

p27^{Kip1}: The “Rosebud” of Diabetic Nephropathy?

GUNTER WOLF* and STUART J. SHANKLAND†

*Department of Medicine, Division of Nephrology and Osteology, University of Hamburg, Hamburg, Germany; and †Department of Medicine, Division of Nephrology, University of Washington, Seattle, Washington.

“Rosebud” was the last enigmatic word of dying protagonist Charles Foster Kane in Orson Welles’ 1941 masterpiece *Citizen Kane*, considered by many cineastes as the best movie ever made. The search for its meaning is the film’s theme. The investigative reporter thinks that by discovering what or who “rosebud” was will possibly provide a simple secret to Kane’s mysterious, complex life. We are facing an anticipated epidemic of ends-tage renal disease in the next decade, which will largely be due to diabetic nephropathy. This reflects our inability to prevent and control the disease; therefore, anything that has promise to contribute to controlling diabetic nephropathy is a cause for considerable excitement.

What is p27^{Kip1}, and is it the missing link in understanding the pathogenesis and potential treatment of diabetic nephropathy? This question is posed by Awazu *et al.* in this issue of *JASN*. In attempting to answer this question, we must first understand the complicated mechanisms of cell cycle regulation, which have been a major focus of many laboratories over the past two decades, culminating in awarding the Nobel prize for medicine in 2001 to Hartwell and Hunt. Depending on the cell type and underlying specific cell injury (1), renal cells respond by undergoing proliferation, apoptosis, de-differentiation, or hypertrophy (2,3).

Morphometric studies on renal specimens from patients with type 1 and 2 diabetes show characteristic glomerular hypertrophy, particularly of mesangial cells, but also of endothelial cells, early in diabetic nephropathy (1,4,5). Hypertrophy is biochemically defined as an increase in protein and RNA content, but without DNA replication (6). Morphometrically, this is accompanied by an increase in cell size. A more detailed analysis of the evolution of diabetic nephropathy that can only be performed in animal models shows a biphasic mesangial growth response, with early proliferation and subsequent hypertrophy (7). These growth responses to the diabetic environment are ultimately governed at the level of the nucleus (5), providing a rationale for the study of cell cycle regulatory

proteins in diabetic nephropathy, as was undertaken by Awazu *et al.* in this issue of *JASN* (8).

It has been known for more than a century that the growth of a cell has two main phases: interphase and mitosis. Figure 1 shows that interphase is further divided into G₁, S, and G₂ phases (9). Nondividing cells enter the G₁ phase from a quiescent G₀ phase, pass through G₁ into the S phase, where DNA replication takes place, and enter mitosis after progressing through G₂.

Once in G₁, a cell can undergo three fates. First, the classic proliferation pathway requires that cells progress from G₁, through S and M phases, followed by cytokinesis (cell division) (Figure 1). Second, upon G₁ entry, cells can exit G₁ or any phase of the cell cycle to undergo apoptosis. Third, and the focus of this editorial, is that after G₁ entry, there is arrest at the G₁/S transition, which prevents DNA synthesis. However, under these circumstances, because the protein content increases in G₁ phase (in anticipation of the cell dividing into two daughter cells), a block at G₁/S phase prevents DNA synthesis, resulting an increase in the protein:DNA ratio, which causes an increase in cell size (hypertrophy) (Figure 1).

Cell Cycle Regulatory Proteins Govern Cell Growth

What regulates the phases of the cell cycle, and how does this relate to diabetic nephropathy? Transitions between the different phases of the cell cycle are governed by positive (cyclins and cyclin-dependent kinases [CDK]) and negative (CDK-inhibitors) cell cycle regulatory proteins (Figure 2). Cyclins bind to and activate specific CDK in each phase of the cell cycle. In early G₁, D-type cyclins (D1, 2, 3) associate with CDK4 and CDK6, and cyclin E associates with CDK2 in late G₁. Cyclin A binds to CDK2 at the G₁/S phase boundary, and it is essential for DNA synthesis. Finally, cyclins B1 and B2 form complexes with cdc2 (formerly CDK1) during mitosis. The activated heterodimer cyclin-CDK complexes exhibit kinase activity and in turn phosphorylate other target proteins that are necessary for cell cycle progression (10).

Although cyclin-CDK may have a role in diabetic nephropathy, Awazu *et al.* (8) and other investigators, Terada, Safirstein, Wolf, Shankland, Preisig, and Mason (11–17), have focused on specific CDK-inhibitors in the pathogenesis of renal cell hypertrophy. The rationale for this is that, although there is cell cycle entry in diabetes, cell cycle progression is halted. CDK-inhibitors are relatively small molecules that bind to cyclin-CDK complexes, inhibit their activity, and thus halt cell cycle progression (Figure 2). Two main families of CDK-inhibitors exist (18): the INK4 family inhibit only D-type

Correspondence to Dr. Stuart J. Shankland, Associate Professor of Medicine, Director, Nephrology Fellowship Program, Division of Nephrology, Box 356521, University of Washington Medical Center, 1959 NE Pacific Avenue, Seattle, WA 98195-6521. Phone: 206-543-3792; Fax: 206-685-8661; E-mail: stuartjs@u.washington.edu

1046-6673/1403-0819

Journal of the American Society of Nephrology
Copyright © 2003 by the American Society of Nephrology

DOI: 10.1097/01.ASN.0000057518.58420.E4

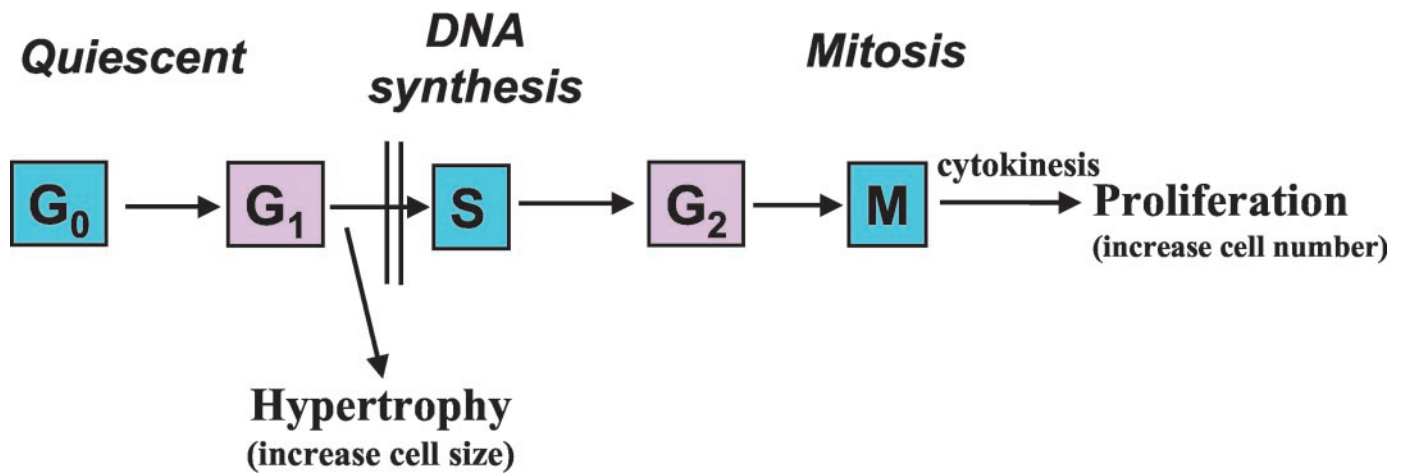


Figure 1. Phases of the cell cycle. Sequential transition through each phase of the cell cycle results in proliferation. In contrast, when cells engage G₁ phase but arrest at G₁/S, they undergo hypertrophy.

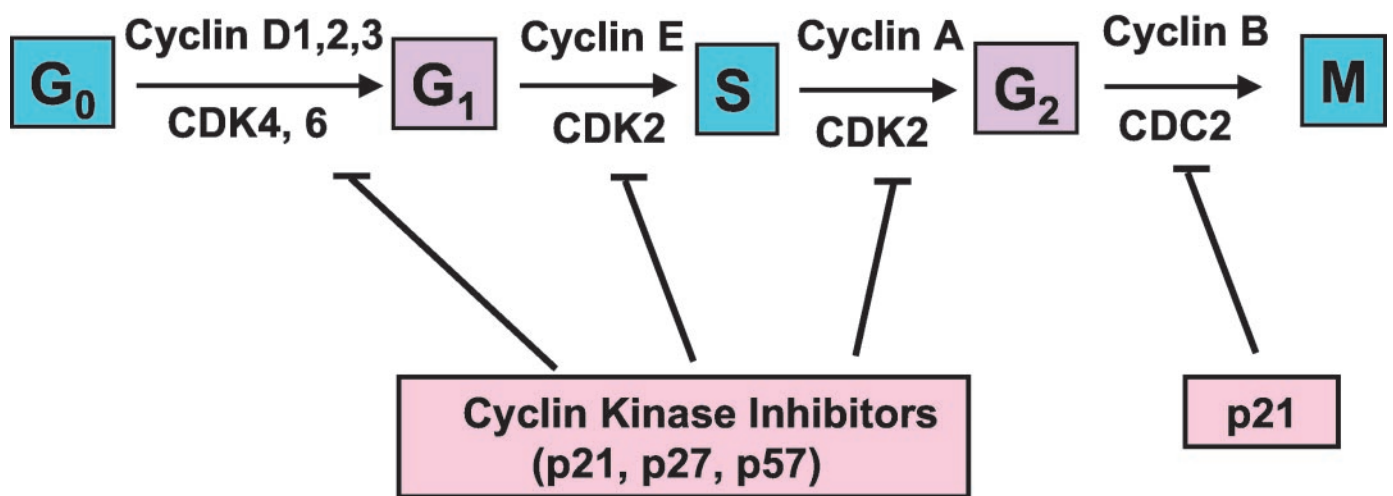


Figure 2. Cell cycle regulatory proteins. Progression through each phase of the cell cycle requires that cyclins bind to and activate a partner cyclin dependent kinase (CDK). Specific CDK-inhibitors (p21, p27, p57) inactivate cyclin-CDK complexes in different phases of the cell cycle; in so doing, they prevent cell cycle progression.

cyclin-CDK complexes in G₁; the Cip/Kip (p21^{Cip1} and p27^{Kip1}) family are more promiscuous and inhibit CDK2, CDK4, and CDK6 in most phases of the cell cycle. An increase in CDK-inhibitors arrests cells in the G₁ phase of cell cycle, and a decrease in CDK-inhibitors is required for cycle progression and proliferation.

CDK-Inhibitors and Diabetic Nephropathy

Why study CDK-inhibitors in diabetic hypertrophy? The answer begins by examining the cell cycle kinetics during diabetic nephropathy. Cultured mesangial cells exposed to high glucose (450 mg/dl) undergo a biphasic growth response, being pro-proliferation early (within 24 h), followed by anti-proliferation due to G₁ arrest (19). Studies have shown that G₁ phase arrest is necessary for glucose-induced cellular hypertrophy and also for the increase in extracellular matrix proteins. Earlier studies focused on transforming growth factor- β (TGF- β)

and showed that glucose-induced G₁ phase arrest is mediated in part by the autocrine synthesis and activation of the anti-proliferative and hypertrophic cytokine (19,20). However, there is a growing body of literature showing that specific CDK-inhibitors are also critical determinants of diabetic nephropathy (see below) and that these are both dependent and independent of TGF- β .

Exposing cultured mesangial cells to high glucose increases the CDK-inhibitors, p21^{Cip1} and p27^{Kip1}, but does not influence mRNA abundance (13). The glomerular (predominantly mesangial) expression of p21^{Cip1} and p27^{Kip1} also increases in experimental type 1 and 2 diabetic nephropathy (14, 21). Treatment with an angiotensin-converting enzyme (ACE) inhibitor reduces p27^{Kip1} expression and prevents renal hypertrophy of diabetic rats (22). The increase in p27^{Kip1} requires protein kinase C activation and is also partly dependent on the induction of TGF- β (13). More recently, studies have shown

that high glucose increases p27^{Kip1} protein expression through posttranscriptional mechanisms involving MAP kinases that directly phosphorylate this protein (23). Mason and colleagues showed that glucose-induced connective tissue growth factor increases expression of the CDK-inhibitors p15, p21^{Cip1}, and p27^{Kip1} in mesangial cells undergoing hypertrophy (24). Taken together, these studies demonstrate that the levels of specific CDK-inhibitors, especially p27, increase in cultured cells exposed to high glucose.

What does the increase in p27^{Kip1} mean? The increase in p27^{Kip1} abundance due to high glucose concentrations causes this CDK-inhibitor to associate with and inhibit CDK2. This leads to cell cycle arrest at G₁/S and prevents DNA synthesis (13). In contrast to wild-type mesangial cells, when p27^{Kip1} knockout (–/–) mesangial cells are exposed to high glucose, they do not undergo G₁ arrest (25). However, reconstituting p27^{Kip1} in p27 –/– cells inhibits cells at G₁/S in the presence of high glucose (25). These *in vitro* results clearly demonstrate that p27^{Kip1} is required for glucose-induced cell cycle arrest.

A definitive and functional role for p27^{Kip1} in G₁-phase arrest and glucose-induced hypertrophy comes from studies utilizing p27^{Kip1} –/– mesangial cells. In contrast to wildtype (+/+) mesangial cells, high glucose fails to induce hypertrophy in p27^{Kip1} –/– mesangial cells (25). However, reconstituting p27^{Kip1} in p27^{Kip1} –/– cells with an inducible vector system restored the hypertrophic phenotype induced by high glucose. These studies show that p27 is required for glucose-induced hypertrophy in cultured mesangial cells.

The role for CDK-inhibitor p27 in diabetic hypertrophy *in vivo* has been enigmatic until the current study by Awazu *et al.* in the current issue of *JASN* (8). The authors induced experimental type 1 diabetes in p27^{Kip1} +/+ and –/– mice by streptozotocin injection and showed that blood glucose and BP were comparable between diabetic p27^{Kip1} +/+ and –/– mice. However, in contrast to the increase in the kidney weight to body weight ratio, glomerular volume, and mesangial expansion in diabetic p27^{Kip1} +/+ animals at 12 wk, these measures of hypertrophy did not increase in diabetic p27 –/– animals (8). Moreover, albuminuria only developed in diabetic p27^{Kip1} +/+ animals and was not detected in diabetic p27 –/– mice. Furthermore, despite a similar increase in glomerular TGF- β expression in diabetic p27^{Kip1} +/+ and –/– mice, the glomerular protein expression of fibronectin increased only in diabetic p27^{Kip1} wild-type mice.

The studies described above by Awazu *et al.* (8) focused on established glomerular hypertrophy. Is there a role for the CDK-inhibitor p27 in early experimental diabetic hypertrophy? Studies have shown that at 4 wk after the development of hyperglycemia, glomerulosclerosis, tubulointerstitial sclerosis, and vascular sclerosis, indices were significantly less in diabetic p27^{Kip1} –/– mice compared with the diabetic wild-type (26). Most interestingly, diabetic heterozygotic p27^{Kip1} +/- animals exhibited scores between those of p27^{Kip1} +/+ and –/– mice, suggesting that these animals are haplotype insufficient (26).

Is p27^{Kip1} the whole story, or do other CDK-inhibitors also have a role in mediating renal cell hypertrophy? Mice deficient

for the CDK-inhibitor p21^{Cip1} are partially protected from diabetic glomerular hypertrophy (15), and the lack of p21 also attenuates glomerular hypertrophy after a significant reduction in renal mass (12). Recent studies have also shown that although TGF- β inhibits proliferation in double p21^{Cip1}/p27^{Kip1} –/– mesangial cells (16), the hypertrophic growth effects of TGF- β were significantly reduced in the absence of both p21^{Cip1} and p27^{Kip1}. This finding indicates that more than one CDK-inhibitor is likely pivotal in mediating the maximal hypertrophic response (16).

Taken together, there is now a growing literature showing that the CDK-inhibitor p27^{Kip1} has a critical role beyond simply regulating proliferation. The study by Awazu *et al.* (8) shows that p27^{Kip1} also regulates hypertrophy (8). Studies have also shown that p27^{Kip1} safeguards against apoptosis (27, 28). What research directions should we be focusing on in the future? These are numerous and include testing the possibility that targeted reductions of p27^{Kip1} could potentially protect patients from diabetic nephropathy. We need to understand why p27^{Kip1} increases predominantly in mesangial cells and not in other renal cell types, how p27^{Kip1} expression is regulated post-translationally in diabetes, and if p27^{Kip1} mediates other forms of renal hypertrophy.

We all know the sled named Rosebud that had been tossed into a furnace is not the answer for Kane's life. It is rather the emblem of the security, hope, and innocence of childhood, which a man can spend his life seeking to regain. Similarly, knowing what p27^{Kip1} is does not necessarily explain how its control ameliorates diabetic nephropathy. It will certainly be a fascinating story to decipher the exact role of this cell cycle protein in the evolution of diabetic nephropathy and to determine how its manipulation can bring benefit to humans akin to those described in Awazu's study in mice.

References

1. Mogensen CE, Andersen MJ: Increased kidney size and glomerular filtration rate in early juvenile diabetes. *Diabetes* 22: 706–712, 1973
2. Shankland SJ, Wolf G: Cell cycle regulatory proteins in renal disease: Role in hypertrophy, proliferation, and apoptosis. *Am J Physiol* 278: F515–F529, 2000
3. Ekholm SV, Reed SI: Regulation of G1 cyclin-dependent kinases in mammalian cell cycle. *Curr Opin Cell Biol* 12: 676–684, 2000
4. Osterby R, Gundersen HJ: Glomerular size and structure in diabetes mellitus. I. Early abnormalities. *Diabetologia* 11: 225–229, 1975
5. Wolf G, Ziyadeh FN: Molecular mechanisms of diabetic renal hypertrophy. *Kidney Int* 56: 393–405, 1999
6. Fine LG: The biology of renal hypertrophy. *Kidney Int* 29: 619–634, 1986
7. Young BA, Johnson RJ, Alpers CE, Eng E, Gordon K, Floege J, et al: Cellular events in the evolution of experimental diabetic nephropathy. *Kidney Int* 47: 935–944, 1995
8. Awazu M, Omori S, Ishikura K, Hida M, Fujita H: The lack of cyclin kinase inhibitor p27(Kip1) ameliorates progression of diabetic nephropathy. *J Am Soc Nephrol* 14: 699–708, 2003
9. Shankland SJ: Cell cycle control and renal disease. *Kidney Int* 52: 294–308, 1997

10. Lees E: Cyclin dependent kinase regulation. *Curr Opin Cell Biol* 7: 773–780, 1995
11. Terada Y, Inoshita S, Nakashima O, Tamamori M, Ito H, Kuwahara M, et al: Cell cycle inhibitors (p27(Kip1) and p21(Cip1)) cause hypertrophy in LLC-PK1 cells. *Kidney Int* 56: 494–501, 1999
12. Megyesi J, Price PM, Tamayo E, Safirstein RL. The lack of a functional p21^{WAF1/CIP1} gene ameliorates progression to chronic renal failure. *Proc Natl Acad Sci USA* 96: 10830–10835, 1999
13. Wolf G, Schroeder R, Ziyadeh FN, Thaiss F, Zahner G, Stahl RAK: High glucose stimulates expression of p27Kip1 in cultured mouse mesangial cells: relationship to hypertrophy. *Am J Physiol* 42: F348–F356, 1997
14. Wolf G, Schroeder R, Thaiss F, Ziyadeh FN, Helmchen U, Stahl RAK: Glomerular expression of p27Kip1 in diabetic db/db mouse: Role of hyperglycemia. *Kidney Int* 53: 869–879, 1998
15. Al-Douahji M, Brugarolis J, Brown PAJ, Stehman-Breen CO, Alpers CE, Shankland SJ: The cyclin kinase inhibitor p21^{WAF/CIP1} is required for glomerular hypertrophy in experimental diabetic nephropathy. *Kidney Int* 56: 1691–1699, 1999
16. Monkawa T, Hiromura K, Wolf G, Shankland SJ: The hypertrophic effect of transforming growth factor-beta is reduced in the absence of cyclin-dependent kinase-inhibitors p21 and p27. *J Am Soc Nephrol* 13: 1172–1178, 2002
17. Franch HA, Shay JW, Alpers RJ, Preisig PA: Involvement of pRB family in TGFbeta-dependent epithelial cell hypertrophy. *J Cell Biol* 129: 245–254, 1995
18. Sherr CJ, Roberts JM: CDK inhibitors: Positive and negative regulators of G 1-phase progression. *Genes Dev* 13: 1501–1512, 1999
19. Wolf G, Sharma K, Chen Y, Ericksen M, Ziyadeh FN. High glucose-induced proliferation in mesangial cells is reversed by autocrine TGF- β . *Kidney Int* 42: 647–656, 1992
20. Ziyadeh FN, Sharma K, Ericksen M, Wolf G: Stimulation of collagen gene expression and protein synthesis in murine mesangial cells by high glucose is mediated by autocrine activation of transforming growth factor-beta. *J Clin Invest* 93: 536–542, 1994
21. Kuan CJ, Al-Douahji M, Shankland SJ. The cyclin kinase inhibitor p21^{WAF1. CIP1} is increased in experimental diabetic nephropathy: Potential role in glomerular hypertrophy. *J Am Soc Nephrol* 9: 986–993, 1998
22. Wolf G, Wenzel U, Ziyadeh FN, Stahl RAK. Angiotensin converting-enzyme inhibitor treatment reduces glomerul p16^{INK4} and p27^{Kip1} expression in diabetic BBdp rats. *Diabetologia* 42: 1425–1432, 1999
23. Wolf G, Hannken T, Zahner G, Shankland SJ, Stahl RAK: p44/42 MAP kinase directly phosphorylates the CDK-inhibitor p27(Kip1) (p27): Role in hypertrophy [Abstract]. *J Am Soc Nephrol* 12: 623A, 2001
24. Wahab NA, Weston BS, Roberts T, Mason RM: Connective tissue growth factor and regulation of the mesangial cell cycle: Role in cellular hypertrophy. *J Am Soc Nephrol* 13: 2437–2445, 2002
25. Wolf G, Schroeder R, Zahner G, Stahl RAK, Shankland SJ. High glucose-induced hypertrophy of mesangial cells requires p27^{Kip1}, an inhibitor of cyclin-dependent kinases. *Am J Pathol* 158: 1091–1100, 2001
26. Wolf G, Schanze A, Wenzel U, Shankland SJ, Stahl RAK, Amann K: p27(Kip1) (p27) knockout mice are protected of diabetic nephropathy [Abstract]. *J Am Soc Nephrol* 13: 164A, 2002
27. Ophascharoensuk V, Fero ML, Hughes J, Roberts JM, Shankland SJ: The cyclin-dependent kinase inhibitor p27Kip1 safeguards against inflammatory injury. *Nat Med* 4: 575–580, 1998
28. Hiromura K, Pippin JW, Fero ML, Roberts JM, Shankland SJ: Modulation of apoptosis by the cyclin-dependent kinase inhibitor p27^{Kip1}. *J Clin Invest* 103: 597–604, 1999

See related article, “The Lack of Cyclin Kinase Inhibitor p27(Kip1) Ameliorates Progression of Diabetic Nephropathy,” on pages 699–708.