Does Hemodialysis Increase Protein Breakdown? Dissociation between Whole-Body Amino Acid Turnover and Regional Muscle Kinetics

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Hemodialysis (HD) is a protein catabolic procedure. Whole-body amino acid turnover studies identify dialysate amino acid loss and reduced protein synthesis as the catabolic events; proteolysis is not increased. Regional amino acid kinetics, however, document enhanced muscle protein breakdown as the cause of the catabolism; muscle protein synthesis also increased but to a lesser magnitude than the increment in protein breakdown. This discordance between whole-body and regional kinetics is best explained by the contrasting physiology between the muscle and the liver. During HD, muscle releases amino acids, which then are taken up by the liver for de novo protein synthesis. There seems to be a somatic to visceral recycling of amino acids. Evidence supporting this concept includes the increased fractional synthetic rate of albumin and fibrinogen during HD. It should be emphasized that region- or organ-specific kinetics vary, and whole-body turnover is a composite of all of the visceral and somatic compartments taken together. Reduced whole-body protein synthesis may be a compensatory adaptation to dialysate amino acid loss with a consequent reduction in plasma amino acid concentration. Notwithstanding the protein catabolic nature of HD, evidence is accumulating that intradialytic nutritional supplementation may blunt its catabolic effect.

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Amino Acid Release across the Arm or the Leg

As shown in Table 1, Gutierrez et al. (7) measured amino acid exchange across the leg (nmol/100 g per min) in normal subjects who underwent sham HD with a bio-incompatible dialyzer and reported 13 and 109% increments in amino acid release during and after dialysis, respectively. When dialyzed with a biocompatible dialyzer, the same subjects did not show increased amino acid release. They also showed that the increase in amino acid release could be blocked by concurrent administration of indomethacin, a nonselective cyclo-oxygenase inhibitor. In sharp contrast, Ikizler et al. (8) and Raj et al. (9) found a marked increase in amino acid release (nmol/100 ml per min) using a biocompatible dialyzer. In Ikizler’s paper, forearm total amino acid balance in 11 patients was −65 pre-dialysis, and the negative balance widened to −334 during dialysis, representing a fractional decrement in balance of 414%. The value returned toward normal 2 h after HD. In Raj’s work, net essential amino acid balances across the leg were −131 and −290, and nonessential amino acid balances were −335 and −2067 before and during HD. The fractional decrements of amino acid balance for essential and nonessential amino acids were, respectively, 121 and 517%. There is no good explanation for these differences except for one factor, the study subjects, normal control versus ESRD patients. In Gutierrez’s study, the increment in amino acid release across the leg was accounted for, in large part, by increased blood flow to the leg after dialysis. In the other two studies, the increase in amino acid release was a consequence of greater arteriovenous concentration gradients, as limb blood flow was not significantly changed.

In Vivo Whole-Body Amino Acid Turnover Kinetics

Modern amino acid turnover kinetics generally uses stable isotopes. These studies are accomplished by primed-continuous infusion of labeled amino acids into the free amino acid pool, where, under isotope and substrate steady-state condition, dilution of the labeled amino acid by either intake or endogenous protein degradation is allowed to occur. By measuring this dilution in the plasma and by quantifying tracer loss as labeled \( ^{13} \text{C} \) carbon dioxide, one determines whole-body protein degradation and amino acid oxidation rates and indirectly estimates protein synthesis by mass balance method (10). Table 2 summarizes the results of whole-body flux studies during HD.

<table>
<thead>
<tr>
<th>Author (Reference)</th>
<th>Type</th>
<th>Subject</th>
<th>Dialyzer Location</th>
<th>Pre-HD</th>
<th>HD</th>
<th>Post-HD</th>
<th>HD/Pre-HD</th>
<th>Fractional Change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gutierrez (7)</td>
<td>Total AA</td>
<td>Normal</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Ikizler (8)</td>
<td>Total AA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Raj (9)</td>
<td>EAA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Gutierrez (7)</td>
<td>EAA</td>
<td>Normal</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Ikizler (8)</td>
<td>Non-EAA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
</tbody>
</table>

Table 1. Amino acid balance across the limb during hemodialysis in normal subjects and ESRD patients

<table>
<thead>
<tr>
<th>Author (Reference)</th>
<th>Type</th>
<th>Subject</th>
<th>Dialyzer Location</th>
<th>Pre-HD</th>
<th>HD</th>
<th>Post-HD</th>
<th>HD/Pre-HD</th>
<th>Fractional Change</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gutierrez (7)</td>
<td>Total AA</td>
<td>Normal</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Ikizler (8)</td>
<td>Total AA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Raj (9)</td>
<td>EAA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Gutierrez (7)</td>
<td>EAA</td>
<td>Normal</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
<tr>
<td>Ikizler (8)</td>
<td>Non-EAA</td>
<td>ESRD</td>
<td>C</td>
<td>Pre-HD</td>
<td>HD</td>
<td>Post-HD</td>
<td>HD/Pre-HD</td>
<td>Fractional Change</td>
</tr>
</tbody>
</table>
leucine to protein incorporation, reflecting protein synthesis, was reduced. This may be due, in part, to leucine loss into the dialysate, accounting for approximately 12% of total flux. The leucine oxidation rate was not increased but was, in fact, reduced during dialysis. The combined effects of dialysate amino acid loss and reduced protein synthesis resulted in a significant reduction in net whole-body protein balance during HD.

Ikizler et al. (8) measured protein turnover in ESRD patients before, during, and for 2 h after HD using $^{13}$C leucine infusion. Dialysis sessions were 4 h in duration, and the dialysis membrane was polysulfone. They reported that HD led to a slight increase in whole-body proteolysis and a nonsignificant reduction in whole-body protein synthesis; net protein balance and plasma leucine were reduced. Dialysate leucine loss was approximately 16% of the leucine disappearance rate or the total flux rate.

In the above two studies, during the postdialysis period, amino acid loss ceased and whole-body protein synthesis improved. Proteolysis was not significantly different from baseline. Amino acid oxidation rate, which was reduced during HD, increased after dialysis. Net protein balance remained more negative compared with baseline, although of lesser magnitude than during dialysis. The catabolic picture after dialysis may be improved. Protein degradation is minimally changed from baseline; increments and decrements are minor in magnitude. By contrast, net protein balance is invariably reduced because of a combination of dialysate amino acid loss and decreased protein synthesis. After dialysis, net protein balance becomes less negative because dialysate amino acid loss ceases and protein synthesis improves.

Table 2. Whole-body amino acid turnover studies in ESRD patients during hemodialysis

<table>
<thead>
<tr>
<th>Author (Reference)</th>
<th>Isotope</th>
<th>Flux</th>
<th>Pre-HD/HD</th>
<th>Pre-HD/Post-HD</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>B</td>
<td>O</td>
</tr>
<tr>
<td>Lim (11)</td>
<td>$^{13}$C leucine</td>
<td>$\mu$mol/kg per h $\rightarrow$</td>
<td>118/117</td>
<td>17/14</td>
</tr>
<tr>
<td></td>
<td>$\Delta\rightarrow$</td>
<td>mg/kgFFM per min $\rightarrow$</td>
<td>3.4/3.7</td>
<td>0.5/0.4</td>
</tr>
<tr>
<td>Ikizler (8)</td>
<td>$^{13}$C leucine</td>
<td>$\Delta\rightarrow$</td>
<td>0.8/0.7</td>
<td>—</td>
</tr>
<tr>
<td>Raj (9)</td>
<td>$^{13}$C $\delta$ phenylalanine</td>
<td>$\Delta\rightarrow$</td>
<td>70/65</td>
<td>15/9</td>
</tr>
<tr>
<td>Veeneman (12)</td>
<td>$^{13}$C valine</td>
<td>$\Delta\rightarrow$</td>
<td>3.9/3.6</td>
<td>0.54/0.52</td>
</tr>
</tbody>
</table>

$^a$Listed under column Flux are the absolute values of the flux rates and fractional changes in each experiment. The latter representing increment or decrement comparing HD with pre-HD and post-HD with pre-HD. Two values are entered into the columns of breakdown (B), oxidation (O), synthesis (S), and net balance (NB) representing before and during and before and after HD. Column D lists the absolute amounts of unlabeled dialysate amino acid (the study amino acid) loss, and values in parentheses represent losses expressed as % of total flux. Only Lim’s study used a bio-incompatible dialyzer; the others used biocompatible membrane.

They found that, in the postabsorptive state, protein degradation and valine oxidation rates were not increased on the HD day; in fact, the values were lower than those obtained on a nondialysis day. Protein synthesis, however, was reduced, as was net protein balance. Dialysate valine loss was approximately 26% of total valine flux.

It is interesting to note that the different experiments listed in Table 2 covered a span of one decade, performed by different investigators at different institutions and with different isotopes, yet the results are consistent. During HD, whole-body protein degradation is minimally changed from baseline; increments and decrements are minor in magnitude. By contrast, net protein balance is invariably reduced because of a combination of dialysate amino acid loss and decreased protein synthesis. After dialysis, net protein balance becomes less negative because dialysate amino acid loss ceases and protein synthesis improves.

Regional Kinetics across the Forearm or Leg

To estimate regional turnover, one needs additional measurements of arterial and venous amino acid concentration, arterial and venous isotopic enrichment, and the blood flow rate across the region. This is usually performed across an arm or a leg (13). Labeled phenylalanine is typically used because it is neither synthesized nor metabolized by muscle; its appearance rate, therefore, represents protein degradation, and its disappearance represents protein synthesis. As shown in Table 3, Ikizler et al. (8) studied 11 ESRD patients and reported forearm kinetics ($\mu$g/100 ml per min) in the sequence of before, during, and after dialysis as follows: Proteolysis was 77, 180, and 127, respectively; protein synthesis was, 56, 123, and 98, respectively; and net balance was $-22$, $-58$, and $-28$, respectively. During HD, forearm muscle proteolysis increased by 134% and protein synthesis increased by 120%, resulting in net
The extent of increment is higher for protein breakdown. Over, protein catabolism. The individual components of protein turnover are consistent. HD leads to an increase in skeletal muscle net protein balance (14). Of note, during the postdialysis period, forearm net muscle protein balance was not different from baseline, although both breakdown and synthesis were significantly higher.

In a similar study, Raj et al. (9) measured leg kinetics (nmol/100 ml per min) using 13C6 phenylalanine and reported a 104 and 40% increase in protein degradation and synthesis, respectively, during HD. This resulted in a 93% reduction in net protein balance.

**Table 3. Regional and intramuscle amino acid kinetics during hemodialysis in ESRD patients**

<table>
<thead>
<tr>
<th>Author</th>
<th>Isotope</th>
<th>Location</th>
<th>Kinetics across Arm and Leg (Fractional Changes Pre-HD to HD)</th>
<th>Intramuscle Kinetics (%/day) FSR Protein (Fractional Changes Pre-HD to HD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ikizler (8)</td>
<td>2H2 phenylalanine</td>
<td>Forearm</td>
<td>↑ 134% ↑ 120% ↓ 164%</td>
<td></td>
</tr>
<tr>
<td>Pupim (14)</td>
<td>2H2 phenylalanine</td>
<td>Forearm</td>
<td>↑ 97% ↑ 85% ↓ 104%</td>
<td></td>
</tr>
<tr>
<td>Raj (9)</td>
<td>13C3 phenylalanine</td>
<td>Leg and muscle</td>
<td>↑ 104% ↑ 40% ↓ 93% ↑ 50%</td>
<td></td>
</tr>
</tbody>
</table>

*FSR, fractional synthesis rate of muscle protein.*

An increase in protein loss of 164% compared with baseline. In a subsequent study, the same investigators reported similar forearm kinetics showing increments of 97% for proteolysis, 85% for protein synthesis, and a 104% reduction in net protein balance (14). Of note, during the postdialysis period, forearm net muscle protein balance was not different from baseline, although both breakdown and synthesis were significantly higher.

In a similar study, Raj et al. (9) measured leg kinetics (nmol/100 ml per min) using 13C6 phenylalanine and reported a 104 and 40% rise in protein degradation and synthesis, respectively, during HD. This resulted in a 93% reduction in net protein balance.

**Three-Compartmental Muscle Kinetics**

Intramuscle protein metabolism can also be quantified directly by the three-compartmental model—artery, muscle, and vein—measuring muscle intracellular individual amino acid concentration and labeled amino acid enrichment; multiple muscle biopsies are needed for this approach (15). Raj et al. (16) studied intracellular amino acid transport kinetics using stable isotopes of three essential and two nonessential amino acids—namely, phenylalanine, leucine, lysine, alanine, and glutamine—before and during HD. They found that during HD, there was increased amino acid efflux from the intracellular compartment into the vein as a result of increased protein breakdown. Despite this, plasma amino acid levels were reduced, intracellular amino acid concentrations remained stable, and fractional muscle synthesis increased by 50% as listed in Table 3.

Overall, the studies of Ikizler, Raj, and Pupim are highly consistent. HD leads to an increase in skeletal muscle net protein catabolism. The individual components of protein turnover, i.e., breakdown and synthesis, both are increased, but the extent of increment is higher for protein breakdown.

**HD Is Protein-Catabolic: A Discussion**

All of the data presented in Tables 1, 2, and 3 indicate that HD is a protein-catabolic event because net protein balance, which is the difference between synthesis and degradation, becomes more negative during HD. Such a negative balance is seen both in the whole-body protein pool and in the regional muscle compartment. The mechanism that contributes to the negative protein balance in the two pools, however, differs. In the muscle compartment, the reduction in protein balance is due to increased protein breakdown that is not adequately compensated by an equivalent increment in synthesis. In the whole-body pool, reduced protein balance is accounted for by dialysate amino acid loss and a compensatory reduction in protein synthesis.

The discordant protein degradation finding between whole-body turnover and regional kinetics during HD, although counterintuitive, is not surprising because skeletal muscle accounts for only approximately 50% of whole-body protein content. It should be underscored that whereas region- or organ-specific kinetics vary, whole-body turnover is a composite results of all taken together. Tessari et al. (17) reported that, in humans, the kidney, the splanchic compartment, and the leg have different rates of protein degradation and synthesis. Failure to demonstrate increased protein breakdown with the whole-body kinetics can be explained by a compensatory downregulation of proteolysis in other body protein pools.

On the basis of the available information, we propose that this dissociation between whole-body and skeletal muscle turnover kinetics is due to the contrasting physiology between the muscle and the liver. In the fasting state, the study condition of all of the experiments listed in Tables 1, 2, and 3, the muscle releases amino acids to replete the plasma pool and the liver extracts these amino acids to synthesizes new proteins. The most direct way to test this hypothesis is to measure simultaneously hepatic vein amino acid concentration and labeled amino acid enrichment as well as hepatic blood flow rate during amino acid turnover study. Although important, these kinds of data are difficult to obtain in humans, but two lines of evidence are already available in the literature to support this concept. First, Felig and Wahren (18) catheterized simultaneously the brachial artery, the femoral vein, and the right main hepatic vein in normal adults and measured amino acid concentrations from these different sites. They found that, in the fasting state, net balance of many amino acids across the leg, i.e., arteriofemoral vein difference, is negative (Figure 1). By contrast, the net balance of many amino acids across the splanchnic bed, that is arteriohepatic vein difference, is positive (Figure 2). These data indicate that, in the fasting state, the muscle releases amino acids, which then are taken up by the liver for de novo protein synthesis. Second, evidence of increased hepatic protein synthesis during HD has already been demonstrated in ESRD patients, and the data are summarized.
in Table 4. Technically, amino acid kinetics could also be used to measure fractional synthesis rate of individual proteins, most commonly albumin and fibrinogen (19). In this method, the isotopes are detached from the infused amino acids and subsequently incorporated into new protein molecules. Raj et al. (9) reported that fractional synthesis rates, %/d, of albumin and fibrinogen increase by 49 and 67%, respectively, during HD. In another, similar protocol, Pupim et al. (21) reported that HD produced a 54% increase in albumin synthesis.

It is not clear whether the two processes—increased muscle protein degradation and increased liver protein synthesis—are intrinsically integrated, or a single HD-related perturbation stimulates both processes. Ikizler and Raj both reported positive correlations between fractional synthesis rates of albumin and fibrinogen and circulating IL-6 levels (9,20,21), suggesting the latter possibility.

This kind of reciprocal change between muscle and hepatic protein metabolism is not unique to dialysis patients. Mansoor et al. (22) found that during primed-constant 13C leucine infusion, patients with severe head injuries had a markedly reduced fractional synthesis rate of protein in the muscle, but the incorporation of tracer into albumin and fibrinogen was increased as compared with the control subjects. Similarly, Fearon et al. (23), using an intravenous loading infusion of 2H5 phenylalanine in cachectic cancer patients who had high C-reactive protein concentrations and hypoalbuminemia, found a higher fractional albumin synthesis than seen in normal subjects.

Despite the absence of accelerated whole-body protein breakdown, HD is, nonetheless, a catabolic event with regard to the whole-body protein economy as a result of a combination of amino acid loss and a compensatory reduction in protein synthesis. As shown in Table 2, dialysate amino acid losses are considerable, ranging from 12 to 26% of total flux. The reduction in whole-body protein synthesis could be explained by substrate deficiency as circulating essential amino acids are invariably reduced and synthesis could be stimulated with amino acid supplementation (12,14,24,25). Kobayashi et al. (26), in a sophisticated experiment, reported that HD-induced reduction in plasma amino acid concentration in normal swine inhibited leg muscle protein synthesis. This inhibition in synthesis was negated when plasma amino acid reduction during HD was alleviated by amino acid infusion. Muscle protein breakdown, in this study, was not changed by plasma amino acid reduction. Of note, despite unequivocal reduction in plasma amino acids, intracellular amino acid concentration was well maintained during HD. Which organs or tissues downregulate synthesis during HD is not clear. It could be any organ or tissue except the muscle and the central nervous system.

Muscle protein synthesis is increased, and central nervous system does not participate much in protein flux because its main source of fuel is glucose. It is possible that the liver, while increasing its synthesis of some urgently needed proteins, might downregulate synthesis of other proteins during a nutrition-deficient situation, such as the fasting state.

Discordance between regional muscle and whole-body protein synthesis kinetics have also been documented in non-HD settings. During recombinant human growth hormone infusion in healthy subjects, Fryburg and Barrett (27) found that protein synthesis, although increased in the forearm muscle, remained unchanged in the whole body. Adey et al. (28) found that patients with chronic renal failure, as compared with normal healthy subjects, have lower synthesis rates of mixed muscle

Figure 1. Net balance of individual amino acids across the leg (arteriofemoral venous differences [A-FV]) in 19 subjects in resting, postabsorptive state. The lines at the top or bottom of the bars represent the SEM. The mean A-FV differences were significantly different from zero for all amino acids except taurine and ornithine. Reprinted from reference 18.

Figure 2. Net balance of individual amino acids across the splanchnic bed (arteriohepatic venous differences [A-HV]) in 12 subjects in the resting, postabsorptive state. The lines at the top of the bars represent the SEM. Only those amino acids whose mean A-HV differences were significantly different from zero are shown. Reprinted from reference 18.
protein and myosin heavy chain but normal whole-body protein synthesis rates.

Notwithstanding the catabolic nature of HD, in the absence of concurrent disease, ESRD patients exhibit increased whole-body protein turnover and improved protein synthesis after maintenance dialysis initiation (2). Furthermore, negative protein balance during HD can be reversed with nutritional supplementation. Pupim et al. (14) studied seven ESRD patients at two separate HD sessions, one with and the other without intradialytic parenteral nutrition. Administration of 45 g of protein and 752 kcal resulted in an increase in plasma amino acid concentrations and a change in net protein balance from negative to positive values both in the forearm and in the whole-body pool. In another elegant study, Venneman et al. (12) measured whole-body valine flux in six ESRD patients on two separate dialysis sessions, one performed during fasting and the other during oral feeding. The supplement consisted of 0.62 g/kg protein and 15 kcal/kg energy divided into six oral feedings given every 30 min. They found that feeding during dialysis resulted in increased protein synthesis, and a change in net protein balance from negative to positive values. It is important to note here that insulin secretion after meal ingestion and insulin-induced protein anabolism both are intact in the ESRD patients (29,30).

**Conclusion**

The impetus for writing this article derives from comments made by nephrologists that they have conflicting impressions regarding the effect of HD on protein metabolism. We thus reviewed most of the literature on the studies of protein/amino acid metabolism during HD. We found that the major discordance is in the results obtained between whole-body and regional kinetics. Regional kinetics across the forearm or the leg showed marked increase in protein breakdown, accompanied by an increase in protein synthesis that is inadequate to compensate for the degradation, thus resulting in a reduction in protein balance across the region. Whole-body turnover studies, however, showed no change in protein synthesis, but there is dialysate amino acid loss, and protein synthesis is reduced. This combination results in a reduction of net protein balance in the whole-body pool. Reduced synthesis may be related, in part, to diminished substrate availability as plasma essential amino acids are reduced during HD. The organ or tissue that downregulates protein synthesis during HD is not identified.

Failure of the whole-body kinetics study to detect an increase in protein degradation is, we propose, due to contrasting physiology between the muscle and the liver; there seems to be a somatic to visceral recycling of amino acids. During HD, which happens also to be in the postabsorptive state in most of the experiments reported, the muscle releases amino acids and the liver recycles these amino acids for de novo protein synthesis.

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**References**


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**Table 4. Fractional synthesis rates of hepatic proteins during HD in ESRD patients**

<table>
<thead>
<tr>
<th>Author (Reference)</th>
<th>Isotope</th>
<th>Protein</th>
<th>FSR (%/d) Pre-HD</th>
<th>FSR (%/d) HD</th>
<th>FSR (%/d) Post-HD</th>
<th>Fractional Changes HD/Pre-HD</th>
<th>Fractional Changes Post-HD/Pre-HD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raj (9)</td>
<td>$^{13}$C$_6$ phenylalanine</td>
<td>Albumin</td>
<td>6.5</td>
<td>9.7</td>
<td>—</td>
<td>↑ 49%</td>
<td>—</td>
</tr>
<tr>
<td>Raj (9)</td>
<td>$^{13}$C$_6$ phenylalanine</td>
<td>Fibrinogen</td>
<td>11.8</td>
<td>19.7</td>
<td>—</td>
<td>↑ 67%</td>
<td>—</td>
</tr>
<tr>
<td>Caglar (20)</td>
<td>L[$^{13}$C]leucine</td>
<td>Albumin</td>
<td>7.4</td>
<td>12.2</td>
<td>10.9</td>
<td>↑ 65%</td>
<td>↑ 47%</td>
</tr>
<tr>
<td>Caglar (20)</td>
<td>L[$^{13}$C]leucine</td>
<td>Fibrinogen</td>
<td>11.2</td>
<td>15.1</td>
<td>17.7</td>
<td>↑ 35%</td>
<td>↑ 58%</td>
</tr>
<tr>
<td>Pupim (21)</td>
<td>L[$^{13}$C]leucine</td>
<td>Albumin</td>
<td>8.3</td>
<td>16.2</td>
<td>11.8</td>
<td>↑ 54%</td>
<td>↑ 51%</td>
</tr>
</tbody>
</table>

*Fractional changes represent increment or decrement comparing HD with pre-HD and post-HD with pre-HD values.*


