

- Standl E, Wilcox RG, Wilhelmssen L, Betteridge J, Birkeland K, Golay A, Heine RJ, Koranyi L, Laaksa M, Mokan M, Norkus A, Pirags V, Podar T, Scheen A, Scherbaum W, Schernthaner G, Schmitz O, Shrha J, Smith U, Taton J, PROactive Investigators: Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive study (PROspective pioglitAzone Clinical Trial In macroVascular Events): A randomised controlled trial. *Lancet* 366: 1279–1289, 2005
4. Wayman N, Hattori Y, McDonald MC, Mota-Filipe H, Cuzzocrea S, Pisano B, Chatterjee PK, Thiemermann C: Ligands of the peroxisome proliferator-activated receptors (PPAR- γ and PPAR- α) reduce myocardial infarct size. *FASEB J* 16: 1027–1040, 2002
 5. Yang T, Soodvalia S: Renal and vascular mechanisms of thiazolidinedione-induced fluid retention. *PPAR Res* 943614, 2008
 6. Zhang H, Zhang A, Kohan DE, Nelson RD, Gonzalez FJ, Yang T: Collecting duct-specific deletion of peroxisome proliferator-activated receptor γ blocks thiazolidinedione-induced fluid retention. *Proc Natl Acad Sci USA* 102: 9406–9411, 2005
 7. Guan Y, Hao C, Cha DR, Rao R, Lu W, Kohan DE, Magnuson MA, Redha R, Zhang Y, Breyer MD: Thiazolidinediones expand body fluid volume through PPAR γ stimulation of ENaC-mediated renal salt absorption. *Nature Med* 11: 861–866, 2005
 8. Nofziger C, Chen L, Shane MA, Smith CD, Brown KK, Blazer-Yost BL: PPAR γ agonists do not directly enhance basal or insulin-stimulated Na⁺ transport via the epithelial Na⁺ channel. *Pflugers Arch* 451: 445–453, 2005
 9. Nofziger C, Brown KK, Smith CD, Harrington W, Murray D, Bisi J, Ashton TT, Maurio FP, Kalsi K, West TA, Baines D, Blazer-Yost BL: PPAR γ agonists inhibit vasopressin-mediated anion transport in the MDCK-C7 cell line. *Am J Physiol Renal Physiol* 297: F55–F62, 2009
 10. Chen L, Yang B, McNulty JA, Clifton LG, Binz JG, Grimes AM, Strum JC, Harrington WW, Chen Z, Balon TW, Stimpson SA, Brown KK: GI262570, a peroxisome proliferator-activated receptor γ agonist, changes electrolytes and water reabsorption from the distal nephron in rats. *J Pharmacol Exp Ther* 312: 718–725, 2005
 11. Vallon V, Hummler E, Rieg T, Pochynyuk O, Bugaj V, Schroth J, Dechenes G, Rossier B, Cunard R, Stockand J: Thiazolidinedione-induced fluid retention is independent of collecting duct α ENaC activity. *J Am Soc Nephrol* 20: 721–729, 2009
 12. Qayyum R, Adomaityte J: A meta-analysis of the effect of thiazolidinediones on blood pressure. *J Clin Hypertens* 8: 19–28, 2006
 13. Barroso I, Gurnell M, Crowley VE, Agostini M, Schwabe JW, Soos MA, Maslen GL, Williams TD, Lewis H, Schafer AJ, Chatterjee VK, O'Rahilly S: Dominant negative mutations in human PPAR γ associated with severe insulin resistance, diabetes mellitus and hypertension. *Nature* 402: 880–883, 1999
 14. Krämer BK, Bergler T, Stoelcker B, Waldegger S: Mechanisms of disease: The kidney-specific chloride channels ClCKA and ClCKB, the Barttin subunit, and their clinical relevance. *Nat Clin Pract Nephrol* 4: 38–46, 2008
 15. Bajwa PJ, Lee JW, Straus DS, Lytle C: Activation of PPAR γ by rosiglitazone attenuates intestinal Cl⁻ secretion. *Am J Physiol Gastrointest Liver Physiol* 297: G82–G89, 2009

Harnessing Transporters to Clear Uremic Toxins

Orson W. Moe

Department of Internal Medicine and Charles and Jane Pak Center of Mineral Metabolism and Clinical Research, University of Texas Southwestern Medical Center, Dallas, Texas

J Am Soc Nephrol 20: 2483–2484, 2009.
doi: 10.1681/ASN.2009101071

J Am Soc Nephrol 20: 2481–2489, 2009

The constancy of the *milieu intérieur* described by Claude Bernard is an absolute prerequisite for the health of the organism. The primacy of the kidney in whole organism homeostasis is best exemplified by the disturbance in the composition and amount of body fluid and solute when this organ fails to function. In simplistic terms, one can envision renal failure as the inability to remove what should be excreted and inability to add what should be added to the body. The inability to excrete the appropriate amount of water and various solutes is well studied and serves as a marker in clinical practice to evaluate the degree of dysfunction and adequacy of therapy. There are also many additional substances that are not properly excreted in renal failure, the accumulation of which contributes to the uremic state, but these are less well studied in terms of their metabolism, mechanism of action, or mode of excretion.

The expanding group of proven and putative uremic toxins has recently been highlighted by the European Uremic Toxin Work Group (<http://EUTox.info>) and the count has reached over 110 moieties.¹ This highly diverse group of substances ranges from inorganic solutes to organic substances including acids, guanidine, peptides, indoles, nucleotides, peptides, and others.² The chemical properties of these moieties are as expansive as their identities, with a broad range of molecular weight (outside the 10- to 30-kD middle molecule class), hydrophobicity, protein binding, a host of post-translational modifications, and a myriad of target organs and mechanisms to impart damage. Many of these molecules are normal constituents of the *milieu intérieur* when their levels are maintained within discreet ranges. Clearly no single biologic system will possess the broad span to handle the excretion of this vast group of molecules. Excretion of these molecules in health includes some contribution from hepatic conjugation, but they largely rely on glomerular filtration or tubular secretion by a host of transporter proteins.

In general, one can devise two broad categories of countermeasures to combat pernicious uremic toxins. First is attempting to directly block their actions (e.g., angiotensin receptor blockade) or neutralize downstream effects on target organs (e.g., alkali replacement), and a second is their enhanced removal either by improvement of dialytic clearance (e.g., large-pore dialysis membranes), inhibition of production (e.g., calcimimetics for parathyroid hormone), or enhancement of endogenous clearance perhaps by pharmacologic means. All of these approaches have been attempted for various uremic toxins over the years. In this issue of *JASN*, the paper by Toyohara and coworkers³ demonstrates success of one of the methods—enhancement of endogenous clearance—which has been attempted the least.

Published online ahead of print. Publication date available at www.jasn.org.

Correspondence: Dr. Orson W. Moe, Department of Internal Medicine and Charles and Jane Pak Center of Mineral Metabolism and Clinical Research, University of Texas Southwestern Medical Center, Dallas, TX 75390. Phone: 214-648-7993; Fax: 214-645-9442; E-mail: orson.moe@utsouthwestern.edu

Copyright © 2009 by the American Society of Nephrology

The mammalian renal proximal tubule, which is likely the most archaic part of the vertebrate nephron, is endowed with a multitude of solute transporters devoted to reabsorption, but also to secretion of unwanted substances. These transporters are usually committed to anionic, cationic, or neutral small organic molecules, but any given transporter typically can accept a vast array of substrates at least when tested in a heterologous system.⁴ Enomoto and Niwa⁵ previously presented us with an elegant summary of the potential roles of organic anion and cation transports in uremic toxin clearance and the reader is referred to this article for details.

The *SLCO* family of genes also encodes many transporters that accept an extremely broad range of substrates. Following up on a finding made by the same group that *SLCO4C1* expression is reduced in a 5/6 nephrectomy model of chronic kidney disease (CKD),⁶ Toyohara and co-workers³ conduct an interesting and informative set of experiments in this issue of *JASN* by overexpressing *SLCO4C1* in mice as a transgene to test whether these animals fare better when confronted with renal failure. Genetic overexpression of the organic anion transporter *SLCO4C1* in mice reduces levels of specific uremic toxins in the plasma such as asymmetric dimethylarginine (ADMA), guanidine succinate, and *trans*-aconitate, which is an inhibitor of the tricarboxylic acid cycle. The reduction of these toxins associates with changes in phenotype such as amelioration of hypertension and histologic indices of renal inflammation. Of note is that elevation of *trans*-aconitate in CKD and its hypertensive and pro-oxidative stress effects is novel. Because the probability of genetic manipulation of humans with CKD is not within a realistic realm in the near future, the authors next searched for agonists that activate the promoter of *SLCO4C1* *in vitro* and found the statin class of drugs are effective. The findings were next tested *in vivo* where the authors found that statins induced native *SLCO4C1* mRNA transcripts, increased ADMA and *trans*-aconitate clearance, and improved markers of left ventricular hypertrophy. Interestingly, there were no detectable effects in blood pressure at the dose of statin administered, suggesting that the reduction of left ventricular hypertrophy may not be secondary to blood pressure changes, which brings forth once again the entity of uremic cardiomyopathy that is unrelated to hypertensive or coronary heart disease.⁷ Finally, the authors provide a very comprehensible metabolomic analysis of the CKD model and effect of transgenic expression of *SLCO4C1* that is a valuable database for all investigators in this field.

This is one of the more convincing studies that show overexpression or induction of a transporter can alleviate some aspects of uremia. However, some caution is order in terms of the interpretation and generalization of the study. There were no data present to indicate ADMA, guanidine succinate, or *trans*-aconitate are direct substrates of the *SLCO1C4* transporter, so their increased clearance could be due to indirect effects of statins or downstream effects from

SLCO1C4 activity. The improvement in clearance is compatible with the uremic toxin model, but the changes in whole animal phenotype were modest. This latter point is because amelioration of a few uremic toxins among hundreds will likely have beneficial but incomplete effects. Moreover, genetic polymorphisms in *SLCO4C1* may exist that affect an individual's ability to handle selected uremic toxins. This caveat is compatible with the human literature of conflicting results of statin therapy on progression of CKD, cardiovascular complications, and mortality (reviewed in reference 8).

Nevertheless, this paper should not be viewed as a test of whether statins can be used to treat certain aspects of CKD—they might with proper validation—or be diminished by previous negative or equivocal clinical trials with statins. Rather, one should embrace a broader view to further explore the role of transporters and their activators in the handling of substances that become toxins in CKD with the prospect of therapeutically manipulating these transporters to enhance their activity in progressive renal disease.

DISCLOSURES

The National Institutes of Health and the Simmons Family Foundation support the author.

REFERENCES

1. Vanholder R, Baurmeister U, Brunet P, Cohen G, Glorieux G, Jankowski J: A bench to bedside view of uremic toxins. *J Am Soc Nephrol*, 19: 863–870, 2008
2. Vanholder R, Van Laecke S, Glorieux G: What is new in uremic toxicity? *Pediatr Nephrol*, 23: 1211–1221, 2008
3. Toyohara T, Suzuki T, Morimoto R, Akiyama Y, Souma T, Shiwaku HO, Takeuchi Y, Mishima E, Abe M, Tanemoto M, Masuda S, Kawano H, Maemura K, Nakayama M, Sato H, Mikkaichi T, Yamaguchi H, Fukui S, Fukumoto Y, Shimokawa H, Inui K, Terasaki T, Goto J, Ito S, Hishinuma T, Rubera I, Tauc M, Fujii-Kuriyama Y, Yabuuchi H, Moriyama Y, Soga T, Abe T: *SLCO4C1* transporter eliminates uremic toxins and attenuates hypertension and renal inflammation. *J Am Soc Nephrol* 20: 2546–2555, 2009
4. Moe OW, Palacin M, Wright S: *Renal Organic Solute Transport*, Philadelphia, Saunders, Elsevier, 2007
5. Enomoto A, Niwa T: Roles of organic anion transporters in the progression of chronic renal failure. *Ther Apher Dial*, 11[Suppl 1]: S27–S31, 2007
6. Mikkaichi T, Suzuki T, Onogawa T, Tanemoto M, Mizutamari H, Okada M, Chaki T, Masuda S, Tokui T, Eto N, Abe M, Satoh F, Unno M, Hishinuma T, Inui K, Ito S, Goto J, Abe T: Isolation and characterization of a digoxin transporter and its rat homologue expressed in the kidney. *Proc Natl Acad Sci U S A*, 101: 3569–3574, 2004
7. Gross ML, Ritz E: Hypertrophy and fibrosis in the cardiomyopathy of uremia—Beyond coronary heart disease. *Semin Dial*, 21: 308–318, 2008
8. Piecha G, Adamczak M, Ritz E: Dyslipidemia in chronic kidney disease: Pathogenesis and intervention. *Pol Arch Med Wewn*, 119: 487–492, 2009

See related article, “*SLCO4C1* Transporter Eliminates Uremic Toxins and Attenuates Hypertension and Renal Inflammation,” on pages 2546–2555.