>2.0 x 10^-6</td>
</tr>
<tr>
<td>S-hydroxyindoleacetic acid</td>
<td>0.73 (0.64 to 0.83)</td>
<td>3.3 x 10^-6</td>
</tr>
<tr>
<td>Quinolinic acid</td>
<td>1.19 (1.10 to 1.28)</td>
<td>5.1 x 10^-6</td>
</tr>
<tr>
<td>LPC18:2</td>
<td>0.90 (0.85 to 0.94)</td>
<td>1.9 x 10^-5</td>
</tr>
<tr>
<td>Sucrose</td>
<td>1.39 (1.19 to 1.63)</td>
<td>4.7 x 10^-5</td>
</tr>
<tr>
<td>LPC18:1</td>
<td>0.91 (0.86 to 0.95)</td>
<td>1.0 x 10^-4</td>
</tr>
<tr>
<td>Inositol</td>
<td>1.19 (1.09 to 1.30)</td>
<td>1.2 x 10^-4</td>
</tr>
</tbody>
</table>

Table 2. Metabolites significantly associated with incident CKD in FHS

LPC, lysophosphatidylcholine.

metabolite had as strong a correlation with eGFR from the initial sample. Further, there was no relationship between the magnitude of metabolite associations with CKD risk and their correlation with baseline eGFR. Thus, the metabolites associated with incident CKD in FHS are not simply markers of impaired renal filtration.

Plasma and Urinary Profiling of Select Metabolites Identifies Markers of Renal Metabolism and Secretion

In addition to identifying metabolites that decrease more than creatinine across the renal arteriovenous circulation, our physiologic data identify several metabolites that increased from artery to vein in all nine individuals undergoing catheterization, indicating net release by the kidney (Table 4). For the current study, we emphasize the increase in plasma
arginine across the kidney (mean V/A, 1.20; \(P = 0.0010\)), consistent with the kidney’s established role in extracting circulating citrulline and converting it to arginine.\(^6,7\) We also note the uniform increase in plasma \(\alpha\)-glycerophosphocholine (mean V/A, 1.24; \(P = 0.0065\)) across the kidney in our study sample, as net release of \(\alpha\)-glycerophosphocholine would be contingent on net uptake of choline or choline derivatives from the circulation. Of note, \(\alpha\)-glycerophosphocholine is an osmolyte known to be produced in the renal medulla in response to changes in extracellular tonicity.\(^8\) Betaine, another renal osmolyte derived from choline, increased from artery to renal vein in seven of the nine individuals undergoing catheterization (Supplementary Table 2).\(^9\)

Using stable isotope dilution, we compared absolute levels of select metabolites in plasma with the corresponding levels in urine obtained from individuals undergoing catheterization. Using the urine-to-plasma creatinine ratio as an index of glomerular filtration, we assessed the urinary fractional excretion of citrulline, choline, and kynurenic acid (Table 5). Across these individuals, the fractional excretion for citrulline and choline were significantly \(>100\%\), consistent with their net uptake and metabolism (e.g., to arginine and \(\alpha\)-glycerophosphocholine, respectively) by the kidney. By contrast, the fractional excretion for kynurenic acid was \(>100\%\), invoking tubular secretion in addition to glomerular filtration as a means of kynurenic acid excretion. Taken together, these data are consistent with the hypothesis that metabolites with mean V/A less than that for creatinine are markers of non-GFR renal function.

Metabolite Predictors of Future CKD after Multivariable Adjustment

Because we found that select candidate metabolites decreased more than creatinine across the renal circulation and confirmed that three of these are markers of renal metabolism or secretion, we hypothesized that these analytes would be associated with incident CKD in FHS, even after accounting for differences in glomerular filtration. Thus, logistic regression models were fitted to assess the association between the nine candidate metabolites and future CKD, adjusting for eGFR, as well as other established CKD risk factors, including age, sex, diabetes, hypertension, and proteinuria at baseline.

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**Table 3.** Individuals undergoing right and left heart catheterization (with renal arteriovenous plasma sampling)

<table>
<thead>
<tr>
<th>Patient</th>
<th>Reason for Procedure</th>
<th>Serum Creatinine (mg/dl)</th>
<th>eGFR (ml/min per 1.73m²)</th>
<th>Age (yr)</th>
<th>Sex</th>
<th>Type 2 Diabetes</th>
<th>ACE/ARB</th>
<th>Coronary Disease</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Preoperative (aneurysm repair)</td>
<td>1.28</td>
<td>58.1</td>
<td>76</td>
<td>M</td>
<td>No</td>
<td>No</td>
<td>Yes</td>
</tr>
<tr>
<td>2</td>
<td>Aortic stenosis</td>
<td>1.1</td>
<td>67.8</td>
<td>84</td>
<td>M</td>
<td>Yes</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>3</td>
<td>Shortness of breath</td>
<td>1.19</td>
<td>65.2</td>
<td>65</td>
<td>M</td>
<td>No</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>4</td>
<td>Preoperative (mitral valve repair)</td>
<td>1.28</td>
<td>59.1</td>
<td>70</td>
<td>M</td>
<td>No</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>5</td>
<td>Aortic stenosis</td>
<td>1.38</td>
<td>51.9</td>
<td>86</td>
<td>M</td>
<td>No</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>6</td>
<td>Aortic stenosis and mitral regurgitation</td>
<td>1.21</td>
<td>61.5</td>
<td>79</td>
<td>M</td>
<td>No</td>
<td>Yes</td>
<td>No</td>
</tr>
<tr>
<td>7</td>
<td>Shortness of breath</td>
<td>1.03</td>
<td>78.6</td>
<td>59</td>
<td>M</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
<tr>
<td>8</td>
<td>Mitral regurgitation</td>
<td>0.9</td>
<td>70.7</td>
<td>49</td>
<td>F</td>
<td>No</td>
<td>No</td>
<td>No</td>
</tr>
<tr>
<td>9</td>
<td>Aortic stenosis</td>
<td>1.02</td>
<td>75.5</td>
<td>76</td>
<td>M</td>
<td>Yes</td>
<td>Yes</td>
<td>Yes</td>
</tr>
</tbody>
</table>

ACE/ARB, angiotensin-converting enzyme inhibitor/angiotensin-receptor blocker; M, male; F, female.

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**Figure 2.** Renal arteriovenous sampling demonstrates heterogeneous metabolite handling. (A) Creatinine levels (peak areas) in aortic and renal venous plasma measured by the mass spectrometer. (B) Correlation between renal venous creatinine levels (peak areas) measured by the mass spectrometer and peripheral venous creatinine (mg/dl) measured by the clinical laboratory. (C) Mean renal venous to arterial metabolite ratios for polar (left panel) and lipid (right panel) metabolites.
Metabolites associated with incident CKD in FHS

Metabolites that undergo net release by the kidney

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>Mean V/A</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Xanthosine</td>
<td>0.45</td>
<td>6.7 × 10⁻⁵</td>
</tr>
<tr>
<td>β-aminoisobutyric acid</td>
<td>0.50</td>
<td>4.0 × 10⁻⁵</td>
</tr>
<tr>
<td>Choline</td>
<td>0.58</td>
<td>1.4 × 10⁻⁵</td>
</tr>
<tr>
<td>Kynurenic acid</td>
<td>0.59</td>
<td>7.0 × 10⁻⁵</td>
</tr>
<tr>
<td>Citrulline</td>
<td>0.65</td>
<td>9.0 × 10⁻⁶</td>
</tr>
<tr>
<td>Aconitate</td>
<td>0.70</td>
<td>9.9 × 10⁻⁵</td>
</tr>
<tr>
<td>Kynurenic acid</td>
<td>0.70</td>
<td>3.1 × 10⁻⁵</td>
</tr>
<tr>
<td>S-hydroxyindoleacetic acid</td>
<td>0.71</td>
<td>9.3 × 10⁻⁴</td>
</tr>
<tr>
<td>Adenosine</td>
<td>0.73</td>
<td>2.8 × 10⁻²</td>
</tr>
<tr>
<td>Isocitrate</td>
<td>0.81</td>
<td>4.0 × 10⁻⁵</td>
</tr>
<tr>
<td>Inositol</td>
<td>0.83</td>
<td>4.5 × 10⁻³</td>
</tr>
<tr>
<td>Trimethylamine-N-oxide</td>
<td>0.85</td>
<td>4.9 × 10⁻³</td>
</tr>
<tr>
<td>Quinolinic acid</td>
<td>0.86</td>
<td>3.1 × 10⁻²</td>
</tr>
<tr>
<td>LPC18:1</td>
<td>0.97</td>
<td>1.9 × 10⁻¹</td>
</tr>
<tr>
<td>LPC18:2</td>
<td>0.99</td>
<td>6.6 × 10⁻¹</td>
</tr>
<tr>
<td>Sucrose</td>
<td>1.00</td>
<td>5.5 × 10⁻¹</td>
</tr>
</tbody>
</table>

Metabolites that undergo net release by the kidney

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>Mean V/A</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serine</td>
<td>1.53</td>
<td>4.6 × 10⁻⁴</td>
</tr>
<tr>
<td>α-glycerophosphocholine</td>
<td>1.24</td>
<td>6.5 × 10⁻³</td>
</tr>
<tr>
<td>Arginine</td>
<td>1.20</td>
<td>9.6 × 10⁻⁴</td>
</tr>
<tr>
<td>Niacinamide</td>
<td>1.17</td>
<td>2.9 × 10⁻³</td>
</tr>
<tr>
<td>Alanine</td>
<td>1.16</td>
<td>1.4 × 10⁻⁴</td>
</tr>
<tr>
<td>Threonine</td>
<td>1.11</td>
<td>3.1 × 10⁻⁴</td>
</tr>
<tr>
<td>Serine</td>
<td>1.53</td>
<td>4.6 × 10⁻⁴</td>
</tr>
</tbody>
</table>

LPC, lysophosphatidylcholine.

Table 4. Selected renal arteriovenous metabolite gradients

Table 5. Fractional excretion of citrulline, choline, and kynurenic acid

DISCUSSION

Our study has two principal findings. First, we provide a broad perspective on the metabolite perturbations that are associated with incipient kidney disease. Second, we highlight a subset of these metabolites that decrease more than creatinine across the renal arteriovenous circulation and find that these metabolites predict incident CKD even after adjustment for eGFR, age, sex, diabetes, hypertension, and proteinuria at baseline. Taken

(logistic model to construct a multimarker panel of five metabolites: kynurenic acid, xanthosine, 5-hydroxyindoleacetic acid, kynurenine, and citrulline. The incremental predictive value of the multimarker panel on top of the base model is shown in Table 7. The OR per SD increment in log multimarker panel for incident CKD was 2.41 (95% CI, 1.93 to 3.02). The addition of the multimarker panel on top of the base model is shown in Table 7. The OR per SD increment in log multimarker panel for incident CKD was 2.41 (95% CI, 1.93 to 3.02). The addition of the multimarker panel led to an improvement in discrimination, as shown by the increase in the c-statistic (0.77–0.83; P<0.0001). Furthermore, the multimarker panel also led to a significant improvement in classification accuracy, with a “category-free” net reclassification improvement of 0.78 (95% CI, 0.60 to 0.95; P<0.0001) and integrated discrimination index of 0.074 (95% CI, 0.050 to 0.097; P<0.0001). These results were minimally attenuated with a more parsimonious multimarker panel consisting of kynurenic acid, xanthosine, and 5-hydroxyindoleacetic acid (Table 7).

DISCUSSION

Our study has two principal findings. First, we provide a broad perspective on the metabolite perturbations that are associated with incipient kidney disease. Second, we highlight a subset of these metabolites that decrease more than creatinine across the renal arteriovenous circulation and find that these metabolites predict incident CKD even after adjustment for eGFR, age, sex, diabetes, hypertension, and proteinuria at baseline. Taken
Table 6. Relationship of baseline metabolite levels and incident CKD

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>Odds Ratio per SD (95% CI)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Kynurenic acid</td>
<td>1.53 (1.25 to 1.88)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Kynurenine</td>
<td>1.49 (1.22 to 1.83)</td>
<td>0.001</td>
</tr>
<tr>
<td>Citrulline</td>
<td>1.48 (1.19 to 1.83)</td>
<td>0.0004</td>
</tr>
<tr>
<td>Choline</td>
<td>1.46 (1.17 to 1.82)</td>
<td>0.0007</td>
</tr>
<tr>
<td>Xanthosine</td>
<td>1.46 (1.21 to 1.76)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>β-aminoisobutyric acid</td>
<td>1.41 (1.15 to 1.72)</td>
<td>0.0008</td>
</tr>
<tr>
<td>Aconitate</td>
<td>1.32 (1.07 to 1.62)</td>
<td>0.0092</td>
</tr>
<tr>
<td>Isocitrate</td>
<td>1.28 (1.05 to 1.58)</td>
<td>0.017</td>
</tr>
<tr>
<td>5-hydroxyindoleacetic acid</td>
<td>0.62 (0.51 to 0.76)</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

Odds ratios for CKD obtained from logistic regressions. All models adjusted for eGFR, age, sex, diabetes, hypertension, and proteinuria at baseline.

Table 7. Multimarker panels and prediction of incident CKD

<table>
<thead>
<tr>
<th>Variable</th>
<th>Multimarker Panel: 5 Metabolites</th>
<th>Multimarker Panel: 3 Metabolites</th>
</tr>
</thead>
<tbody>
<tr>
<td>OR per SD</td>
<td>2.41 (1.93 to 3.02)</td>
<td>2.15 (1.74 to 2.65)</td>
</tr>
<tr>
<td>c-statistics</td>
<td>0.77</td>
<td>0.77</td>
</tr>
<tr>
<td>Base model</td>
<td>0.83</td>
<td>0.82</td>
</tr>
<tr>
<td>Base model +</td>
<td></td>
<td></td>
</tr>
<tr>
<td>multimarker panel</td>
<td></td>
<td></td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.0001</td>
<td>0.0002</td>
</tr>
<tr>
<td>NRI (category-free)</td>
<td>0.78 (0.60 to 0.95)</td>
<td>0.79 (0.61 to 0.96)</td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>IDI</td>
<td>0.074 (0.050 to 0.097)</td>
<td>0.062 (0.041 to 0.083)</td>
</tr>
<tr>
<td>P value</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

Five metabolites were kynurenic acid, xanthosine, 5-hydroxyindoleacetic acid, kynurenine, and citrulline. Three metabolites were kynurenic acid, xanthosine, and 5-hydroxyindoleacetic acid. Values in parentheses are 95% CIs.

Together, these findings highlight the potential value of integrating different axes of renal function in an assessment of kidney health and prognosis.

At present, clinical assessment of kidney function relies primarily on measurement of serum creatinine levels. Because creatinine is freely filtered at the glomerulus, is not reabsorbed, and undergoes only limited tubular secretion, serum creatinine is used as a noninvasive index of GFR. By design, these measures do not assess fundamental renal functions beyond glomerular filtration.

Our study of individuals undergoing catheterization demonstrates the heterogeneous effect of human kidney function on the plasma metabolome. Although arterial and renal venous metabolite levels alone do not quantify the relative effects of glomerular filtration, tubular absorption/secretion, and organ metabolism, we highlight select metabolites at each extreme of V/A (Figure 2C). For example, a metabolite with a V/A > 1 is to some extent being made in the kidney. In that regard, we note that our finding of renal anabolism of arginine, serine, and alanine recapitulates prior studies that have focused on renal amino acid handling. By contrast, metabolites with V/A substantially lower than that for creatinine are to some extent being metabolized and/or secreted, as corroborated by our fractional excretion data for citrulline, choline, and kynurenic acid.

Regardless of CKD cause, tubulointerstitial injury is an invariant pathologic feature of chronic renal disease. Indeed, the degree of tubulointerstitial disease correlates better than glomerular injury with disease prognosis, even for primary glomerular diseases. In the current study, we show that markers of renal metabolism and tubular secretion predict incident CKD in the FHS, even after adjustment for eGFR, age, sex, diabetes, hypertension, and proteinuria at baseline.

Although we hypothesize that the metabolite predictors of CKD in FHS provide information about renal health orthogonal to GFR, we recognize that select risk markers could also play a functional role in CKD progression. Impaired metabolism of citrulline could limit arginine bioavailability for nitric oxide synthesis. Elevated levels of choline, as well as trimethylamine-N-oxide, have recently been implicated in atherogenesis. IDO1 encodes the major enzyme that catalyzes the first and rate-limiting step of tryptophan to kynurenine, which is subsequently metabolized to kynurenic acid. Ischemia reperfusion injury induces renal tubular Id1 expression in mice, and Id1 mice are protected from ischemia reperfusion injury compared with wild-type mice. Further, functional roles for both kynurenine and kynurenic acid have been described in the context of inflammation and vascular tone. Notably, 5-hydroxyindoleacetic acid production results from an alternative pathway of kynurenine, and 5-hydroxyindoleacetic acid levels were lower in incident CKD cases than in persons who remained CKD-free in our study.

We note several limitations of this study. First, serum creatinine measurements provide an imperfect assessment of
GFR, and we acknowledge that more precise measures, such as
inulin clearance, would improve renal phenotyping in the FHS.
Such measures, however, are impractical in the context of a
community-based study. Of note, our study of renal arterio-
venous plasma sampling shows that select CKD predictors are
markers of renal metabolism and secretion, not simply mark-
ers of baseline differences in glomerular filtration. Second,
although we set a conservative significance threshold for me-
tabolite discovery in FHS, we acknowledge the possibility of
false discovery. Furthermore, because the FHS included
middle-aged to older individuals of predominantly European
ancestry, additional cohorts will be required to both validate our
findings and extend their generalizability to younger individuals
or other racial/ethnic groups. Third, given the clinical indica-
tions for right and left heart catheterization, our physiologic
study included several individuals with abnormal kidney
function. Ideally, further study of healthy individuals, along
with more systematic surveys of urinary fractional excretion,
will provide a more thorough picture of normal renal metabolite
handling. Nevertheless, we believe that the complete epide-
miologic and physiologic data sets we present in this study
(Supplementary Tables 1 and 2) will serve as a valuable resource
for understanding the interaction between kidney function and
the metabolome in humans.

Increasing interest has been directed toward the application of
metabolite profiling to plasma from individuals with existing
CKD or ESRD.2–4,30–32 Here, we profile plasma obtained from
individuals up to 8 years before disease onset, on the principle
that non-GFR markers of renal metabolite handling may be
sensitive indicators of incipient kidney dysfunction. Although
metabolite profiling alone may not be sufficiently specific to
identify individuals who go on to develop CKD, it may con-
tribute to clinical models of CKD prediction that contain
other variables. Thus, future efforts will be directed at mea-
suring specific metabolites in more diverse cohorts, across
distinct CKD causes, and adjudicating to what extent these
biomarkers add to established clinical predictors. In parallel,
efforts to understand how nonrenal determinants (e.g., diet
and de novo synthesis) modulate select metabolite levels will be
required to further clarify their relation to CKD risk.

CONCISE METHODS

FHS
The Framingham Offspring Study was initiated in 1971, when 5124
individuals were enrolled into a longitudinal cohort study; 3799 of
these individuals attended the fifth examination of this cohort, which
took place between 1991 and 1995.33 The fifth examination is design-
nated as the baseline examination for the current study. At this and
subsequent quadrennial visits, participants underwent a physician-
administered physical examination, medical history, and routine lab-
oratory tests. The presence of CKD was ascertained at the fifth and
seventh examinations. Of the 3799 attendees at the baseline exami-
nation, we have thus far applied all three components of our
metabolite profiling platform to baseline plasma from 2069 individ-
uals; 378 of these participants were chosen in the context of a nested
case-control study of incident type 2 diabetes,5 and the subsequent
1691 were chosen randomly. Individuals were excluded from the
current study if they had CKD at the baseline exam (n=89), had un-
known CKD status at baseline (n=240), or had unknown CKD status
at the seventh examination (n=306). Therefore, of the 2069 attendees
of the baseline examination with available metabolite data, 1434 in-
dividuals were eligible for the current study. By the seventh exami-
nation (up to 8 years after the baseline examination), 123 individuals
had developed new-onset CKD and were designated as cases, and
1311 individuals remained without CKD.

CKD Assessment
We used eGFR to estimate kidney function, using the abbreviated
Modification of Diet in Renal Disease study equation. CKD was
defined as an eGFR < 60 ml/min per 1.73 m² (i.e., CKD stage 3 or
worse).11,12 The modified Jaffe method was used to measure serum
creatinine, and creatinine levels were calibrated using a two-step pro-
cess as described previously.34 Proteinuria was defined as a value of
trace or higher on a urine dipstick assay (Ames Labstix, Elkhart, IN).

Arterial and Renal Venous Plasma Sampling
We recruited patients referred to the Massachusetts General Hospital
Cardiac Catheterization Laboratory for right and left heart catheter-
ization (n=9). In addition to the steps required in each individual’s
routine clinical care, the protocol included introduction of a Judkins
catheter into the ostium of a renal vein, with plasma sampling from
this renal venous catheter and from a catheter positioned in the ab-
dominal aorta at the level of the renal arteries before coronary artery
catheterization (and administration of iodinated contrast medium).
All participants were fasting at the time of their procedure. First morning
voided urine was obtained from the final four study participants.

The study protocols were approved by the institutional review
boards of Boston University Medical Center and Massachusetts
General Hospital. All participants provided written informed consent.

Metabolite Profiling
We applied three distinct LC-MS–based methods to distinct plasma
drugs for each experimental sample. Amino acids, amino acid de-
rivatives, urea cycle intermediates, nucleotides, and other positively
charged polar metabolites were profiled as previously described using
10 μl of plasma.5 Lipids, including lysophosphatidylcholines, lysop-
osphatidylethanolamines, phosphatidylcholines, sphingomyelins,
cholesterol esters, diacylglycerols, and triacylglycerols, were profiled as
previously described using 10 μl of plasma.35 For organic acids, sug-
ars, bile acids, and other negatively charged polar metabolites, 30 μl
of plasma were used and MS data were acquired using a 5500 QTRAP
triple quadrupole mass spectrometer (AB SCIEX, Foster City, CA) using
electrospray ionization and multiple reaction monitoring in the negative
ion mode. Detailed methods are provided in the Supplemental Methods.

Statistical Analyses
Metabolite levels were log transformed because the raw data did not
exhibit normal distributions. For the study of individuals undergoing
invasive catheterization, we compared arterial and renal venous metabolite levels before iodinated contrast using two-tailed paired t tests. In FHS, we examined the association between the plasma level of each metabolite and incident CKD in FHS. Baseline metabolite levels were compared in persons who developed CKD versus those who did not using two-tailed t tests for the 217 measured metabolites. To account for multiple testing, we used a Bonferroni-corrected P value threshold of 0.00023 (0.05/217).

For nine metabolites meeting this P value threshold (kynurenic acid, kynurenine, citrulline, choline, xanthosine, β-aminoisobutyric acid, aconitate, isocitrate, and 5-hydroxyindoleacetic acid), we performed logistic regression analyses to estimate the OR of CKD at different metabolite values. Metabolites were analyzed as continuous variables (log transformed and scaled to SD of 1), and regressions were adjusted for eGFR, age, sex, hypertension, diabetes, and proteinuria at baseline. To assess the joint predictive ability of these metabolites, we first ran a stepwise logistic model including all nine metabolites. Five metabolites—kynurenic acid, xanthosine, 5-hydroxyindoleacetic acid, kynurenine, and citrulline—remained significant in the multivariable model (P threshold=0.05). We constructed a multimarker score based on the regression coefficients of these five metabolites, and then assessed whether a model including all clinical risk factors plus the multimarker panel improves CKD prediction compared with the model including clinical risk factors only. We used c-statistics to compare model discrimination, “category-free” net reclassification improvement to assess the ability of the model to correctly reclassify risk groups, and integrated discrimination improvement to examine the ability of the model to increase average sensitivity without reducing average specificity. We repeated these analyses for a more parsimonious multimarker panel composed of kynurenic acid, xanthosine, and 5-hydroxyindoleacetic acid because this subset of metabolites had the strongest P values in the stepwise logistic model.

All analyses were performed using SAS software, version 9.1.3 (SAS Institute, Cary, NC).

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REFERENCES

None.

DISCLOSURES

None.

REFERENCES


