

SUPPLEMENTARY APPENDIX**Supplementary Table S1:**

Serum sample	biopsy	
43 aPLA2R1 +	11 PLA2R1 + 2 PLA2R1 - 10 not available	43 positive
39 aPLA2R1 -	11 PLA2R1 + 10 PLA2R1 - 18 not available	11 positive
13 not available	6 PLA2R1 + 2 PLA2R1 - 5 not available	6 positive
		60 positives

aPLA2R1 = anti-PLA2R1 antibodies ; PLA2R1 = PLA2R1 in depositions positive = patients are considered to have anti-PLA2R1 related disease if either antibodies were present in serum or PLA2R1 antigen was detectable in renal biopsy (see Methods).

Supplementary Table S2:

Exon	Forward primer	Reverse primer	annealing temperature
1	<u>AGTGGGCTCCAGGGCTC</u>	<u>AGGACTGCTGACACACGAAC</u>	64
2	<u>AGATCTGTGGGAAATGGCTG</u>	<u>GGTCCAGGCTTCGTACTTC</u>	57
3	<u>TGAGTATCTGCCCAAGAGG</u>	<u>CTTCAATATCAATCCCAGTGC</u>	57
4	<u>GGCCCTGAAGTTTAGAGATTG</u>	<u>TTTGGGTGAGAGTTTGGG</u>	57
5	<u>TGAGCCTGCTCTGTTCTCA</u>	<u>TGGCTTACACCAAATCCTC</u>	57
6	<u>CATGCCCTGGACTGTACTGC</u>	<u>CACAGTCCCCTTGACATAACAGA</u>	64
7	<u>CACTAGCTCATCCGGAAAATG</u>	<u>GGGGTTCCCTTATTATGTGG</u>	57
8	<u>TCAAACCATAGCAGGGAAAGG</u>	<u>CACAGGTGGCCTTCAGTACA</u>	64
9	<u>GCACATGGCAGCTATAAGA</u>	<u>GACAGGGCATCACACATCAC</u>	57
10	<u>AGAAGTGGGTAGTTAATGGC</u>	<u>GGGTTTCACTTCCATATAAGC</u>	57
11	<u>TACTCTGGGCCATTCTGTG</u>	<u>CCCTGTTAACCTGTGTTGATCC</u>	57
12	<u>TTGCCATCTGAGCACAAAG</u>	<u>TCTGTCAGGCACTGTTCTGG</u>	57
13	<u>TTCCCTCCACTAGATCAAGCTC</u>	<u>GGGAATTAAACAACACTTGGG</u>	57
14	<u>GCATCAGCTTCTGC</u>	<u>GTGCAGTTTGTATGGAG</u>	64
15	<u>AGGGATGCCCTTTGATCT</u>	<u>CTGTCCCAGGTGTTCCACT</u>	57
16	<u>GCCTCTGGAGCTCCCTCT</u>	<u>AAAAGGTGGGGATAAGGTACA</u>	64
17	<u>TGTACCTATCCCGACCTTT</u>	<u>AAACAATCTGACGTGCTCA</u>	64
18	<u>GGTTTCTCAAGATGCTAGGC</u>	<u>CACTCATGTTGGTTAGAGAAG</u>	57
19	<u>TGGAGACTGGACATGCAGAA</u>	<u>CCCATTCTGGTGCTGTCTT</u>	64
20	<u>CCTGCCAGTTAGCTGTTG</u>	<u>TAGAAACGAGGGGCAGTGTG</u>	64
21	<u>CTGTAACACCTTGGAGGAGG</u>	<u>GGTCAAACTGCAAACCTCGT</u>	64
22	<u>AAATAGGATGCAGTGATTACCC</u>	<u>GGCACAGAGAGGAATGTGTT</u>	64
23	<u>TGCGAATAAAGGAATAATGGTG</u>	<u>CATGCCACATTAGCCACAG</u>	57
24	<u>CAACATGGATCATAGTGACAAGG</u>	<u>AGGCTGTTCATAGTTGGTC</u>	64
25	<u>TCGATGTGGATTGTAGCA</u>	<u>ATCACCAACCAGACAGAGG</u>	57
26	<u>CATGAAACCCAATCTCAGAG</u>	<u>GGTTTCAAATTCTACACTGGAC</u>	64
27	<u>GCCTTAACTCTGACGTCAA</u>	<u>AGCCAAGCATCTCTGCTA</u>	64
28	<u>AGGGGCTGTTTATGGTT</u>	<u>CCAGAATGAAGGTTGCTGGT</u>	64
29 and 30	<u>GTCATGGGCCATAGTTGCT</u>	<u>TGGCATCTGTGCTGTAAAGG</u>	64

All primer sequences given are from 5' to 3'. Underlined primers were used for sequencing. For exon 9 the following primer was used for sequencing (TTCAAATTGATGGTGGAAATTGA). PCR was performed using the AmpliTaq Gold 360 PCR mix (Applied Biosystems); for the amplification of exon 4, 5 and 7 AmpliTaq Gold (Applied Biosystems) was used.

Supplementary Table S3: population of European Ancestry , 1000 Genomes Project, release 10 March 2012

Population	Number
Utah residents (CEPH) with Northern & Western European ancestry (CEU)	85
Toscani in Italy (TSI)	98
British from England and Scotland (GBR)	89
Finnish from Finland (FIN)	93
Iberian population in Spain (IBS)	14
Total European ancestry (CEU)	379