

Supplemental Information

Single Cell Profiling of Acute Kidney Injury Reveals Novel Transcriptional Signatures, Pro-Fibrotic Phenotype and Epithelial-to-Stromal Crosstalk

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Supplemental Figure 1. UIR induced proximal tubule dedifferentiation, tubular damage and gene expression changes. Related to Figure 1.

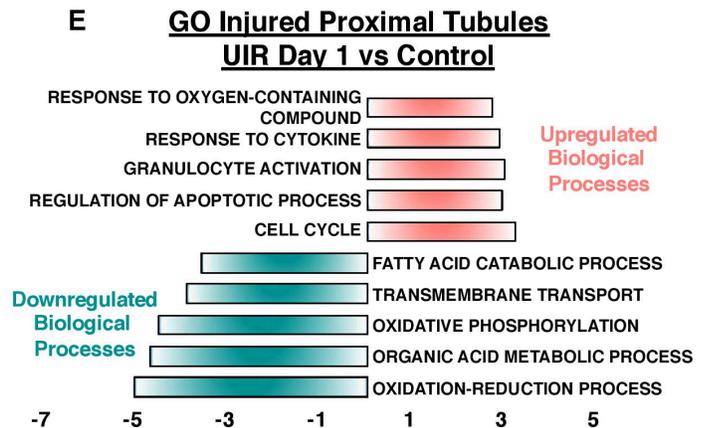
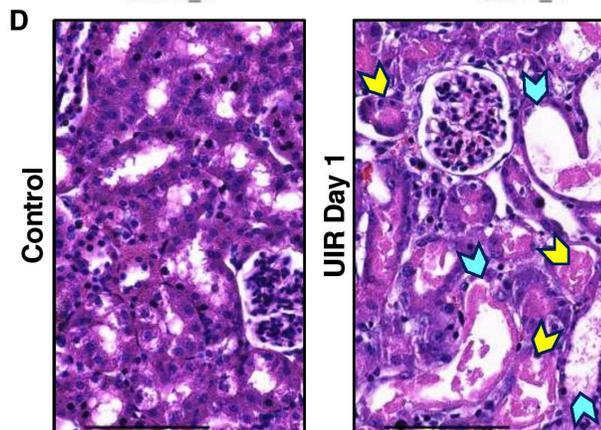
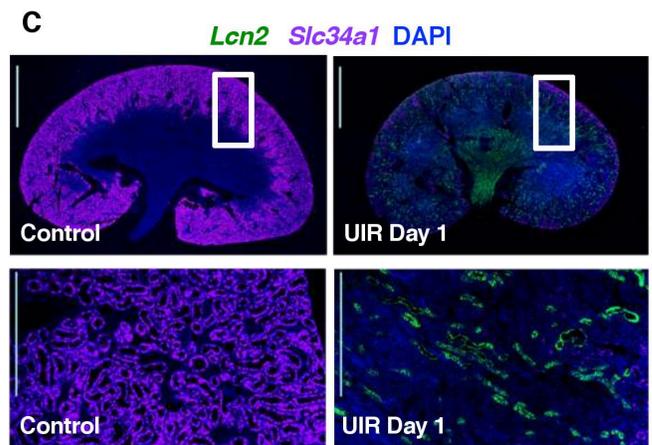
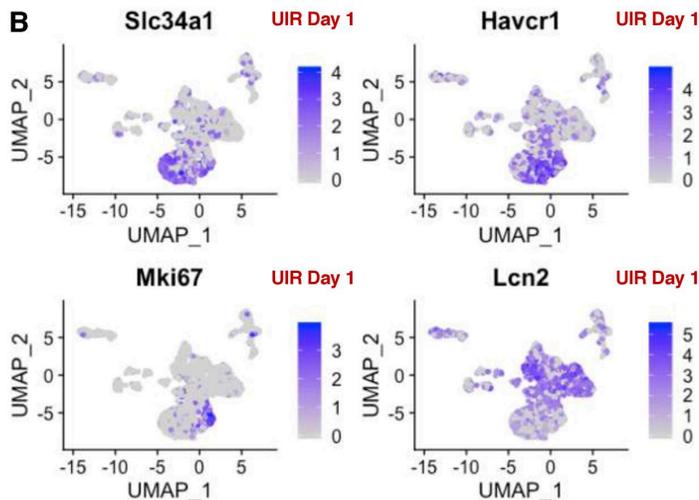
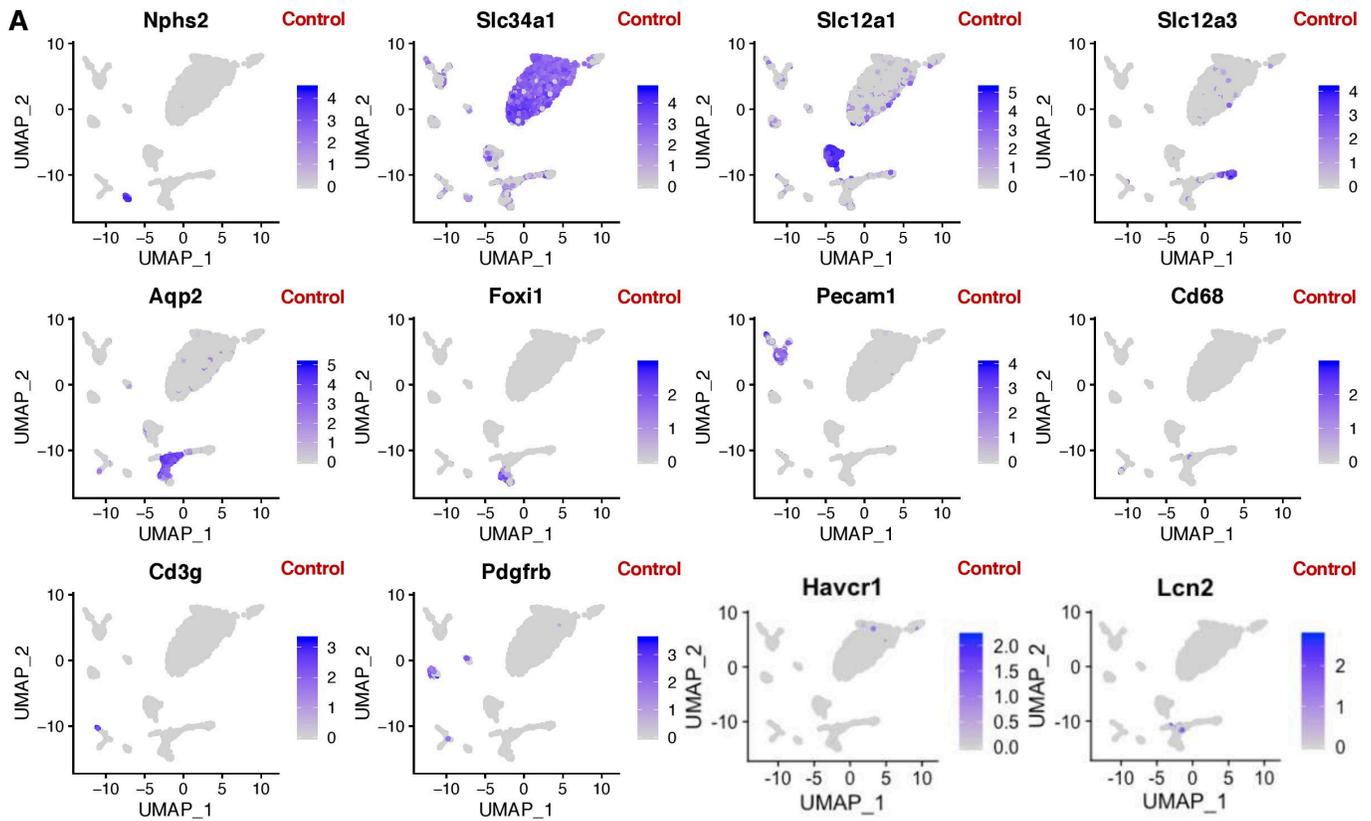
(A) Feature Plots for renal cell populations identified in the Control kidney by the scRNA-seq. Podocytes (*Nphs2*), proximal tubules (*Slc34a1*), loop of Henle (*Slc12a1*), distal tubules (*Slc12a3*), collecting duct principal (*Aqp2*) and intercalated (*Foxi1*) cells, endothelial (*Pecam1*), macrophages (*Cd68*), T cells (*Cd3g*), stromal/pericyte (*Pdgfrb*). Both injury markers *Havcr1* and *Lcn2* are nearly absent.

(B) Feature Plots identify UIR Day 1 cell populations, including sub-clusters of injured (high *Kim1*, *a.k.a.* *Havcr1*, and low *Slc34a1*) and cycling (*Mki67* positive) proximal tubules, the injured *Lcn2*-positive distal tubule, loop of Henle and collecting duct. The “Mixed Identity Cells” are positioned between the proximal tubules, distal tubules, loop of Henle and collecting duct.

(C) RNAscope showing *Lcn2* elevation in the distal nephron tubule and collecting duct of UIR Day 1. *Lcn2* (green), *Slc34a1* (purple), 4x (2500 μ m scale) and 10x (500 μ m scale).

(D) H&E staining reveals AKI induced tubular dilation (blue pointers) and cast formation (yellow pointers) not detectable in the Control. 40x, 100 μ m scale.

(E) GO Biological Process of “Injured Prox” UIR Day 1 vs Control, $-\log_2(\text{pValue})$.



Supplemental Figure 2. AKI results in formation of the Mixed Identity Cells in the adult kidney.

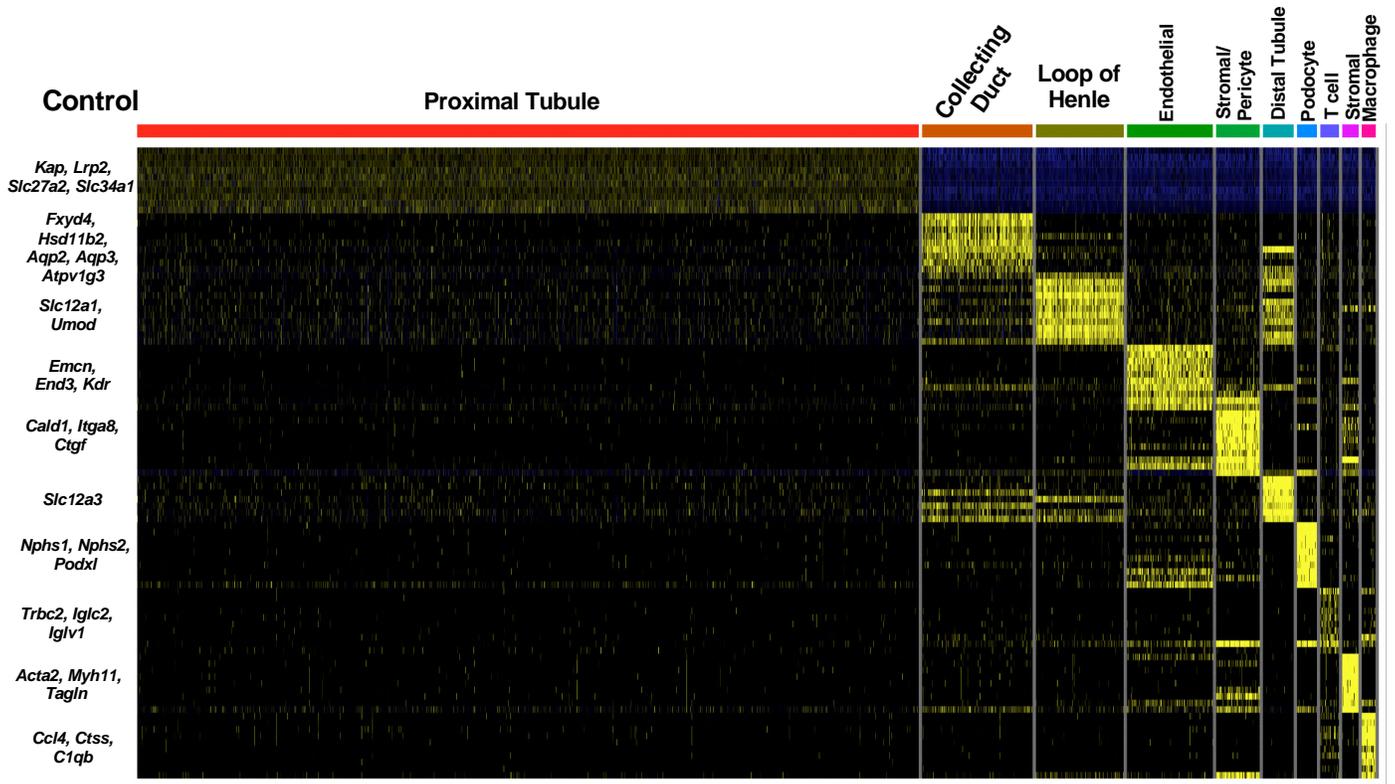
Related to Figures 1 and 2.

(A) Heatmap shows the relative marker gene expression in the Control renal cell populations. The “Mixed Identity Cells” with ectopic expression of multiple compartments markers are not identified.

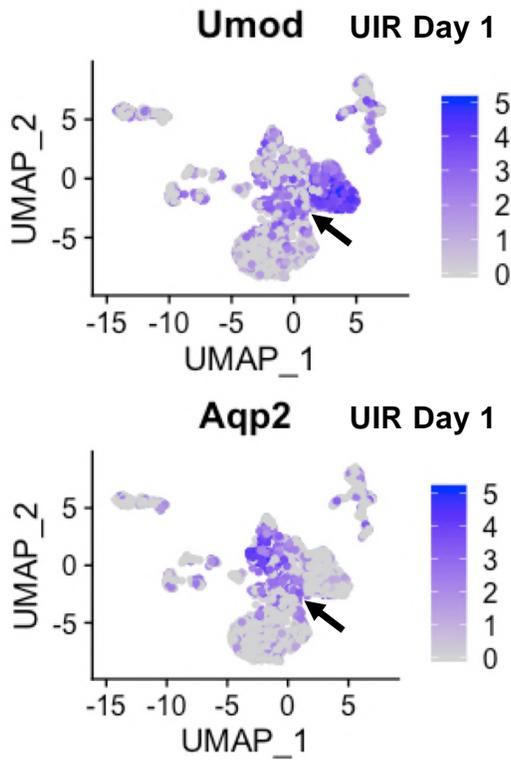
(B) Feature plots show the expression of the loop of Henle marker *Umod* and the collecting duct marker *Aqp2* in the UIR Day 1. The intermediately located “Mixed Identity Cells” pointed with the arrow show overlapping expression of both markers.

(C) Combined feature plot shows the overlapping expression of the proximal and distal nephron tubule injury markers *Kim1* and *Lcn2* in the UIR Day 1 “Mixed Identity Cells” (shown with the arrow).

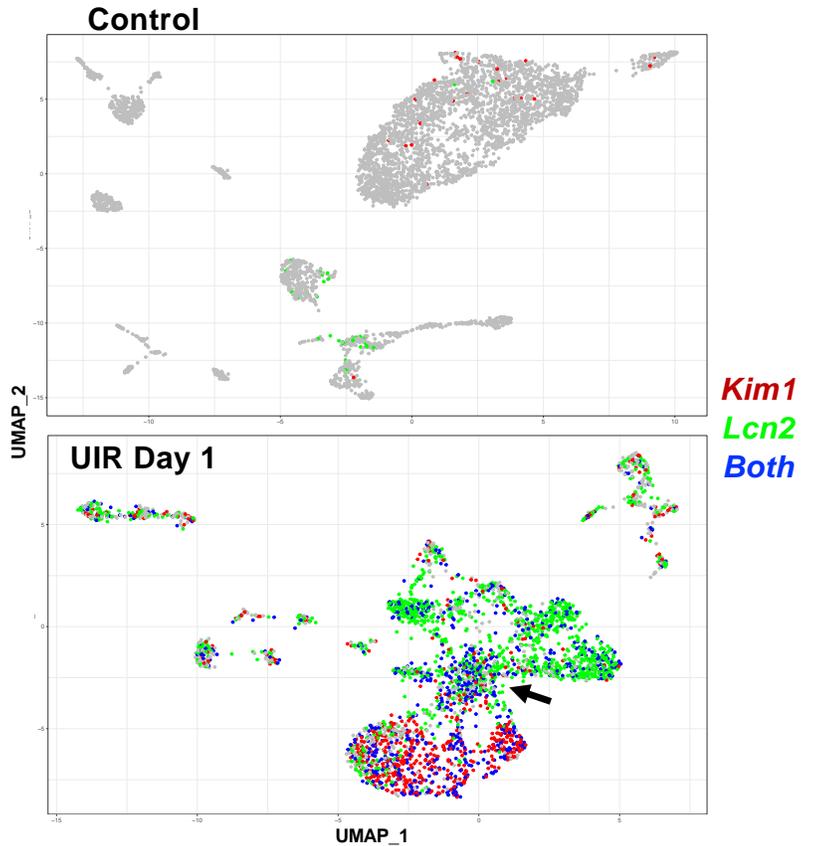
A



B



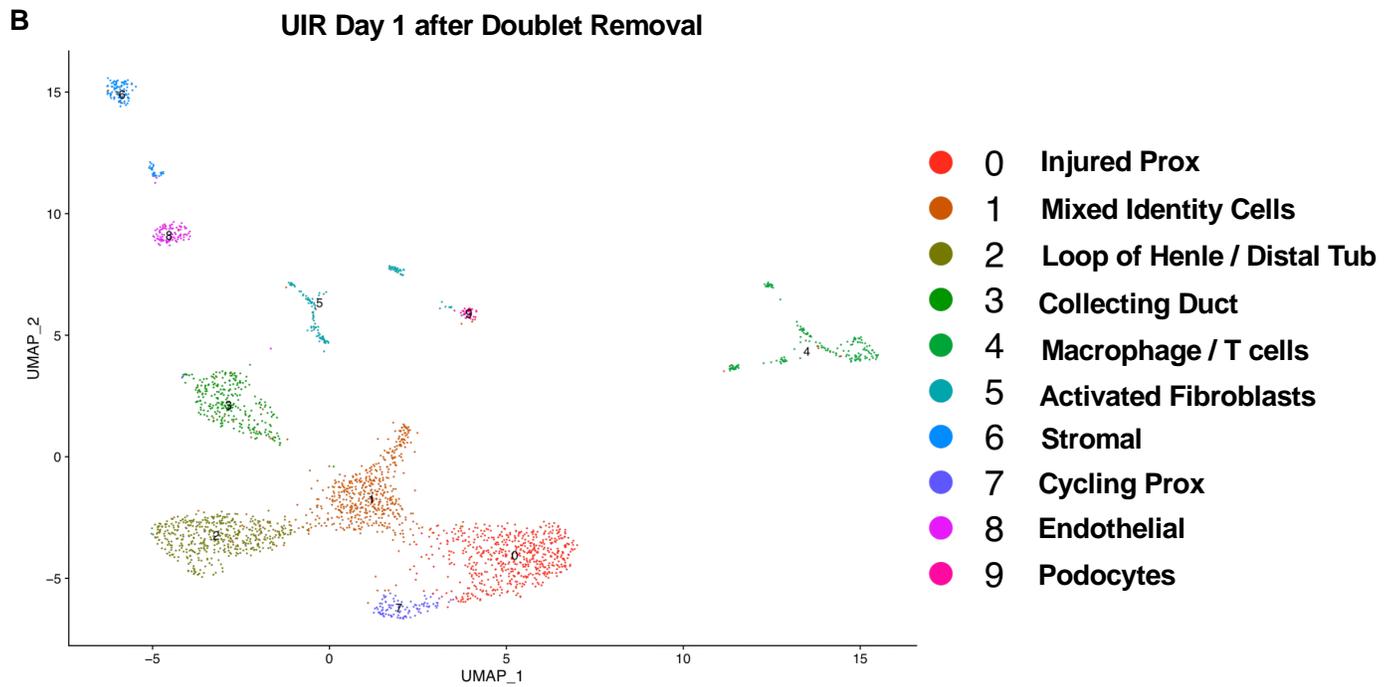
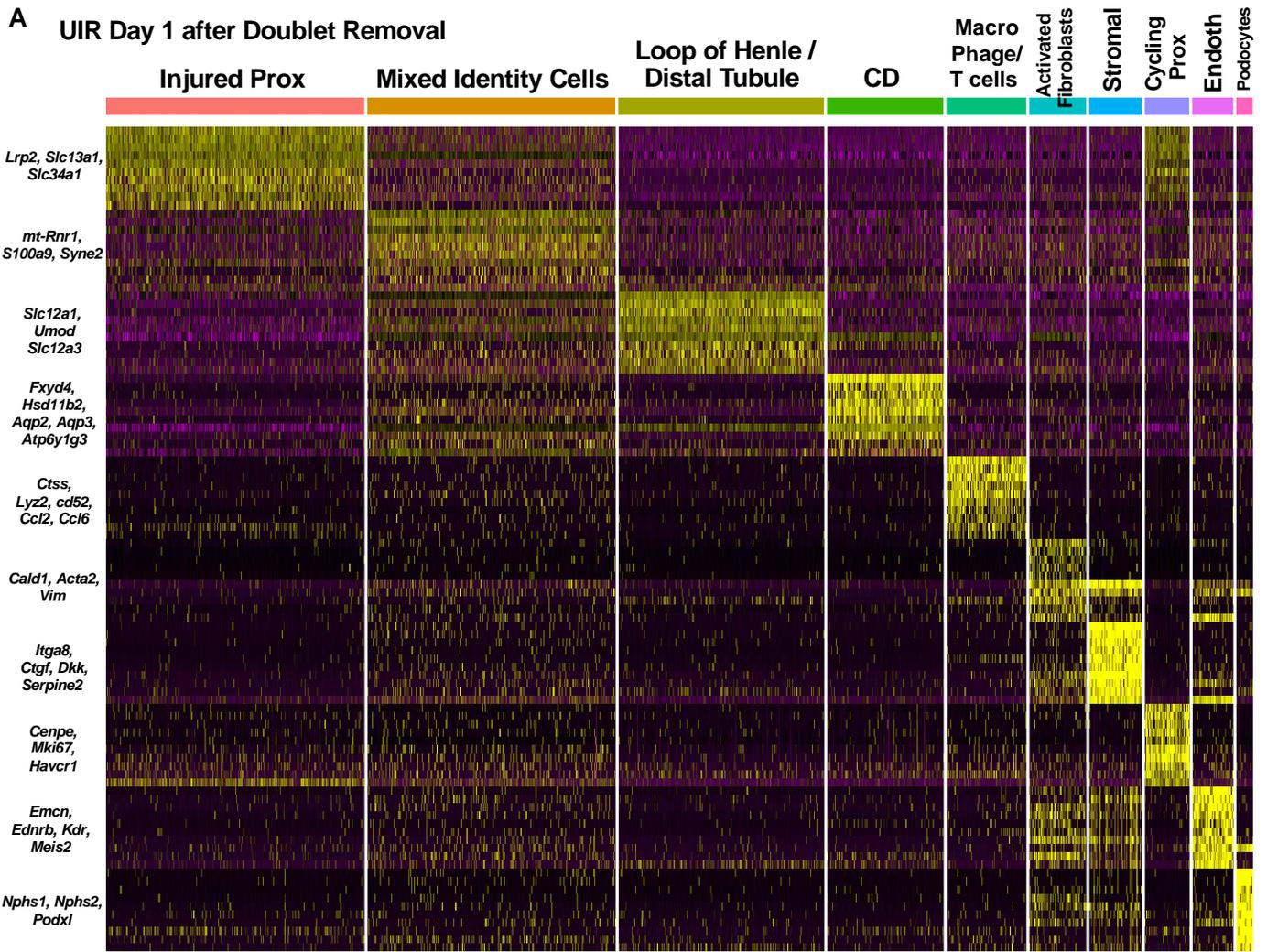
C



Supplemental Figure 3. Doublet removal does not affect the Mixed Identity Cells presence. Related to Figures 1 and 2.

(A) Heatmap shows the relative marker gene expression in UIR Day 1 renal cell types after doublet removal using Doublet Decon. The Mixed Identity cluster remains present and shows stochastic expression of markers of many different cell types. Please note that the heatmap shows genes elevated in renal cell populations relative to each other, based on the z-score. The yellow is above the mean, black is the mean, and purple/blue is below the mean, which represents the lowest expression levels. While other injured renal cell types exhibit some minor degree of “inappropriate” gene expression, the Mixed Identity Cells cluster is defined by remarkably elevated levels of many renal cell types markers. Endoth, Endothelial; Prox, Proximal Tubules; Distal Tub, Distal Tubule.

(B) UMAP shows UIR Day 1 renal cell populations after the doublet removal. The figure shows that presence of Mixed Identity cells (Cluster 1) is not affected by the doublet removal.



Supplemental Figure 4. UIR induces quantitative changes in mixed identity and proximal tubular dedifferentiation. Related to Figures 1 and 2.

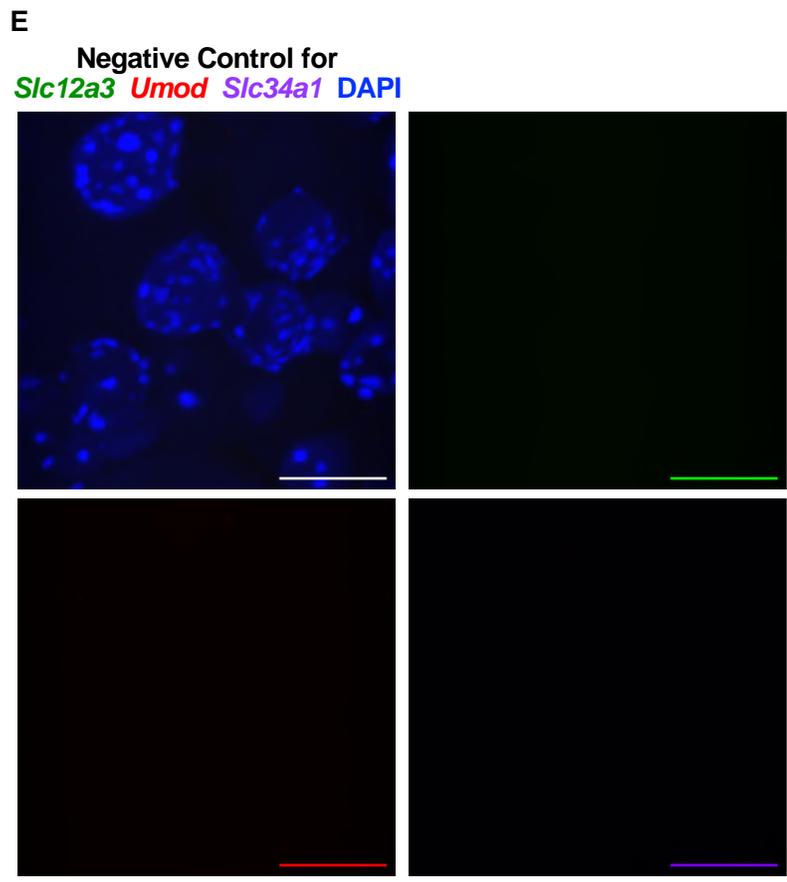
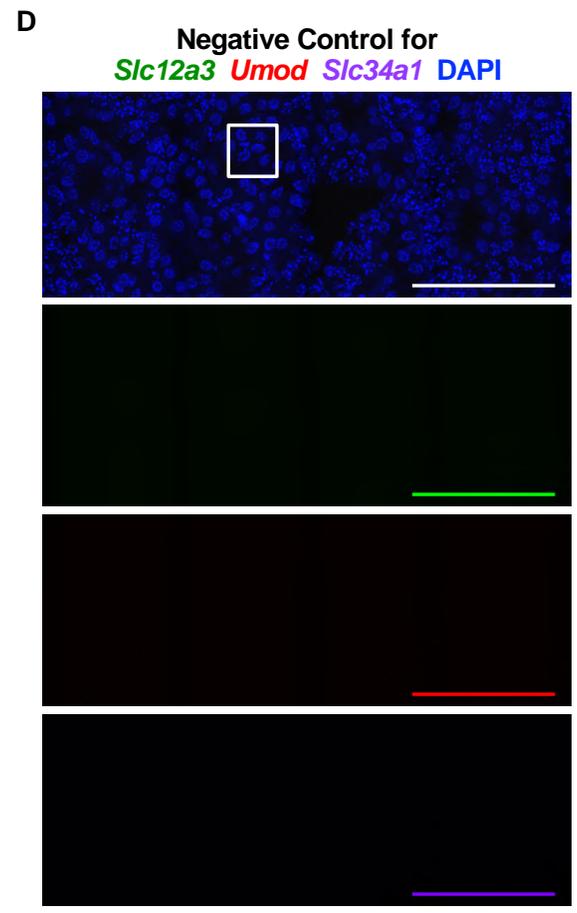
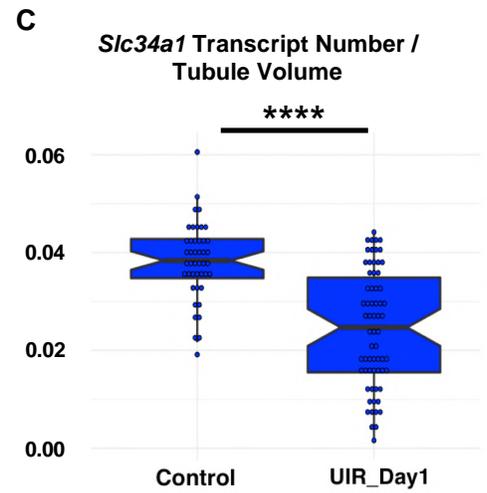
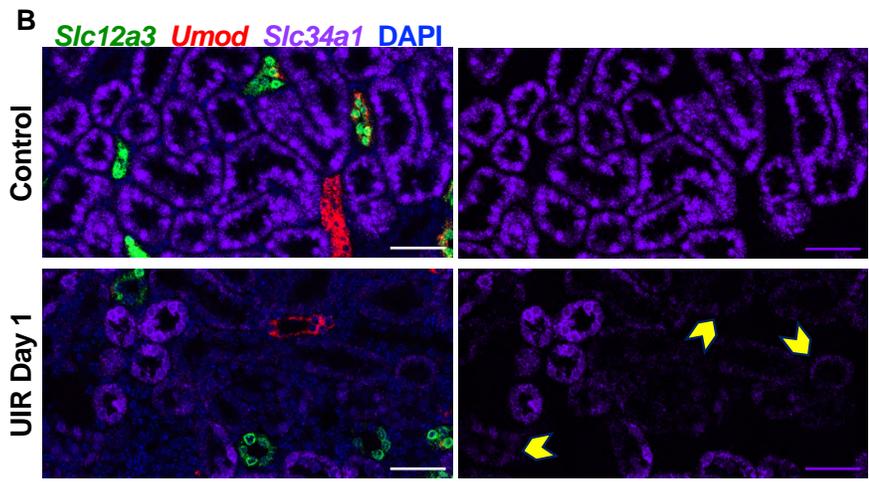
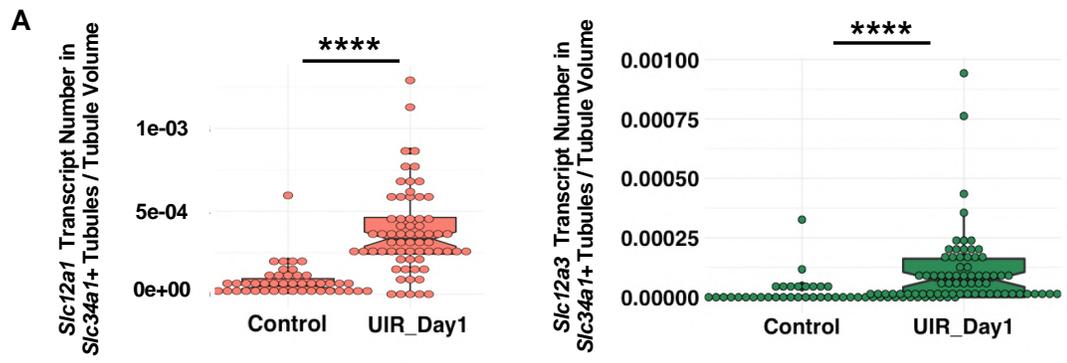
(A) IMARIS quantification of *Slc12a1* and *Slc12a3* transcripts in the *Slc34a1*-positive UIR Day 1 vs Control tubules, 12 Z-stacks (50-70 tubules) per group, Student's *t* test, **** $p < 0.0001$. Data is presented as the transcript number normalized to the tubule volume.

(B) RNAscope with *Slc34a1* (purple), *Umod* (red) and *Slc12a3* (green) probes, DAPI (blue), Control and UIR Day 1. 60x 2x Nyquist zoom, 0.10 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from $\sim 6\mu\text{m}$ Z-stack, 12 stitched images, scale 50 μm . Tubules with reduced *Slc34a1* expression are shown with the yellow pointers.

(C) IMARIS quantification of *Slc34a1* in the UIR Day 1 vs Control *Slc34a1*, $n=12$ Z-stacks (50-70 tubules) per group, Student's *t* test, **** $p < 0.0001$. Data is shown as *Slc34a1* transcript number normalized to the tubule volume.

(D) Negative control for RNAscope with *Slc34a1* (purple), *Umod* (red) and *Slc12a3* (green) probes, DAPI (blue). 60x 2x Nyquist zoom, 0.10 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from $\sim 6\mu\text{m}$ Z-stack, 12 stitched images, scale 50 μm . Negative control was processed in the same batch with the experimental samples. Higher magnification from panel E is highlighted with the white frame.

(E) Negative control for RNAscope with *Slc34a1* (purple), *Umod* (red) and *Slc12a3* (green) probes, DAPI (blue). 60x 6x Nyquist zoom, 0.03 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from $\sim 6\mu\text{m}$ Z-stack, scale 10 μm . Negative control was processed in the same batch with the experimental samples.



Supplemental Figure 5. AKI persists through UIR Day 4 and 7 and resolves by UIR Day 14. Related to Figure 3.

(A) H&E shows the pronounced renal tubular injury at Day 2, which starts resolving at Day 4 and 7.

UIR Day 11 and 14 exhibit normal renal histology. 40x. Tubular dilation (blue pointers), cast formation (yellow pointers).

(B) UMAP shows renal cell populations in the UIR Day 11.

(C) *Slc34a1* CISH, UIR Day 11, 4x, 40x.

(D) *Kim1* (green), *Aqp2* (red), *Slc34a1* (purple), DAPI (blue) RNAscope, UIR Day 11, 4x, 60x.

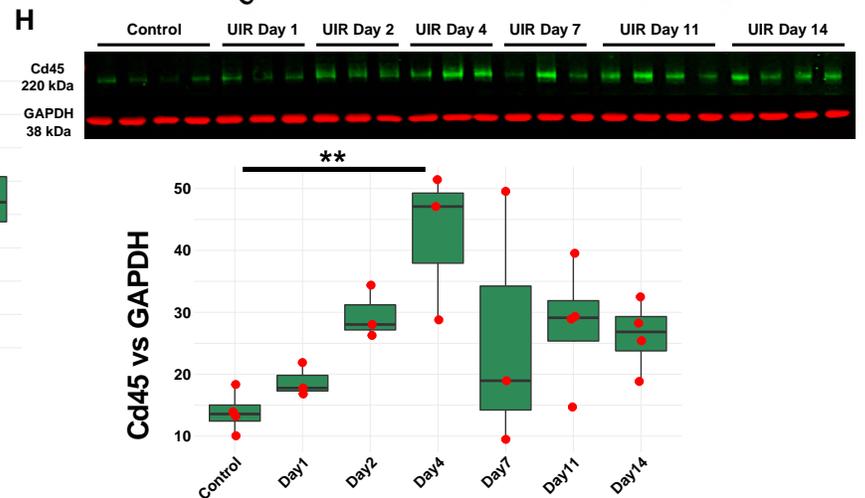
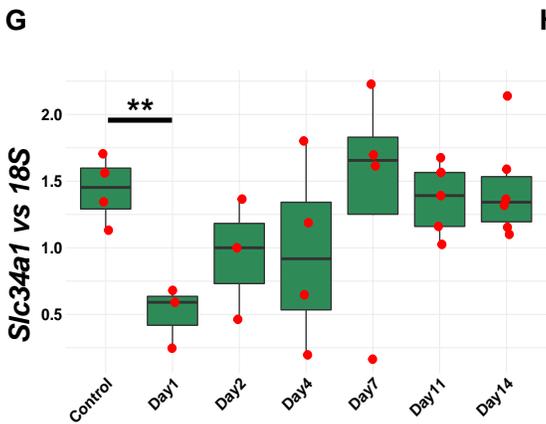
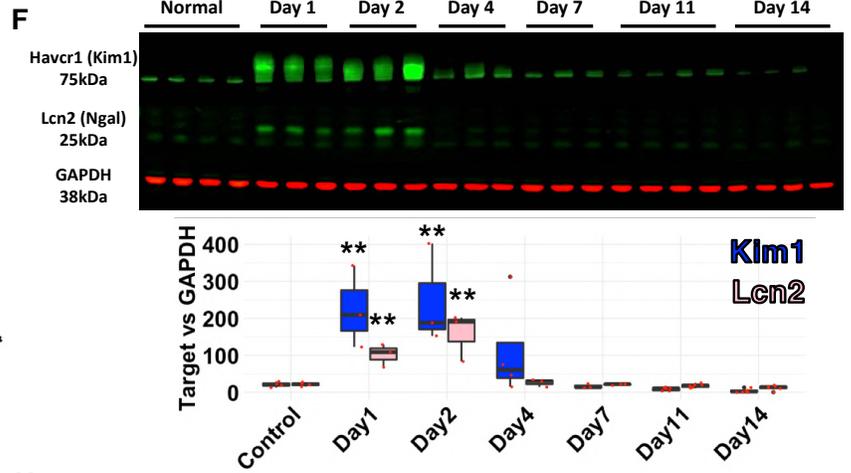
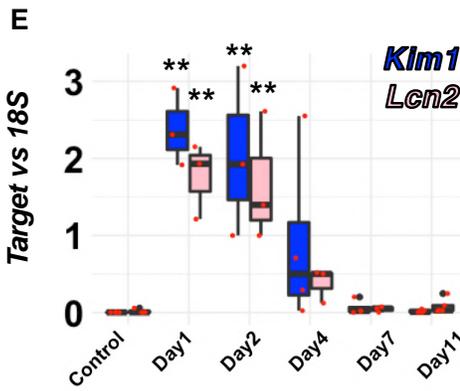
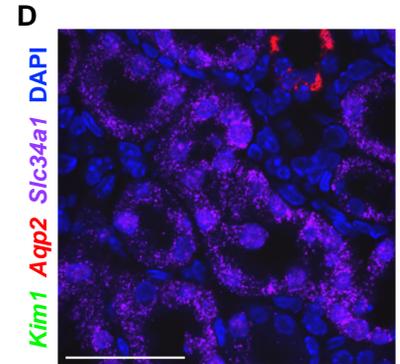
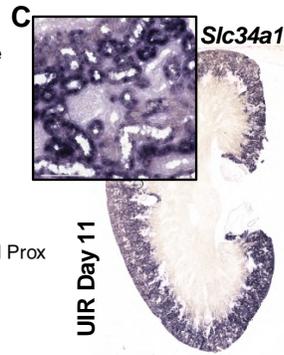
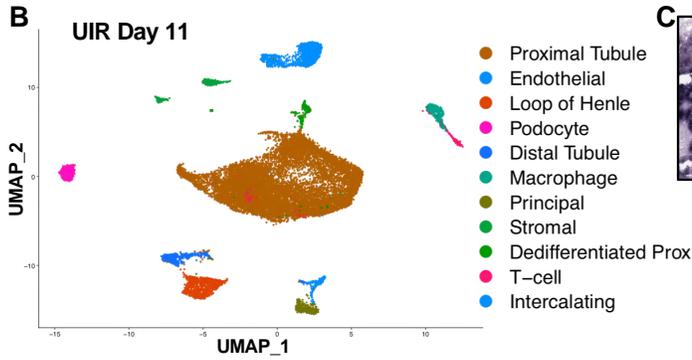
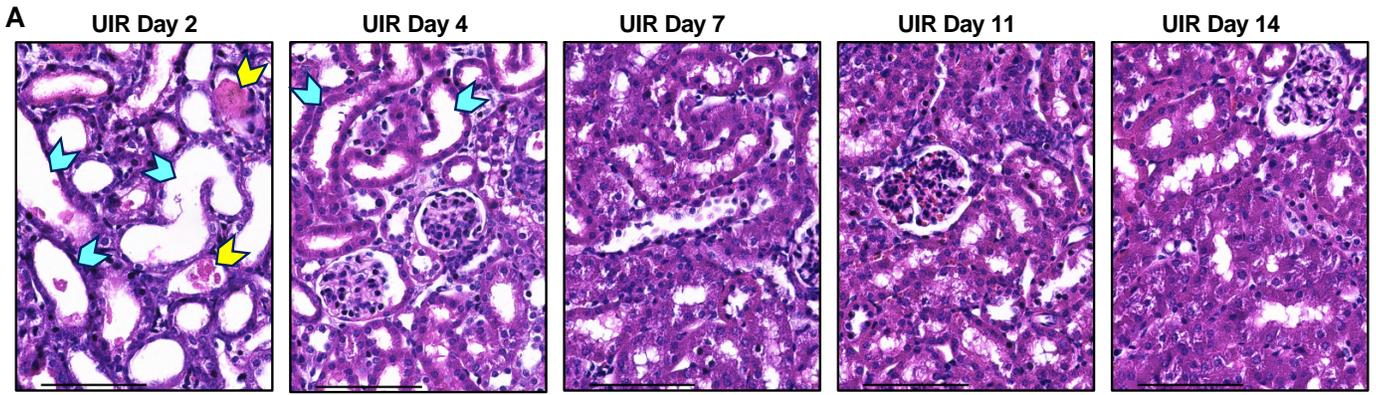
(E) qPCR shows *Kim1* and *Lcn2* expression over the AKI course, one-way ANOVA with Bonferroni and Holm, n=3-6 per group, ** pValue<0.01 compared to Control.

(F) Western blots image and quantification show *Kim1* and *Lcn2* expression over the AKI course, one-way ANOVA with Bonferroni and Holm, n=3-4 per group, ** pValue<0.01 compared to Control.

(G) qPCR shows *Slc34a1* expression over the AKI course, Student's *t* test, n=3-6 per group, ** pValue=0.0040 compared to Control.

(H) Western blot image and quantification shows *Cd45* expression over the AKI course, one-way ANOVA with Bonferroni and Holm, n=3-4 per group, ** pValue<0.01 compared to Control.

Scale 4x, 2500µm, 40x, 100µm, 60x, 25µm.

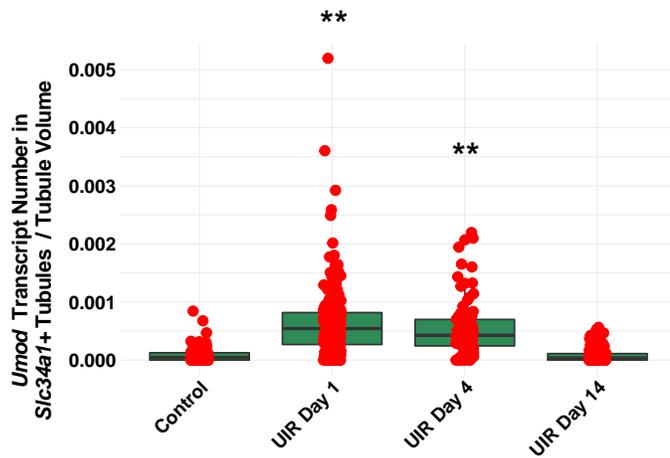


Supplemental Figure 6. AKI induced unique gene expression signatures in the Injured Proximal tubules and Mixed Identity Cells. Related to Figure 3.

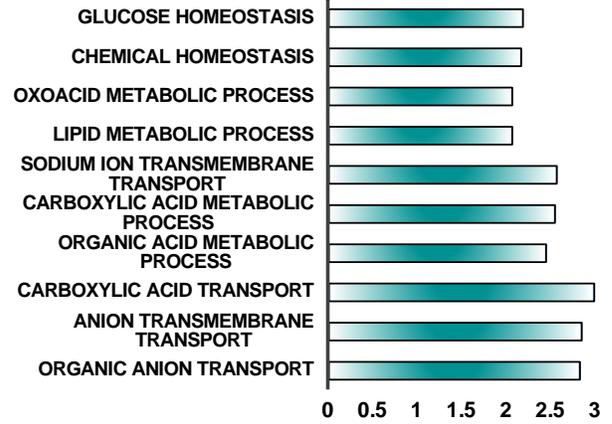
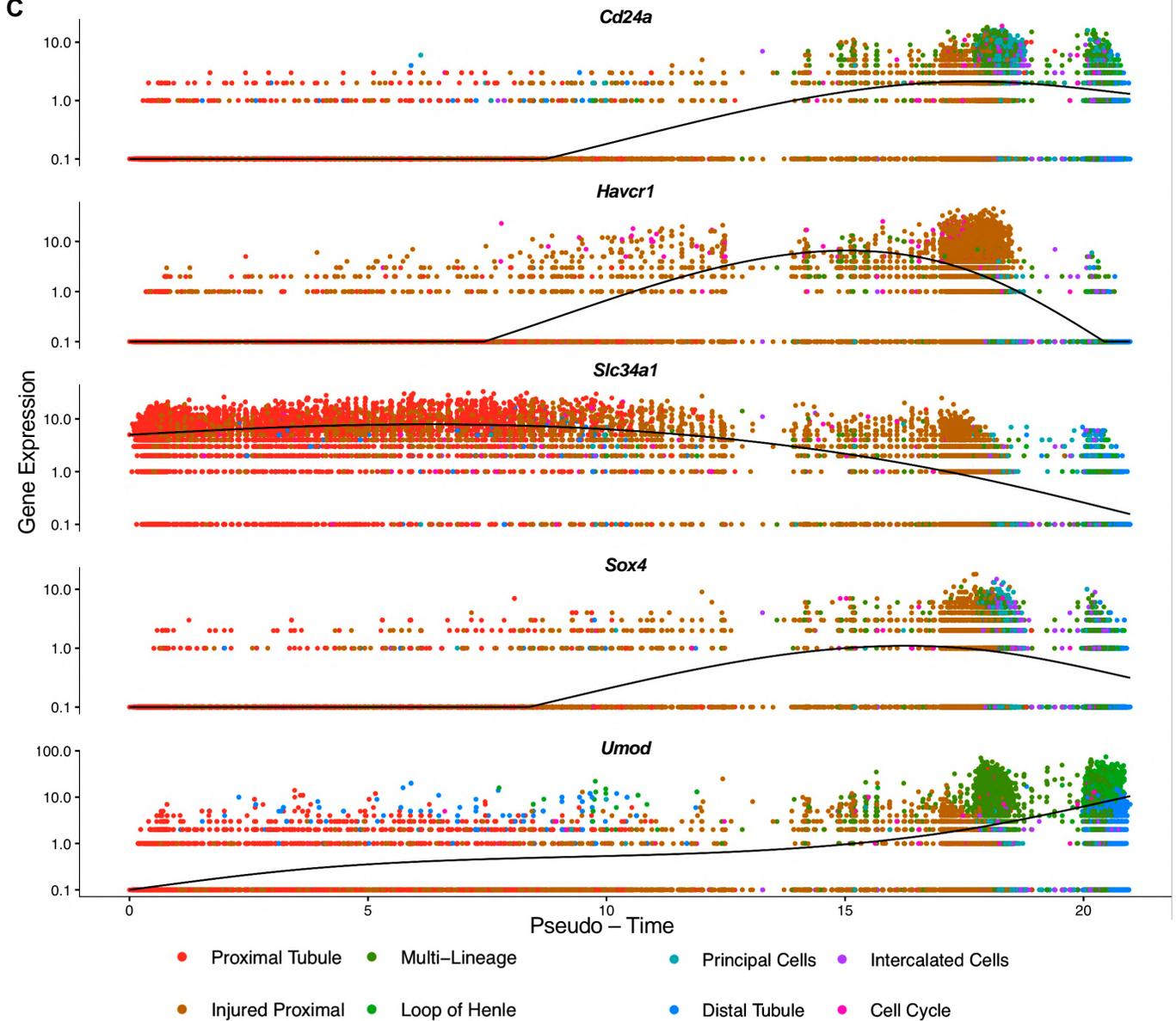
(A) IMARIS quantification of *Umod* transcripts in the UIR Day 1 (n=4 animals), 4 (n=2 animals) and 14 (n=2 animals) vs Control (n=3 animals) tubules, 12 ~6 μm Z-stack images per group, one-way ANOVA with Bonferroni and Holm, ** pValue<0.01 compared to Control. Data is presented as the transcript number normalized to the tubule volume.

(B) GO Biological Process analysis of the UIR Day 14 marker genes, $-\log(2)$ pValue.

(C) Trajectory analysis of the UIR Day 1, 4, 7, 14 and the Control demonstrates the changes of kidney injury marker (*Kim1*), proximal tubular (*Slc34a1*) and loop of Henle (*Umod*) markers and renal developmental genes (*Cd24a*, *Sox4*) in the renal cell populations over the Pseudo – Time.

A**B**

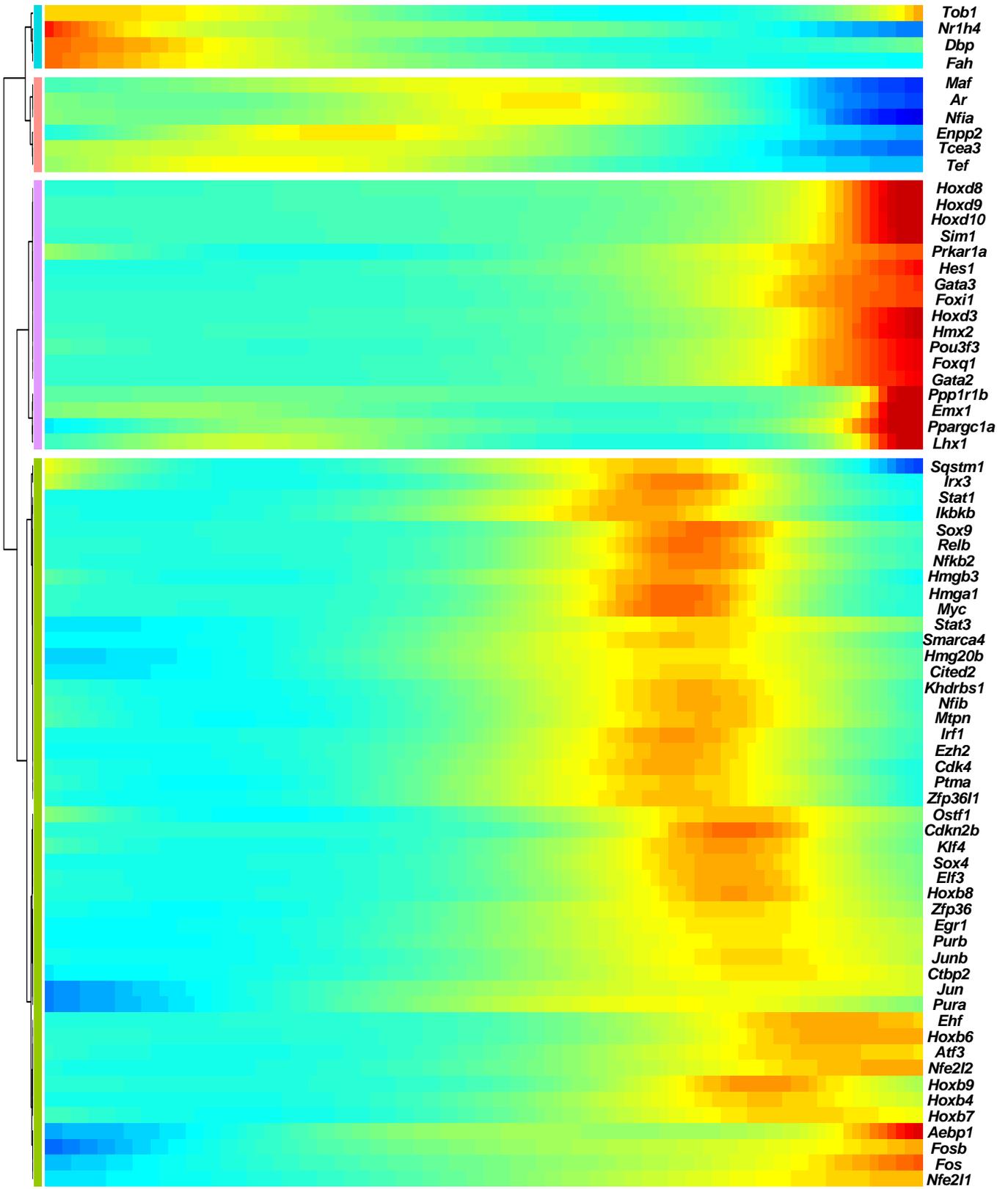
**GO Proximal Tubule Markergene
UIR Day 14**

**C**

Supplemental Figure 7. The trajectory analysis reveals changes in transcription factor expression over AKI response. Related to Figure 3.

(A) The linear heatmap shows transcription factors expression changes in the Control, UIR Day 1, 4, 7 and 14 renal tubular cell clusters (proximal tubules, injured proximal tubules, cell cycle proximal tubules, mixed identity cells, distal tubules, loop of Henle, collecting duct) over a Pseudo – Time. The cells are clustered based upon similarity of transcription factors expression. Note the significant presence of renal developmental genes, including *Sox4* and *Hox* genes.

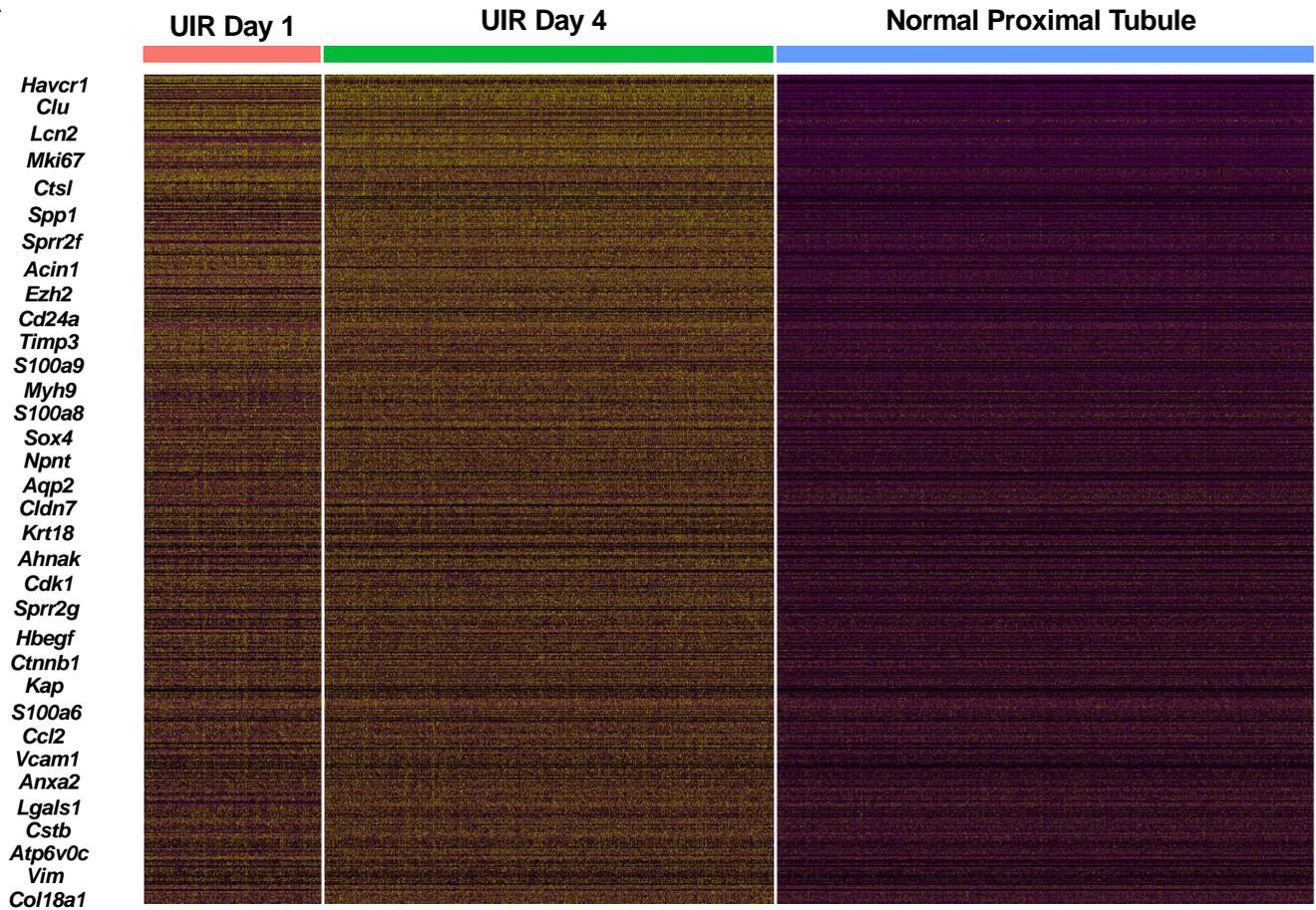
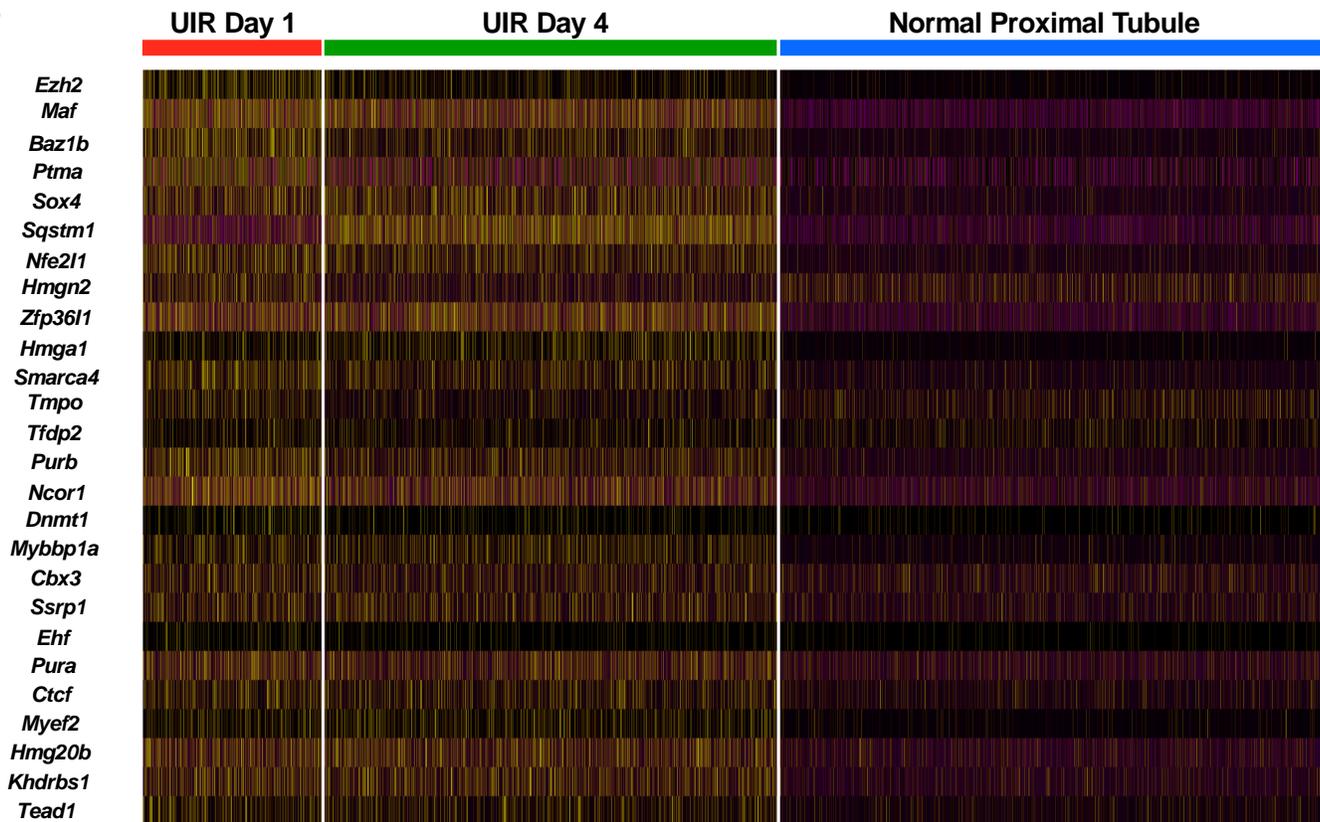
A



Supplemental Figure 8. Injury induced gene expression analysis reveals enrichment of apoptotic, pro-fibrotic and developmental factors in the UIR induced proximal tubules and mixed identity cells. Related to Figure 3.

(A) Heatmap shows the relative gene expression in the UIR Day 1 and 4 renal clusters (proximal tubules, injured proximal tubules, mixed identity cells) compared to adult normal murine proximal tubules. Please note that the heatmap shows genes elevated in renal cell populations relative to each other, based on the z-score. The yellow is above the mean, black is the mean, and purple/blue is below the mean, which represents the lowest expression levels. Highlighted genes are labeled on the left side of the heatmap; the complete list of genes elevated in the UIR Day 1 and 4 compared to the normal is shown in the Supplemental Table 4.

(B) Heatmap shows the relative transcription factor expression in the UIR Day 1 and 4 renal clusters (proximal tubules, injured proximal tubules, mixed identity cells) compared to adult normal murine proximal tubules. Individual transcription factors elevated in the UIR Day 1 and 4 clusters compared to the normal are labeled on the left side of the heatmap.

A**B**

Supplemental Figure 9. *Sox4* labels proximal tubules dedifferentiation throughout the AKI course.

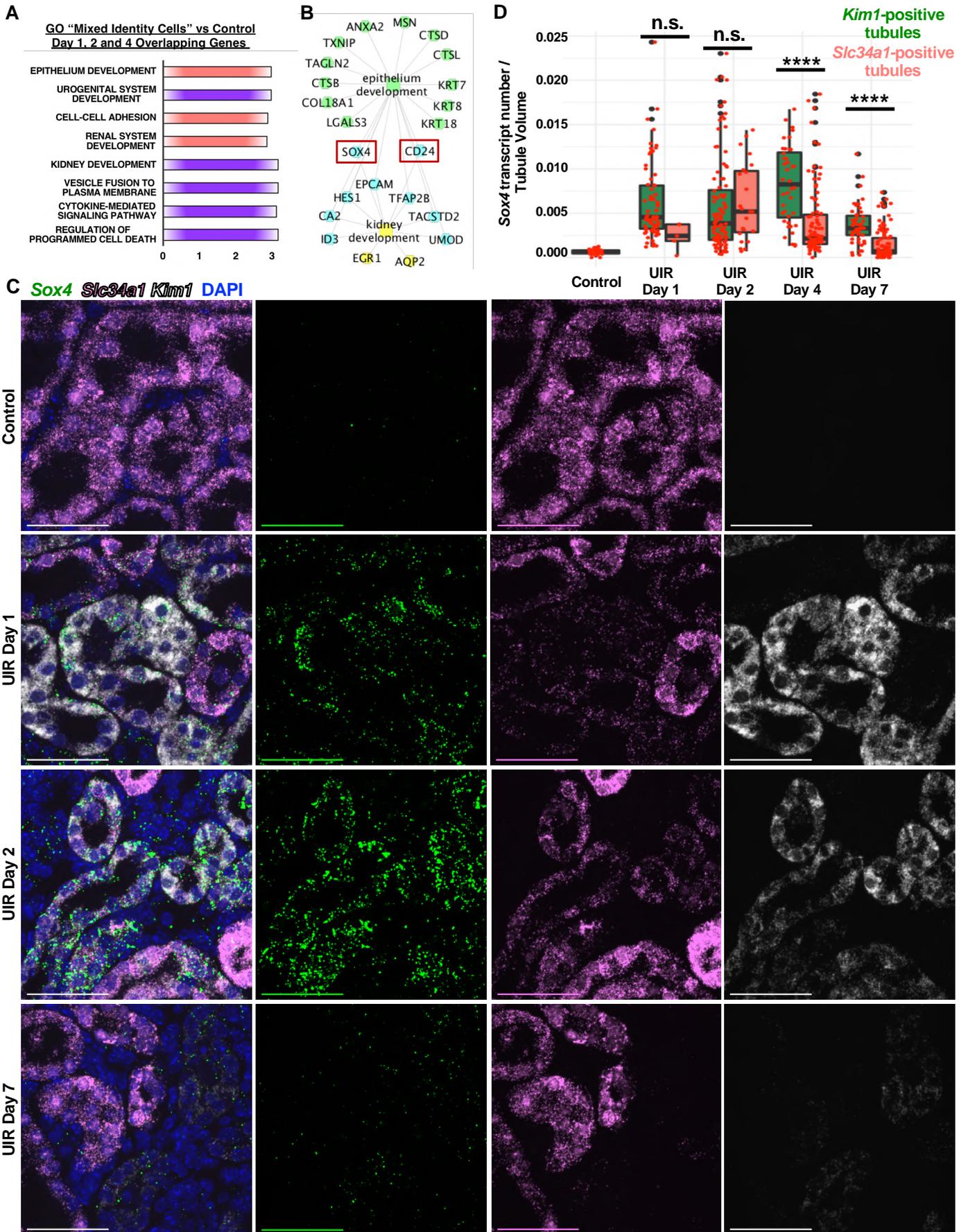
Related to Figure 4.

(A) GO Biological processes of genes overlapping in the “Mixed Identity Cells” at Day 1, 2 and 4, -log(2) pValue. Renal developmental pathways highlighted in salmon color.

(B) ToppCluster analysis of the 99 genes overlapping between the UIR Day 1, 2 and 4 in the “Mixed Identity Cells” shows the enrichment of kidney (green) and epithelium development (yellow) biological processes. Genes involved in both processes are labeled with blue polygons. *Sox4* and *Cd24a* are highlighted in red. The analysis is done with 0.05 pValue cutoff and Bonferroni correction, the graph was made using the Fruchterman-Reingold algorithm.

(C) RNAscope with *Slc34a1* (pink), *Sox4* (green) and *Kim1* (white) probes, DAPI (blue), Control and UIR Day 1, 2, 7. 60x Nyquist zoom, 0.14 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from $\sim 6 \mu\text{m}$ Z-stack, scale 25 μm .

(D) IMARIS quantification of *Sox4* in the UIR Day 1, 2, 4 and 7 vs Control *Slc34a1* vs *Kim1*-positive renal tubules, n=12 Z-stacks (50-70 tubules) per group, Student's *t* test, **** $p < 0.0001$, n.s. – not significant. Data is shown as *Sox4* transcript number normalized to the tubule volume.



Supplemental Figure 10. *Cd24a* marks distal nephron tubule segment injury and correlates with *Lcn2*. Related to Figure 4.

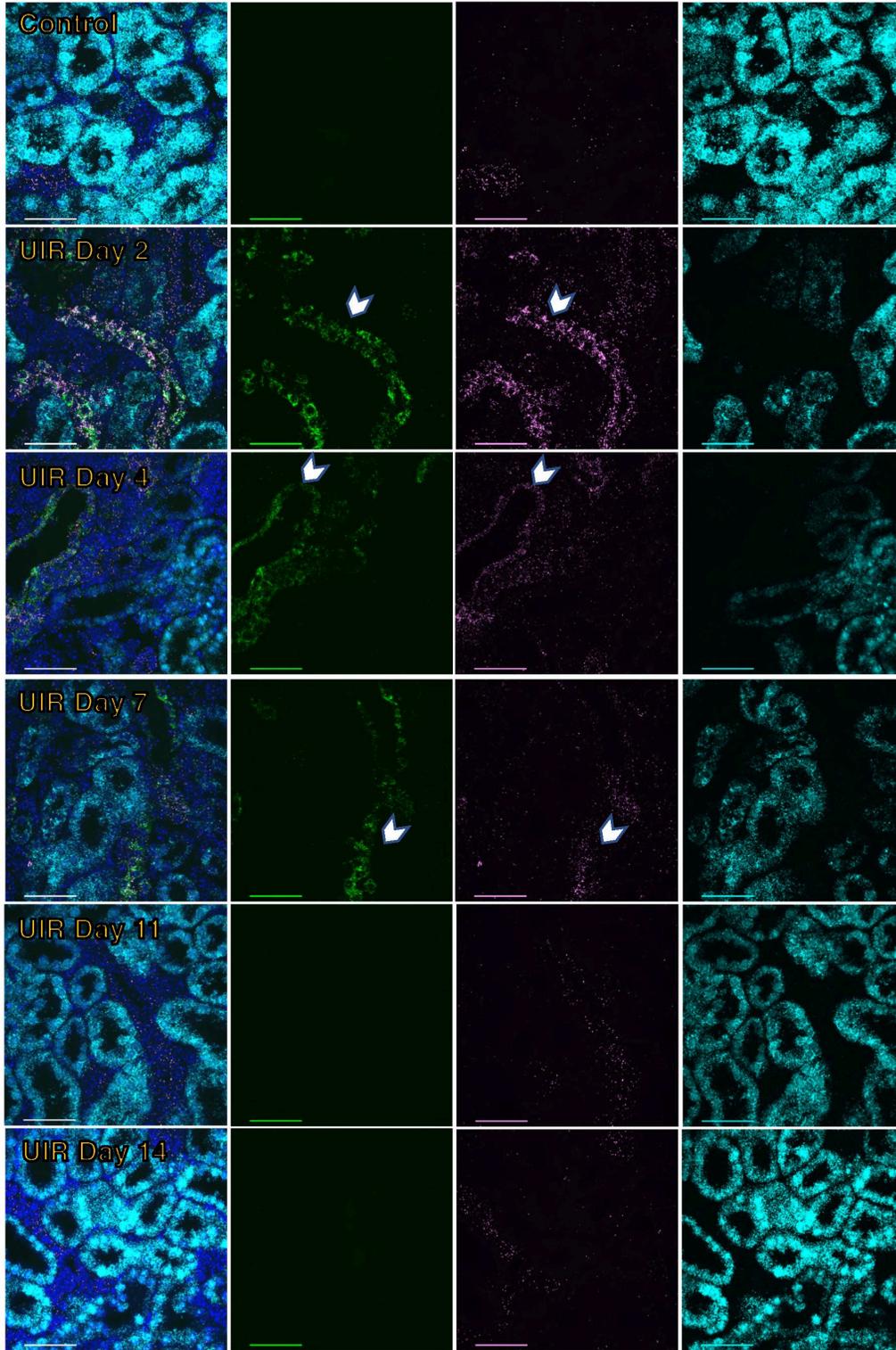
(A) 60x RNAscope images show *Lcn2* (green) and *Cd24a* (pink) colocalization at UIR Day 2, 4 and 7 highlighted with pointers, 60x Nyquist zoom, 0.21 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from Z-stack, scale 50 μm .

(B) Pearson's correlation analysis of *Cd24a* and *Lcn2* transcript numbers in UIR Day 1, n=9 Z-stacks (~60 tubules).

(C) IMARIS quantification of *Cd24a* in the UIR Day 1, 2, 4 and 7 and Control *Slc34a1*-positive renal tubules, n=9 Z-stacks (~60 tubules) per group, analyzed with one-way ANOVA with Bonferroni and Holm, ** $p < 0.01$ compared to the Control.

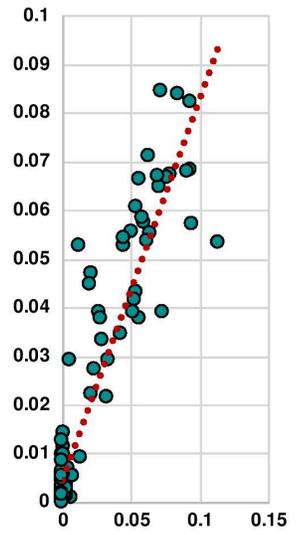
(D) IMARIS quantification of *Cd24a* in the *Lcn2* vs *Slc34a1*-positive renal tubules at UIR Day 1, 2, 4 and 7, n=9 Z-stacks (~60 tubules) per group, Student's *t* test, **** $p < 0.0001$.

A *Lcn2* *Cd24a* *Slc34a1* DAPI

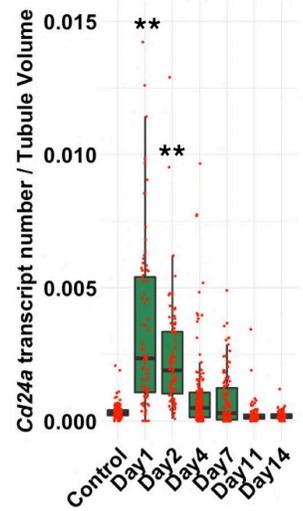


B *Lcn2* and *Cd24a* Correlation

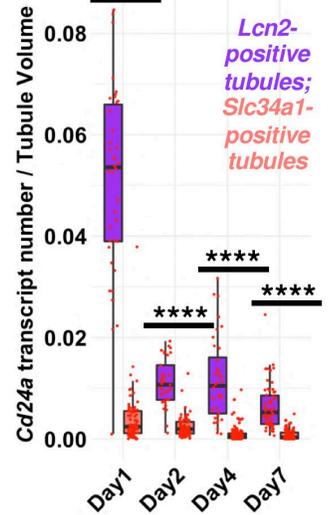
RSQ = 0.861; pValue < 0.0001



C *Cd24a* in *Slc34a1*⁺

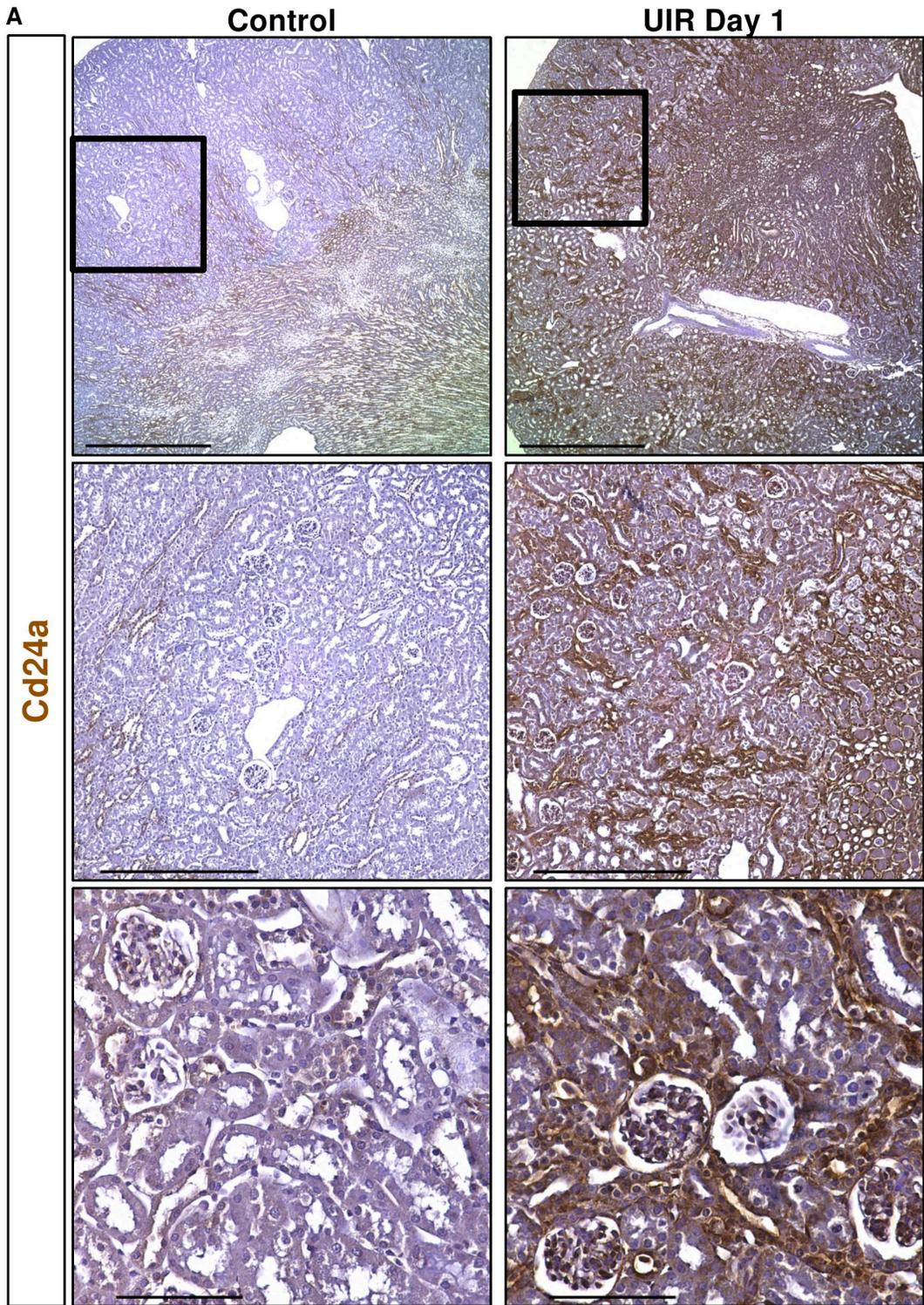


D



Supplemental Figure 11. UIR Day 1 exhibits marked Cd24a elevation in the cortical and medullary tubules. Related to Figure 4.

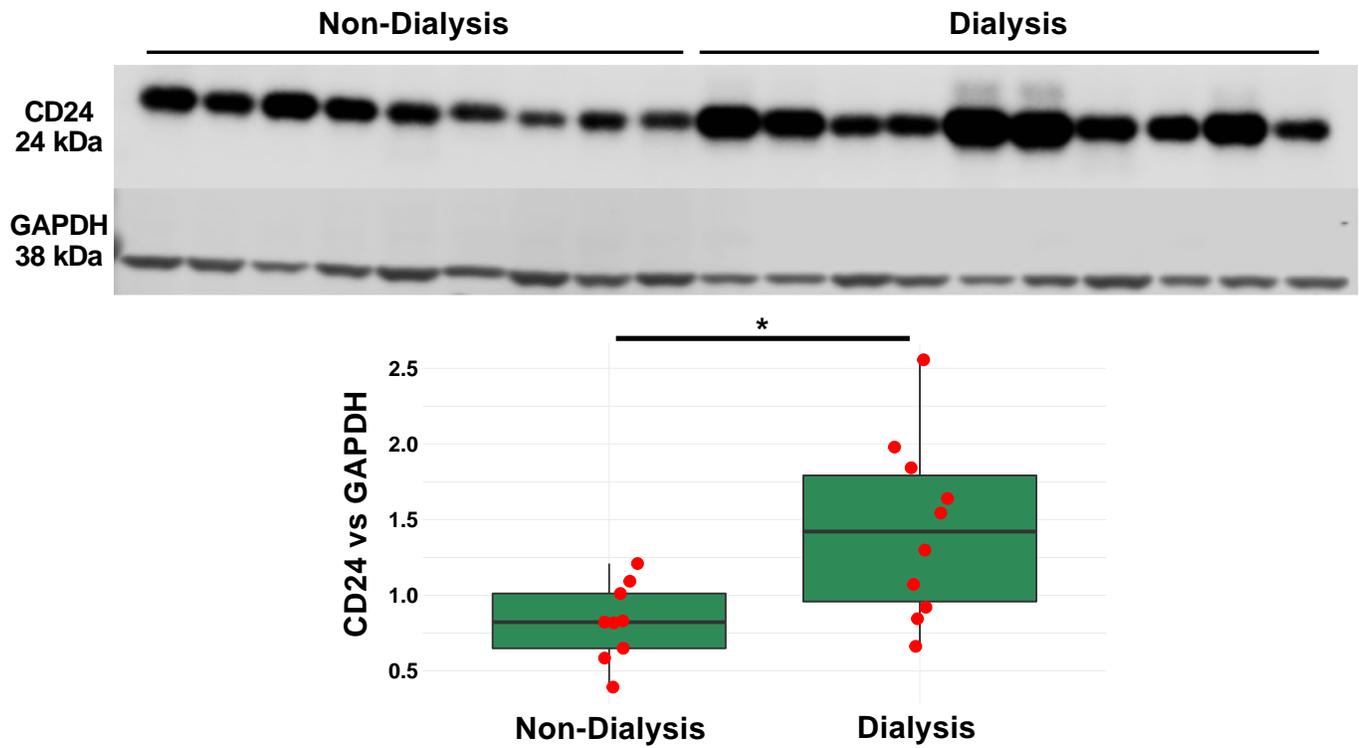
(A) Cd24a immunohistochemistry at UIR Day 1 vs Control. Black boxed show the 10x zoom into the cortex. 4x, 1000 μ m, 10x, 500 μ m, 40x, 100 μ m scale.



Supplemental Figure 12. Validation of AKI induced genes in the human kidney samples. Related to Figure 4.

(A) Western blot showing CD24 expression in human kidney biopsies from non-dialysis and dialysis patients, n=9-10 per group, Student's *t* test, * pValue<0.05, Fold Change 1.753 compared to Control.

A



Supplemental Figure 13. Pro-fibrotic signaling in the injured kidney. Related to Figures 5, 6 and 7.

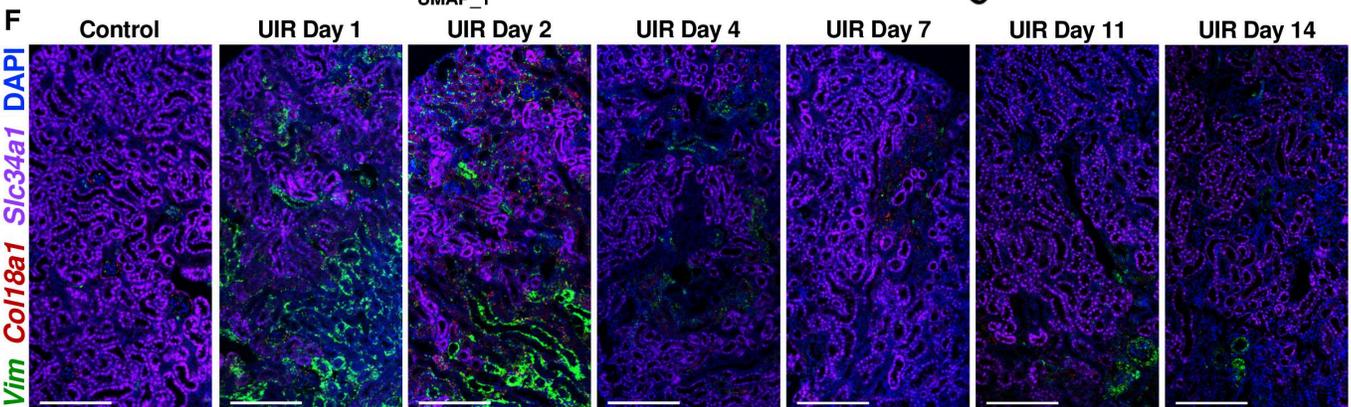
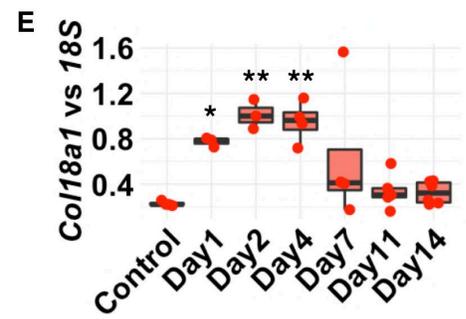
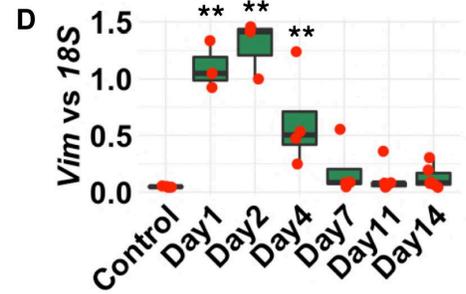
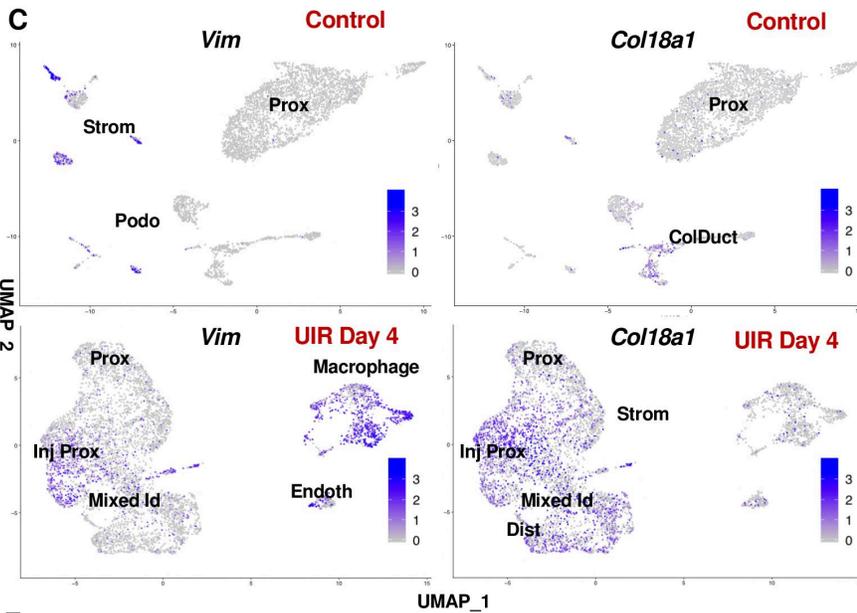
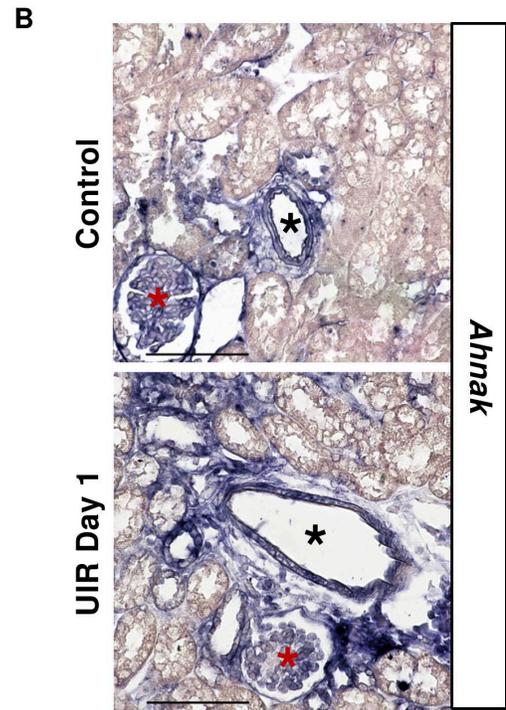
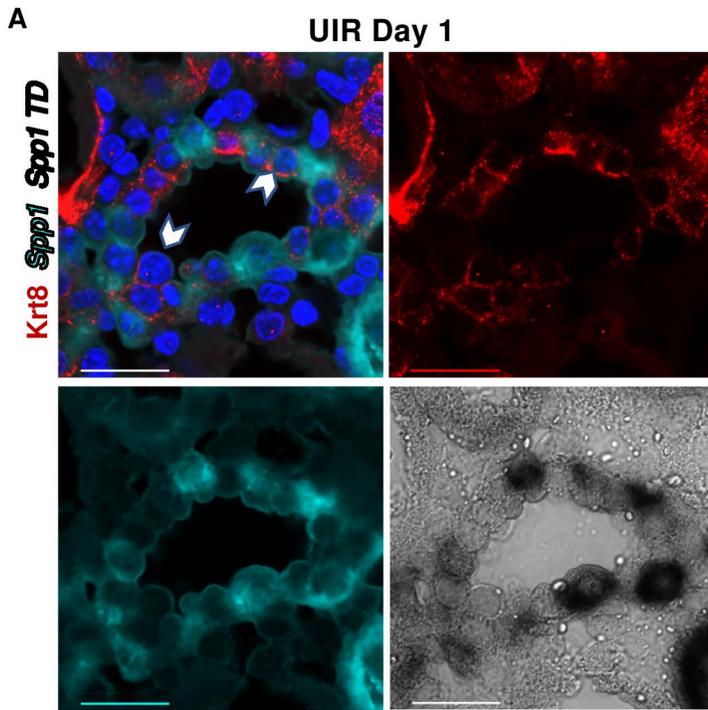
(A) *Spp1* CISH (cyan), Krt8 IF (red), DAPI (blue), UIR Day 1, 60x 0.09 px/ μ m Nyquist zoom, scale 25 μ m. Pointers show *Spp1* and Krt8 overlap. The combined scRNA-seq and CISH results define the precise cell types with elevated *Spp1* and *Krt8* elevated expression following UIR. TD, transmitted detector, showing the chromogenic *Spp1* CISH signal.

(B) *Ahnak* CISH shows perivascular (black stars) and glomerular (red stars) expression in the Control and UIR Day 1. 40x, 100 μ m scale.

(C) Feature Plots show *Vim* and *Col18a1* in the UIR Day 4 vs Control renal cell populations. Prox, proximal tubules, Podo, podocytes, Strom, stromal, Endoth, endothelial, Inj Prox, Injured Proximal tubules, Mixed Id, mixed identity cells, Dist, distal tubules, ColDuct, collecting duct.

(D and E) qPCR shows *Vim* and *Col18a1* gene expression changes over the AKI course, analyzed with one-way ANOVA with Bonferroni and Holm, * $p < 0.05$, ** $p < 0.01$ compared to the Control.

(F) RNAscope images show strong *Slc34a1* (purple) proximal tubule marker and low *Vim* (green) and *Col18a1* (red) expression in the interstitial and periglomerular spaces of the Control kidney. UIR causes significant *Slc34a1* decline and *Vim* and *Col18a1* elevation in the proximal tubules and in the stroma, which resolves at Day 11 and 14, with some remaining expression around the glomeruli. 10x, 500 μ m scale.



Supplemental Figure 16. Negative controls for *Sox4*, *Cd24a*, *Lcn2*, *Kim1*, *Slc34a1*, *Vim*, *Col18a1* RNAscope probes.

(A) Negative control for *Slc34a1* (pink), *Sox4* (green) and *Kim1* (white) RNAscope probes, DAPI (blue), 0.14 $\mu\text{m}/\text{px}$, Maximal Intensity Projection (MaxIP) from Z-stack, scale 50 μm .

(B) Negative control for *Cd24a* (pink), *Lcn2* (green) and *Slc34a1* (cyan) RNAscope probes, DAPI (blue), 60x Nyquist zoom, 0.21 $\mu\text{m}/\text{px}$, MaxIP from Z-stack, scale 50 μm .

(C) Negative control for *Col18a1* (red), *Vim* (green) and *Slc34a1* (purple) RNAscope probes, DAPI (blue), 60x Nyquist zoom, 0.21 $\mu\text{m}/\text{px}$, MaxIP from Z-stack, scale 50 μm .

