

Cyst Number but Not the Rate of Cystic Growth Is Associated with the Mutated Gene in Autosomal Dominant Polycystic Kidney Disease

Peter C. Harris,* Kyongtae T. Bae,[†] Sandro Rossetti,* Vicente E. Torres,* Jared J. Grantham,[‡] Arlene B. Chapman,[§] Lisa M. Guay-Woodford,^{||} Bernard F. King,* Louis H. Wetzel,[‡] Deborah A. Baumgarten,[§] Philip J. Kenney,^{||} Mark Consugar,* Saulo Klahr,[†] William M. Bennett,[¶] Catherine M. Meyers,** Qin (Jean) Zhang,[†] Paul A. Thompson,[†] Fang Zhu,[†] J. Philip Miller,[†] and the CRISP Consortium

*Division of Nephrology and Hypertension and Department of Radiology, Mayo Clinic College of Medicine, Rochester, Minnesota; [†]Department of Radiology, Medicine, and Division of Biostatistics, Washington University School of Medicine, St. Louis, Missouri; [‡]Kidney Institute and the Department of Internal Medicine, Kansas University Medical Center, Kansas City, Kansas; [§]Division of Nephrology and Department of Radiology, Emory University School of Medicine, Atlanta, Georgia; ^{||}Division of Genetics and Translational Medicine and Department of Radiology, University of Alabama at Birmingham, Birmingham, Alabama; [¶]Legacy Good Samaritan Hospital, Portland, Oregon; and **National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, Maryland

Data from serial renal magnetic resonance imaging of the Consortium of Radiologic Imaging Study of PKD (CRISP) autosomal dominant polycystic kidney disease (PKD) population showed that cystic expansion occurs at a consistent rate per individual, although it is heterogeneous in the population, and that larger kidneys are associated with more rapid disease progression. The significance of gene type to disease progression is analyzed in this study of the CRISP cohort. Gene type was determined in 183 families (219 cases); 156 (85.2%) had PKD1, and 27 (14.8%) had PKD2. PKD1 kidneys were significantly larger, but the rate of cystic growth (PKD1 5.68%/yr; PKD2 4.82%/yr) was not different ($P = 0.24$). Cyst number increased with age, and more cysts were detected in PKD1 kidneys ($P < 0.0001$). PKD1 is more severe because more cysts develop earlier, not because they grow faster, implicating the disease gene in cyst initiation but not expansion. These insights will inform the development of targeted therapies in autosomal dominant PKD.

J Am Soc Nephrol 17: 3013–3019, 2006. doi: 10.1681/ASN.2006080835

Autosomal dominant polycystic kidney disease (ADPKD) is characterized by progressive cyst development and expansion, resulting in ESRD in the majority of patients (1,2). ADPKD is genetically heterogeneous with two loci identified—*PKD1* (16p13.3) and *PKD2* (4q21)—that encode the proteins polycystin-1 and polycystin-2 (3–5). The majority of patients (approximately 85%) have PKD1, with PKD2 accounting for most of the remainder (6–8). Genetic modifying factors, as well as the environment, also significantly influence the course of this disease (9,10). On the basis of detected somatic mutations in isolated cystic linings and cell lines that are derived from single cysts, plus animal models, it has been proposed that cyst initiation is a two-hit process (11–15). However, dosage changes of a polycystin molecule also may result in cyst development (16–20), and heterogeneity of a

developing cyst questions whether a second hit is always necessary as an initiating event (21).

PKD2 is consistently a milder disease as evidenced in age at ESRD (PKD1 54.3 yr; PKD2 74 yr) and age at diagnosis of the disease and of hypertension (7,22,23). The Consortium of Radiologic Imaging Study of PKD (CRISP) was established to determine from a prospective, longitudinal study whether radiologic measures of kidney and cyst volumes by magnetic resonance imaging (MRI) could be used as an early means to monitor disease progression (24). This study showed that kidney and cyst volumes increase in most patients and that larger kidneys are associated with a decline in renal function (25). Previously, no significant difference was found between PKD1 and PKD2 kidneys by ultrasound analysis (7,26), but preliminary data from the CRISP study (before the genotyping was complete) showed that PKD1 kidneys are significantly larger than PKD2, consistent with correlations between renal size and function (25). Here, with completed genotyping data, PKD1 and PKD2 kidneys are compared in more detail, and insights are provided about the process of cystogenesis.

Published online ahead of print. Publication date available at www.jasn.org.

Address correspondence to: Dr. Peter C. Harris, Division of Nephrology, Mayo Clinic, 200 First Street SW, Rochester, MN 55905. Phone: 507-266-0541; Fax: 507-266-9315; E-mail: harris.peter@mayo.edu

Materials and Methods

Radiologic and Statistical Analyses

Detailed descriptions of the structure of this study, the baseline characteristics of the cohort, and details of evaluations during the course of the study have been published previously (24,25,27). Patients who had ADPKD ($n = 241$), were 15 to 46 yr of age, and had a GFR of >70 ml/min at enrollment were evaluated at baseline and annually over 3 yr for renal function and MRI of the kidney with coronal T₁- and T₂-weighted images to calculate renal and cyst volumes. Volumes were measured on a 3-mm slice-by-slice basis by two analysts who were blinded to genotype, as described previously in detail (24). The annual percentage change of total kidney and cyst volume was determined by regressing log-transformed (on a base-10 scale) against time (baseline to year 3) for individuals using a mixed linear model. To count the number of cysts, we chose a middle section of the left kidney on coronal T₂-weighted images, and any cyst with a diameter of ≥ 4 mm was recorded by a single analyst who was blinded to genotype.

The percentage change of corrected iothalamate clearance was calculated by dividing the slope estimate by the intercept value to measure GFR. Logistic regression was used to evaluate hypertension in PKD1 and PKD2 groups after adjustment for gender and age. Statistical methods used in this study included mixed-model ANOVA (and t test). Cross-tabulation comparisons were examined using χ^2 methods.

Genotyping Analysis

Details of the genetic study will be described elsewhere. Briefly, samples for genotyping were available from 239 patients from 202 different families. The *PKD1* and *PKD2* genes were screened in each family by denaturing HPLC (28), and mutation-negative cases (plus controls) were analyzed using a commercial diagnostic test (Athena Diagnostics, Worcester, MA) that uses direct sequencing. Larger deletion mutations also were sought by field inversion gel electrophoresis (29). The overall detection rate was 90.1%. Linkage analysis with markers flanking *PKD1* and *PKD2* also was used to identify gene type in three large families in whom no mutation was detected.

Results

Mutation analysis of the CRISP cohort identified likely pathogenic changes in 180 pedigrees (211 patients) with linkage identifying the gene in three more families (eight patients). Twenty-seven (14.8%) pedigrees (34 patients) had PKD2, and 156 (85.2%) pedigrees (185 patients) had PKD1, a distribution similar to that previously found in clinical ADPKD populations (6,7). Despite that the patients with PKD2 were significantly older, they were less likely to be hypertensive and had smaller kidney and cyst volumes at baseline (Table 1, Figure 1, A and B). The age- and gender-adjusted PKD2 kidney and cyst volumes were, respectively, 59.8 and 43.2% of their PKD1 counterparts. Although there was no difference in GFR or renal blood flow (another early marker of kidney function [30]) between PKD1 and PKD2 at baseline, there was significantly more urinary albumin in PKD1 cases (Table 1). Kidney and cyst volumes consistently increased in both the PKD1 and PKD2 populations, and the absolute rate of change was greater in PKD1 (25) (Figure 1, A through D, Table 1). However, this was due to the larger baseline sizes of the kidneys; the rates of growth for kidney and cyst volume were not significantly different (Table 1, Figure 1, A through D). This indicates that gene type does not strongly influence the size of ADPKD

kidneys by modulating the relative rate of cyst growth. Gender, however, was associated with both the absolute and relative rates of kidney and cystic expansion in the ADPKD population, with male patients showing more rapid expansion (Table 2).

Analysis of cyst number at baseline in all patients showed that PKD2 kidneys have significantly fewer cysts (55.9% of those found in PKD1 kidneys) and that there is a correlation between cyst number and kidney volume (Figure 1, E and F, Table 1; see the Materials and Methods section for details). In both PKD1 and PKD2, the number of cysts was correlated with the age of the patient, illustrating that new cysts develop during the course of the disease. Although the slopes of the regression lines that depict the relationship between age and cyst number are not significantly different between the PKD1 and PKD2 populations ($P = 0.77$; Figure 1E), the intersect to the y axis is significantly lower for PKD2 (approximately 14 cysts per MRI section; $P < 0.0001$), suggesting more aggressive early onset of cystogenesis in PKD1. Representative MRI images showing examples of younger and older PKD1 and PKD2 kidneys illustrate the differences in terms of cyst number, as well as total cystic volume (Figure 2), although there is considerable heterogeneity within each of the genic populations (Figure 1, E and F). Overall, these data indicate that PKD2 kidneys are smaller because they develop fewer cysts, especially at the early stages of the disease.

Discussion

Hypertension and urinary albumin excretion were significantly more common in PKD1 than PKD2, consistent with these variables' being associated with more severe disease (24,25). However, the major new conclusion from this study is that the genic effect is at the level of cyst initiation; the rate of cystic enlargement is not modulated by the disease gene. Although it is logical that the disease mutation is involved in cyst initiation, similar rates of cystic growth in the two disorders has not been shown previously. Therefore, two distinct phases to cystogenesis, a disease gene-related initiation phase and a gene-independent cyst enlargement phase, have been defined.

Gender was associated with the rate of cyst expansion in the total cohort. Previously, male individuals have been associated with more severe disease in ADPKD (2), and although this has been demonstrated in PKD2 (31), recent data on PKD1 have not shown a significant difference (22,32) in age at ESRD. Our data indicate that gender may be important, however, to the rate of cystic expansion, suggesting a hormonal influence on the process. The faster expansion of cyst volume in male individuals is consistent with the stimulating effect of testosterone on cAMP accumulation and chloride and fluid secretion by MDCK cells (33). A hormonal effect was identified previously in polycystic liver disease, in which more severe disease in women is thought to be promoted by estrogen exposure (34,35).

Our data lead us to suggest that new cysts develop during the life of the patient, although the expansion of microscopic cysts initiated *in utero* (36) to a level where they are recorded (≥ 4 mm), may be significant; and differential rates of growth of PKD1 and PKD2 microcysts cannot be ruled out. That cysts continue to develop in childhood and adulthood also is indi-

Table 1. Comparison of PKD1 versus PKD2 imaging/clinical parameters^a

Variable	PKD1 (n = 185)	PKD2 (n = 34)	P ^b
Baseline values			
hypertension	120 (65%)	17 (50%)	0.0998
			0.0205
mean age (yr)	31.77	35.42	0.0281
	<i>31.74</i>	<i>35.40</i>	0.028
mean kidney volume (ml)	1010.90	688.11	<0.0001
	<i>1040.85</i>	<i>622.56</i>	<0.0001
mean cyst volume (ml)	384.03	199.26	0.0011
	<i>397.56</i>	<i>171.60</i>	<0.0001
mean GFR (ml/min per 1.73 m ²)	96.92	96.67	0.96
	<i>95.97</i>	<i>99.71</i>	<i>0.39</i>
mean 24-h urinary albumin (g/L)	27.94	16.59	0.0108
	<i>28.35</i>	<i>16.31</i>	0.0078
mean number of cysts ^c	30.79	18.76	<0.0001
	<i>31.47</i>	<i>17.03</i>	<0.0001
total RBF ^d (ml/min per 1.73 m ² ; PKD1 n = 85; PKD2 n = 15)	731.1	731.02	0.9264
	<i>734.75</i>	<i>752.34</i>	<i>0.7681</i>
Changes per year ^e	(n = 179)	(n = 33)	
GFR (ml/min)	−1.11	−0.89	0.89
	<i>−0.91</i>	<i>−0.32</i>	<i>0.72</i>
kidney volume change (ml)	71.38	34.80	0.0055
	<i>74.94</i>	<i>32.217</i>	0.001
rate of kidney volume change (%)	5.55	4.69	0.24
	<i>5.68</i>	<i>4.82</i>	<i>0.24</i>
cyst volume change (ml)	70.69	28.72	0.0015
	<i>74.61</i>	<i>24.98</i>	0.0001
rate of cyst volume change (%)	12.21	11.40	0.58
	<i>12.46</i>	<i>12.08</i>	<i>0.79</i>

^aValues adjusted for age, gender, and the age and gender interaction (except for mean age, adjusted for gender only) are shown in italics. PKD, polycystic kidney disease; RBF, renal blood flow.

^bSignificant *P* values are in boldface.

^cMeasured from representative magnetic resonance imaging (MRI) section (see Materials and Methods for details).

^dRBF was available only on a subset of cases.

^eMean calculated over 3 yr of follow-up.

cated because, although the rate of cystic expansion is similar in both kidneys in an individual (25), there is considerable heterogeneity in cyst size (Figure 2). These concepts are presently being tested in conditional mouse models of ADPKD (37) and by further observations of the CRISP cohort. Because PKD1 kidneys have more cysts even at young ages (Figure 1, E and F), the rate of cyst initiation at early ages, including in the fetus, may be important.

The reason that fewer cysts develop in PKD2 is not known, but it seems to fit neatly with the concept that cystogenesis is a two-hit process that requires a somatic mutation for cyst initiation (11). Several factors suggest that the *PKD1* gene may sustain a higher level of somatic insults than *PKD2*. Most notable among these is the larger size of the coding region (approximately 12.9 kb compared with approximately 3 kb) and the GC richness of the DNA, resulting in a higher level of CpG dinucleotides that are known warm spots for

mutations (38). In addition, special factors, such as a polypyrimidine tract in IVS21 and six pseudogenes that match much of the 5' two thirds of *PKD1*, may increase the somatic mutation level at *PKD1* (29,39,40). A significant level of *de novo* germline mutations at *PKD1* emphasize that new mutations occur at a significant level at this locus (38). It is possible, however, that there are other reasons that a *PKD1* germline mutation might be more likely to result in cyst development. For example, polycystin-1 may be more important during renal development than polycystin-2, or some *PKD1* mutations may generate stable proteins that can act as dominant negatives and thus have an enhanced detrimental effect.

A comparison of patients within the *PKD1* or *PKD2* populations show that there is considerable variability in the rate at which cysts grow (Figure 1, C and D), although similar individual rates are found for the right and left kidneys (25). This

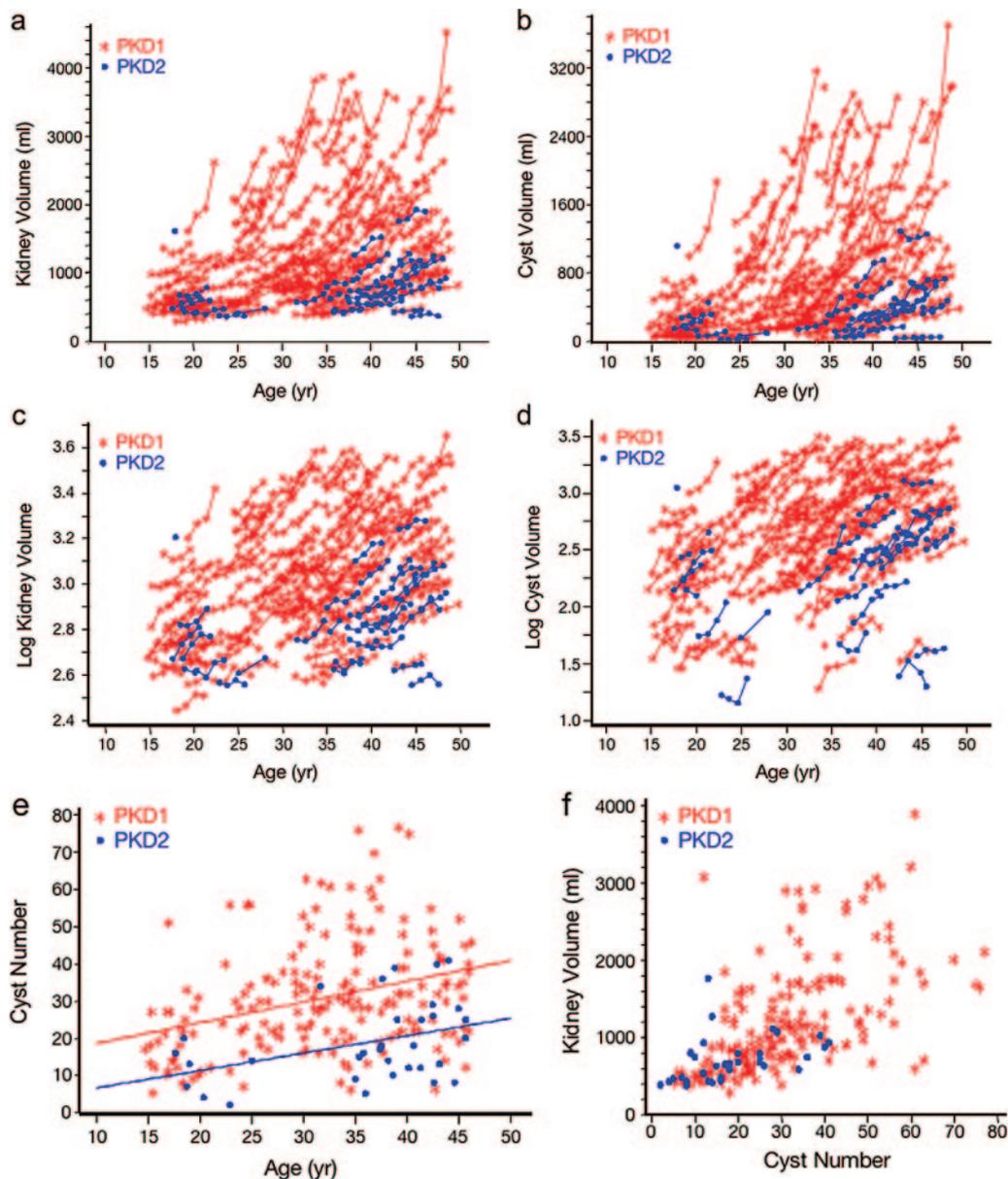


Figure 1. Plot of kidney (a and c) or cyst volume (b and d) on a linear (a and b) or log scale (c and d) versus age for all genotyped Consortium of Radiologic Imaging Study of PKD (CRISP) participants comparing patients with PKD1 (red asterisks) and PKD2 (blue circles). Patients with PKD2 generally have smaller kidney and cyst volumes, but the rate of increase (c and d) is similar to that of patients with PKD1. Cyst number compared with age (e) or kidney volume (f) in the PKD1 and PKD2 populations. Patients with PKD2 generally have lower cyst numbers. Regression lines (e) show that the relative rate of development of new cysts in the PKD1 and PKD2 populations is similar but that more cysts develop in PKD1 at a younger age.

rate seems to be independent of the disease gene and reflects genetic modifying effects, as well as environmental influences and gender. Similarly, considerable variation in cyst number is seen within the PKD1 and PKD2 populations (Figure 1, E and F), probably influenced by allelic effects, genetic modifiers, the environment, and stochastic factors on the rate of cyst initiation. These findings have implications for identifying quantitative trait loci that modulate disease severity and the development of effective therapeutics.

Conclusion

We have defined cystogenesis as a two-phase process: Cyst initiation, associated with the disease mutation, and cyst expansion, which is disease gene independent. Both phases vary between individuals. Therefore, quantitative trait loci or potential therapies may have an influence on the rate of cyst formation by preventing somatic mutations or by regulating the growth of cysts. Assuming that the downstream changes that are associated with cystogenesis as a result of disruption of the

Table 2. Comparison of gender with baseline values and measures of disease progression in the total ADPKD population^a

Variable	Male (n = 88)	Female (n = 131)	P ^b
Baseline values			
mean age (yr)	33.43	33.31	0.8224
kidney volume (ml)	861.52	751.00	0.0441
cyst volume (ml)	266.90	253.67	0.7028
cyst number ^c	25.86	22.59	0.0847
Change per year ^d			
kidney volume change (ml)	66.28	40.41	0.0062
rate of kidney volume change (%)	5.88	4.59	0.0163
cyst volume change (ml)	63.01	36.04	0.0039
rate of cyst volume change (%)	13.83	10.66	0.0029

^aValues are adjusted for age and genotype. ADPKD, autosomal dominant PKD.

^bSignificant P values are in boldface.

^cMeasured from representative MRI sections (see Materials and Methods for details).

^dMean calculated over 3 yr of follow-up.

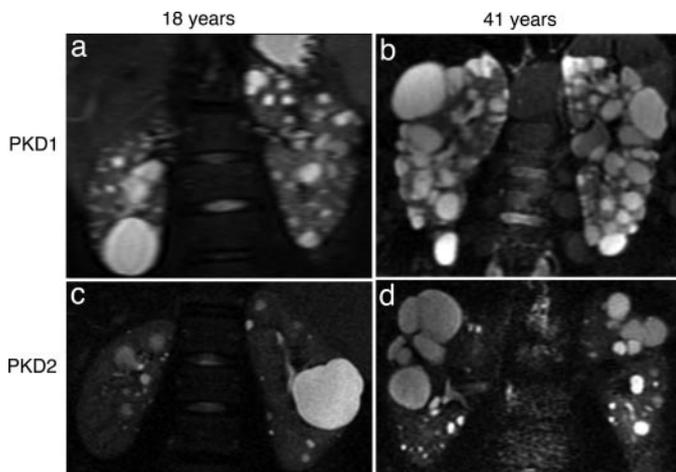


Figure 2. Coronal T₂-weighted, single-shot fast spin echo magnetic resonance images from patients with PKD1 (a and b) and PKD2 (c and d) at 18 yr (a and c) and 41 yr (b and d). Renal cysts in PKD1 are more numerous, diffusely distributed, and heterogeneous in size than those in PKD2.

polycystin complex are similar in PKD1 and PKD2, it is likely that factors that target cystic growth may be equally effective in both disorders. Most therapies that presently are under consideration, such as the clinical evaluation of vasopressin receptor antagonists (41), fall into this second group.

Acknowledgments

This study was supported by National Institute of Diabetes and Digestive and Kidney Diseases cooperative agreements (DK56934, DK56956, DK56957, and DK56961), with additional support for this ancillary study (DK56957-S1) for genetic analysis. The CRISP study also was supported by General Clinical Research Centers at each institution.

The study has been accepted as an abstract to the annual meeting of the American Society of Nephrology; November 17, 2006; San Diego, CA.

We thank the study coordinators Jody Mahan, Beth Stafford, Lorna Stevens, Kristin Cornwell, Vickie Kubly, Diane Watkins, Sharon Langley, and Pam Trull and Mary Virginia Gaines for managerial support. John McAuliffe, William Seltzer, Lynne Leclair, and Mark Smith at Athena Diagnostic are thanked for assistance in the fee-for-service direct sequence mutation analysis.

References

1. Grantham JJ, Chapman AB, Torres VE: Volume progression in autosomal dominant polycystic kidney disease: The major factor determining clinical outcomes. *Clin J Am Soc Nephrol* 1: 148–157, 2006
2. Gabow PA, Johnson AM, Kaehny WD, Kimberling WJ, Lezotte DC, Duley IT, Jones RH: Factors affecting the progression of renal disease in autosomal-dominant polycystic kidney disease. *Kidney Int* 41: 1311–1319, 1992
3. International Polycystic Kidney Disease Consortium: Polycystic kidney disease: The complete structure of the *PKD1* gene and its protein. *Cell* 81: 289–298, 1995
4. Hughes J, Ward CJ, Peral B, Aspinwall R, Clark K, San Millan JL, Gamble V, Harris PC: The polycystic kidney disease 1 (*PKD1*) gene encodes a novel protein with multiple cell recognition domains. *Nat Genet* 10: 151–160, 1995
5. Mochizuki T, Wu G, Hayashi T, Xenophontos SL, Veldhusien B, Saris JJ, Reynolds DM, Cai Y, Gabow PA, Pierides A, Kimberling WJ, Breuning MH, Deltas CC, Peters DJM, Somlo S: *PKD2*, a gene for polycystic kidney disease that encodes an integral membrane protein. *Science* 272: 1339–1342, 1996
6. Peters DJM, Sandkuijl LA: Genetic heterogeneity of polycystic kidney disease in Europe. *Contrib Nephrol* 97: 128–139, 1992
7. Torra R, Badenas C, Darnell A, Nicolau C, Volpini V, Revert L, Estivill X: Linkage, clinical features, and prognosis of autosomal dominant polycystic kidney disease types 1 and 2. *J Am Soc Nephrol* 7: 2142–2151, 1996
8. Daoust MC, Reynolds DM, Bichet DG, Somlo S: Evidence for a third genetic locus for autosomal dominant polycystic kidney disease. *Genomics* 25: 733–736, 1995

9. Paterson AD, Magistroni R, He N, Wang K, Johnson A, Fain PR, Dicks E, Parfrey P, St George-Hyslop P, Pei Y: Progressive loss of renal function is an age-dependent heritable trait in type 1 autosomal dominant polycystic kidney disease. *J Am Soc Nephrol* 16: 755–762, 2005
10. Fain PR, McFann KK, Taylor MR, Tison M, Johnson AM, Reed B, Schrier RW: Modifier genes play a significant role in the phenotypic expression of PKD1. *Kidney Int* 67: 1256–1267, 2005
11. Qian F, Watnick TJ, Onuchic LF, Germino GG: The molecular basis of focal cyst formation in human autosomal dominant polycystic kidney disease type 1. *Cell* 87: 979–987, 1996
12. Brasier JL, Henske EP: Loss of the polycystic kidney disease (PKD1) region of chromosome 16p13 in renal cyst cells supports a loss-of-function model for cyst pathogenesis. *J Clin Invest* 99: 194–199, 1997
13. Watnick TJ, Torres VE, Gandolph MA, Qian F, Onuchic LF, Klinger KW, Landes G, Germino GG: Somatic mutation in individual liver cysts supports a two-hit model of cystogenesis in autosomal dominant polycystic kidney disease. *Mol Cell* 2: 247–251, 1998
14. Nauli SM, Rossetti S, Kolb RJ, Alenghat FJ, Consugar MB, Harris PC, Ingber DE, Loghman-Adham M, Zhou J: Loss of polycystin-1 in human cyst-lining epithelia leads to ciliary dysfunction. *J Am Soc Nephrol* 17: 1015–1025, 2006
15. Wu G, D'Agati V, Cai Y, Markowitz G, Park JH, Reynolds DM, Maeda Y, Le TC, Hou J Jr, Kucherlapati R, Edelmann W, Somlo S: Somatic inactivation of Pkd2 results in polycystic kidney disease. *Cell* 93: 177–188, 1998
16. Jiang ST, Chiou YY, Wang E, Lin HK, Lin YT, Chi YC, Wang CK, Tang MJ, Li H: Defining a link with autosomal dominant polycystic kidney disease in mice with congenitally low expression of Pkd1. *Am J Pathol* 168: 205–220, 2006
17. Lantinga-van Leeuwen IS, Dauwerse JG, Baelde HJ, Leonard WN, van de Wal A, Ward CJ, Verbeek S, DeRuiter MC, Breuning MH, de Heer E, Peters DJM: Lowering of Pkd1 expression is sufficient to cause polycystic kidney disease. *Hum Mol Genet* 13: 3069–3077, 2004
18. Thivierge C, Kurbegovic A, Couillard M, Guillaume R, Cote O, Trudel M: Overexpression of PKD1 causes polycystic kidney disease. *Mol Cell Biol* 26: 1538–1548, 2006
19. Pritchard L, Sloane-Stanley JA, Sharpe J, Aspinwall R, Lu W, Buckle V, Strmecki L, Walker D, Ward CJ, Alpers CE, Zhou J, Wood WG, Harris PC: A human PKD1 transgene generates functional polycystin-1 in mice and is associated with a cystic phenotype. *Hum Mol Genet* 9: 2617–2627, 2000
20. Qian Q, Hunter LW, Li M, Marin-Padilla M, Prakash YS, Harris PC, Somlo S, Torres VE, Sieck GC: PKD2 haploinsufficiency alters intracellular calcium in vascular smooth muscle cells. *Hum Mol Genet* 12: 1875–1880, 2003
21. Nishio S, Hatano M, Nagata M, Horie S, Koike T, Tokuhisa T, Mochizuki T: Pkd1 regulates immortalized proliferation of renal tubular epithelial cells through p53 induction and JNK activation. *J Clin Invest* 115: 910–918, 2005
22. Hateboer N, van Dijk MA, Bogdanova N, Coto E, Saggarmalik AK, San Millan JL, Torra R, Breuning M, Ravine D: Comparison of phenotypes of polycystic kidney disease types 1 and 2. *Lancet* 353: 103–107, 1999
23. Torra R, Badenas C, Perez-Oller L, Luis J, Millan S, Nicolau C, Oppenheimer F, Mila M, Darnell A: Increased prevalence of polycystic kidney disease type 2 among elderly polycystic patients. *Am J Kidney Dis* 36: 728–734, 2000
24. Chapman AB, Guay-Woodford LM, Grantham JJ, Torres VE, Bae KT, Baumgarten DA, Kenney PJ, King BF Jr, Glockner JF, Wetzel LH, Brummer ME, O'Neill WC, Robbin ML, Bennett WM, Klahr S, Hirschman GH, Kimmel PL, Thompson PA, Miller JP: Renal structure in early autosomal dominant polycystic kidney disease (ADPKD): The Consortium for Radiologic Imaging Studies of Polycystic Kidney Disease (CRISP) cohort. *Kidney Int* 64: 1035–1045, 2003
25. Grantham JJ, Torres VE, Chapman AB, Guay-Woodford LM, Bae KT, King BF Jr, Wetzel LH, Baumgarten DA, Kenney PJ, Harris PC, Klahr S, Bennett WM, Hirschman GN, Meyers CM, Zhang X, Zhu F, Miller JP: Volume progression in polycystic kidney disease. *N Engl J Med* 354: 2122–2130, 2006
26. O'Neill WC, Robbin ML, Bae KT, Grantham JJ, Chapman AB, Guay-Woodford LM, Torres VE, King BF, Wetzel LH, Thompson PA, Miller JP: Sonographic assessment of the severity and progression of autosomal dominant polycystic kidney disease: The Consortium of Renal Imaging Studies in Polycystic Kidney Disease (CRISP). *Am J Kidney Dis* 46: 1058–1064, 2005
27. Bae KT, Commean PK, Lee J: Volumetric measurement of renal cysts and parenchyma using MRI: Phantoms and patients with polycystic kidney disease. *J Comput Assisted Tomogr* 24: 614–619, 2000
28. Rossetti S, Chauveau D, Walker D, Saggarmalik A, Winearls CG, Torres VE, Harris PC: A complete mutation screen of the ADPKD genes by DHPLC. *Kidney Int* 61: 1588–1599, 2002
29. European Polycystic Kidney Disease Consortium: The polycystic kidney disease 1 gene encodes a 14 kb transcript and lies within a duplicated region on chromosome 16. *Cell* 77: 881–894, 1994
30. King BF, Torres VE, Brummer ME, Chapman AB, Bae KT, Glockner JF, Arya K, Felmlee JP, Grantham JJ, Guay-Woodford LM, Bennett WM, Klahr S, Hirschman GH, Kimmel PL, Thompson PA, Miller JP: Magnetic resonance measurements of renal blood flow as a marker of disease severity in autosomal dominant polycystic kidney disease. *Kidney Int* 64: 2214–2221, 2003
31. Magistroni R, He N, Wang K, Andrew R, Johnson A, Gabow P, Dicks E, Parfrey P, Torra R, San-Millan JL, Coto E, Van Dijk M, Breuning M, Peters D, Bogdanova N, Ligabue G, Albertazzi A, Hateboer N, Demetriou K, Pierides A, Deltas C, St George-Hyslop P, Ravine D, Pei Y: Genotype-renal function correlation in type 2 autosomal dominant polycystic kidney disease. *J Am Soc Nephrol* 14: 1164–1174, 2003
32. Rossetti S, Burton S, Strmecki L, Pond GR, San Millan JL, Zerres K, Barratt TM, Ozen S, Torres VE, Bergstralh EJ, Winearls CG, Harris PC: The position of the polycystic kidney disease 1 (PKD1) gene mutation correlates with the severity of renal disease. *J Am Soc Nephrol* 13: 1230–1237, 2002
33. Sandhu S, Silbiger SR, Lei J, Neugarten J: Effects of sex hormones on fluid and solute transport in Madin-Darby canine kidney cells. *Kidney Int* 51: 1535–1539, 1997
34. Sherstha R, McKinley C, Russ P, Scherzinger A, Bronner T, Showalter R, Everson GT: Postmenopausal estrogen ther-

- apy selectively stimulates hepatic enlargement in women with autosomal dominant polycystic kidney disease. *Hepatology* 26: 1282–1286, 1997
35. Gabow PA, Johnson AM, Kaehny WD, Manco-Johnson ML, Duley IT, Everson GT: Risk factors for the development of hepatic cysts in autosomal dominant polycystic kidney disease. *Hepatology* 11: 1033–1037, 1990
 36. Reeders ST, Zerres K, Gal A, Hogenkamp T, Propping P, Schmidt W, Waldherr R, Dolata MM, Davies KE, Weatherall DJ: Prenatal diagnosis of autosomal dominant polycystic kidney disease with a DNA probe. *Lancet* 2: 6–8, 1986
 37. Piontek KB, Huso DL, Grinberg A, Liu L, Bedja D, Zhao H, Gabrielson K, Qian F, Mei C, Westphal H, Germino GG: A functional floxed allele of Pkd1 that can be conditionally inactivated in vivo. *J Am Soc Nephrol* 15: 3035–3043, 2004
 38. Rossetti S, Strmecki L, Gamble V, Burton S, Sneddon V, Peral B, Roy S, Bakkaloglu A, Komel R, Winearls CG, Harris PC: Mutation analysis of the entire PKD1 gene: Genetic and diagnostic implications. *Am J Hum Genet* 68: 46–63, 2001
 39. Burn TC, Connors TD, Dackowski WR, Petry LR, Van Raay TJ, Millholland JM, Venet M, Miller G, Hakim RM, Landes GM, Klinger KW, Qian F, Onuchic LF, Watnick T, Germino GG, Doggett NA: Analysis of the genomic sequence for the autosomal dominant polycystic kidney disease (PKD1) gene predicts the presence of a leucine-rich repeat. *Hum Mol Genet* 4: 575–582, 1995
 40. Watnick TJ, Gandolph MA, Weber H, Neumann HPH, Germino GG: Gene conversion is a likely cause of mutation in PKD1. *Hum Mol Genet* 7: 1239–1243, 1998
 41. Torres VE, Wang X, Qian Q, Somlo S, Harris PC, Gattone VH: Effective treatment of an orthologous model of autosomal dominant polycystic kidney disease. *Nature Med* 10: 363–364, 2004