Reactive Oxygen Species Promote Caspase-12 Expression and Tubular Apoptosis in Diabetic Nephropathy

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ABSTRACT
Apoptosis of tubular epithelial cells contributes to the tubular atrophy that accompanies diabetic nephropathy. Reactive oxygen species (ROS) promote tubular apoptosis, but the mechanisms by which this occurs are incompletely understood. Here, we sought proapoptotic genes that ROS differentially upregulate in renal proximal tubular cells of diabetic (db/db) mice. We performed microarray analysis using total RNA from freshly isolated renal proximal tubules of nondiabetic, diabetic, and diabetic transgenic mice overexpressing catalase in the proximal tubule (thereby attenuating ROS). We observed greater expression of caspase-12 in the proximal tubules of the diabetic mice compared with the nondiabetic and diabetic transgenic mice. Quantitative PCR and immunohistochemistry confirmed the enhanced expression of caspase-12, as well as members of the endoplasmic reticulum stress–induced apoptotic pathway.

Ex vivo, albumin induced caspase-12 activity and expression (protein and mRNA) and mRNA expression of the CCAT/enhancer-binding protein homologous protein in freshly isolated wild-type proximal tubules but not in catalase-overexpressing proximal tubules. In vitro, albumin stimulated activity of both caspase-12 and caspase-3 as well as expression of caspase-12 and CCAT/enhancer-binding protein homologous protein in a human proximal tubule cell line (HK-2). The free radical scavenger tiron inhibited these effects. Furthermore, knockdown of caspase-12 with small interfering RNA reduced albumin-induced apoptosis in HK-2 cells. Taken together, these studies demonstrate that albuminuria may induce tubular apoptosis through generation of ROS and the subsequent expression and activation of endoplasmic reticulum stress genes in the diabetic kidney.


Diabetic nephropathy (DN) is the leading cause of ESRD.1–5 Although glomerular lesions are central in initiating kidney damage, studies within the past three decades have demonstrated that tubulointerstitial fibrosis and tubular atrophy play a key role in nephropathy progression, leading to ESRD.6–12

The underlying mechanism(s) of tubular atrophy in diabetes, however, are incompletely delineated. One attractive mechanism is apoptosis, which has been demonstrated to mediate cell death in a variety of renal diseases, including DN.13–16 Indeed, apoptosis was detected in renal proximal tubular cells (RPTCs) of mice,13,17 rats,18 and patients

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with diabetes, suggesting that tubular apoptosis may precede tubular atrophy in diabetes.

Reactive oxygen species (ROS) mediate renal cell apoptosis induced by hyperglycemia, angiotensin II, and albumin. High glucose induced ROS generation and stimulated renin-angiotensin system gene expression in RPTCs. Furthermore, transgenic (Tg) mice overexpressing rat catalase (CAT) in their RPTCs exhibited attenuated ROS generation, hypertension, and tubular apoptosis in diabetic kidneys in vivo, supporting an important role for ROS in tubular apoptosis in diabetes.

This study aimed to identify proapoptotic genes that are differentially upregulated by ROS in RPTCs of type 2 diabetic db/db mice by using DNA chip microarray. Caspase-12, an endoplasmic reticulum (ER) stress–induced proapoptotic gene, was differentially upregulated in RPTCs of db/db mice. Increased caspase-12 expression in RPTCs was confirmed by immunohistochemistry and real-time quantitative-PCR (RT-qPCR). Finally, we also demonstrated that albumin induced caspase-12 expression and RPTC apoptosis, and its action was mediated, at least in part, via ROS generation.

RESULTS

Physiologic Parameters in Tg Mice

Table 1. Physiologic parameters of mice at age 12 weeks

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Body Weight (g)</th>
<th>Blood Glycemia (mM)</th>
<th>SBP (mmHg)</th>
<th>ACR (µg/mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>db/m+</td>
<td>24.5 ± 1.1</td>
<td>12.8 ± 2.3</td>
<td>115 ± 5</td>
<td>1.73 ± 1.14</td>
</tr>
<tr>
<td>CAT-Tg</td>
<td>26.9 ± 3.7</td>
<td>13.7 ± 2.2</td>
<td>106 ± 5</td>
<td>0.30 ± 0.27</td>
</tr>
<tr>
<td>db/db</td>
<td>47.5 ± 2.2</td>
<td>28.6 ± 5.0</td>
<td>134 ± 9b</td>
<td>1.35 ± 0.44</td>
</tr>
<tr>
<td>db/db CAT-Tg</td>
<td>49.7 ± 3.2</td>
<td>29.6 ± 3.0</td>
<td>111 ± 8c</td>
<td>3.03 ± 1.61</td>
</tr>
</tbody>
</table>

*aP < 0.001.

Table 2. Physiologic parameters of mice at age 20 weeks

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Body Weight (g)</th>
<th>Blood Glycemia (mM)</th>
<th>Kidney Weight (g)</th>
<th>Kidney/Body Weight (g/g)</th>
<th>SBP (mmHg)</th>
<th>ACR (µg/mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>db/m+</td>
<td>29.2 ± 1.4</td>
<td>11.5 ± 1.0</td>
<td>0.330 ± 0.030</td>
<td>0.0100 ± 0.0010</td>
<td>108 ± 8</td>
<td>0.98 ± 0.71</td>
</tr>
<tr>
<td>CAT-Tg</td>
<td>32.1 ± 4.8</td>
<td>12.7 ± 1.5</td>
<td>0.430 ± 0.050</td>
<td>0.0130 ± 0.0047</td>
<td>112 ± 7</td>
<td>1.09 ± 0.23</td>
</tr>
<tr>
<td>db/db</td>
<td>56.4 ± 2.0a</td>
<td>27.1 ± 4.6b</td>
<td>0.450 ± 0.035</td>
<td>0.0080 ± 0.006</td>
<td>131 ± 8c</td>
<td>16.00 ± 3.71a</td>
</tr>
<tr>
<td>db/db CAT-Tg</td>
<td>59.0 ± 2.9a</td>
<td>18.8 ± 5.2c</td>
<td>0.460 ± 0.078</td>
<td>0.0078 ± 0.0015</td>
<td>118 ± 1d</td>
<td>10.00 ± 1.23ac</td>
</tr>
</tbody>
</table>

*aP < 0.001.

Table 3. Gene ontology

<table>
<thead>
<tr>
<th>Gene Title</th>
<th>Abbreviation</th>
<th>db/m+ versus db/db</th>
<th>db/db versus db/db CAT-Tg</th>
<th>Average Expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>Baculoviral IAP repeat-containing 4</td>
<td>Birc4</td>
<td>−0.358 0.004</td>
<td>0.368 0.004</td>
<td>2.787</td>
</tr>
<tr>
<td>Bcl2 modifying factor</td>
<td>Bmf</td>
<td>−1.617 0.010</td>
<td>1.617 0.010</td>
<td>4.953</td>
</tr>
<tr>
<td>Caspase-12</td>
<td>Casp12</td>
<td>−0.865 0.007</td>
<td>0.857 0.007</td>
<td>3.221</td>
</tr>
<tr>
<td>E2F transcription factor 1</td>
<td>E2f1</td>
<td>−0.249 0.007</td>
<td>0.246 0.006</td>
<td>2.939</td>
</tr>
<tr>
<td>Neurotrophic tyrosine kinase, receptor, type 2</td>
<td>Ntrk2</td>
<td>−0.606 0.002</td>
<td>0.574 0.002</td>
<td>2.717</td>
</tr>
<tr>
<td>Purinergic receptor P2X, ligand-gated ion channel, 1</td>
<td>P2rx1</td>
<td>−1.958 0.011</td>
<td>2.303 0.008</td>
<td>2.939</td>
</tr>
<tr>
<td>Tial 1 cytotoxic granule–associated RNA binding protein-like 1</td>
<td>Tial1</td>
<td>−0.481 0.011</td>
<td>0.638 0.007</td>
<td>6.886</td>
</tr>
<tr>
<td>Tnf receptor–associated factor 1</td>
<td>Traf1</td>
<td>−0.936 0.009</td>
<td>0.949 0.009</td>
<td>2.818</td>
</tr>
</tbody>
</table>

*P < 0.05 versus db/m+.

**P < 0.05 versus db/db.

with diabetes, suggesting that tubular apoptosis may precede tubular atrophy in diabetes.
and db/db CAT-Tg mice (Table 1). SBP, body weight, blood glucose level, and ACR were significantly higher in 20-week-old male db/db mice than in db/m+ mice (Table 2). Conversely, db/db CAT-Tg mice had higher body weight and blood glucose level than db/m+ mice but exhibited normal SBP and attenuated ACR as compared with db/db mice (Tables 1 and 2). These results indicate that CAT overexpression in RPTCs effectively prevented hypertension and attenuated albuminuria in db/db mice.

Gene Chip Microarray Analysis
Data were normalized from the various probe sets on the chips for analysis. Gene Ontology database was used to screen for the probe sets involved in the apoptotic processes and that resulted in a list of eight upregulated apoptotic genes in db/db mice, compared with db/m+ and db/db CAT-Tg mice based on a value of $P < 0.01$ (Table 3).

Validation of Caspase-12 Expression in Mouse Kidneys by RT-PCR and Immunohistochemistry
Conventional RT-PCR and RT-qPCR for mouse caspase-12 mRNA expression were used to validate DNA microarray findings (Figure 1, A and B, respectively). Baseline expression of caspase-12 mRNA in 20-week-old db/db mice was threefold higher as compared with db/m+ by RT-qPCR (Figure 1B). This increase was significantly ($P < 0.05$) attenuated in db/db CAT-Tg mice. Immunoblotting of caspase-12 confirms the upregulation of caspase-12 expression in 20-week-old db/db mice as compared with db/m+ but normalized in db/db CAT-Tg mice (Figure 1C).

Immunohistochemistry revealed caspase-12 expression in the distal tubules but in very few renal proximal tubules (RPTs) of db/m+ mouse kidneys (Figure 1D, a). RPTs near the glomeruli (proximal convoluted tubules) stained negative for caspase-12, whereas proximal straight tubules stained positive (Figure 1D, d); however, increased immunostaining for caspase-12 in the convoluted segment of RPTs was apparent in diabetic db/db mice (Figure 1D, b and e), compared with nondiabetic db/m+ controls (Figure 1D, a and d). CAT overexpression effectively attenuated caspase-12 expression in RPTs of db/db CAT-Tg mice (Figure 1D, c and f).

Apoptotic RPTs Express Caspase-12
Immunofluorescence analysis of caspase-12 expression and terminal transferase-mediated deoxyuridine triphosphate nick end-labeling (TUNEL) assay in 20-week-old mouse kidneys revealed that db/db mice (Figure 2b) had a much greater number of apoptotic cells than db/m+ (Figure 2a) and db/db CAT-Tg mice (Figure 2c). Magnification at ×600 in db/db mice demonstrated that RPTs overexpressing caspase-12 contained TUNEL-positive apoptotic cells (Figure 2d).

Caspase-12 Expression Is ER Stress Dependent
Because ER stress induces caspase-12 expression, we investigated the expression of ER chaperone 78-kD glucose-regulated protein/lG heavy chain–binding protein (GRP78/
Albumin Induces Caspase-12 and CHOP but not GRP78/BiP Expression in 12-Week-Old Mice

High concentrations of albumin have been implicated in the induction of caspase-12 expression and apoptosis in RPTCs in vitro. To investigate the correlation of albuminuria with caspase-12, GRP78/BiP, and CHOP expression in vivo in RPTs, we examined caspase-12, GRP78/BiP, and CHOP protein and mRNA expression in RPTs as well as in kidney sections of 12-week-old animals. Immunoblotting (Figure 4, A and B) and RT-qPCR (Figure 4C) as well as immunostaining (Figure 4D) showed no significant differences in caspase-12 protein and mRNA expression in db/m+ db/db, and db/db CAT-Tg mice. Likewise, no significant differences could be detected in GRP78/BiP and CHOP mRNA expression (Figure 5, A and B) and protein expression (Figure 5, C and D) and GRP78/BiP expression (Figure 5E) in 12-week-old mice. Thus, the differences in caspase-12, CHOP, and GRP78/BiP expression observed at 20 weeks of age were likely due to increased ACR in the db/db and db/db Cat-Tg mice compared with db/m+ and Cat-Tg mice (Tables 1 and 2).

Albumin Induces ROS Generation and Caspase-12 and CHOP Expression Ex Vivo

To ascertain ROS dependence of caspase-12 and CHOP up-regulation, we performed ex vivo studies on RPTs freshly isolated from adult male wild-type (wt) mice (C57BL/6J) and CAT-Tg mice (C57BL/6J). RPTs were incubated with 0, 30, and 60 μg/ml fat-free BSA in serum-free medium for 16 hours. Albumin (60 μg/ml) induced ROS generation (Figure 6A) and increased caspase-12 activity (Figure 6B) in RPTs of wt mice but not of RPTs of CAT-Tg mice. Likewise, albumin stimulated caspase-12 and CHOP mRNA expression in RPTs of wt mice without inducing significant changes in caspase-12 and CHOP mRNA expression in RPTs of Cat-Tg mice (Figure 7, A, B, and D). Albumin also induced caspase-12 protein expression in RPTs of wt mice but had no effect in RPTs of Cat-Tg mice (Figure 7E). GRP78/BiP mRNA and protein expression did not differ in RPTs of wt and CAT-Tg mice in response to albumin (Figure 7, A, C, and F).

Albumin Induces ROS Generation and Caspase-12 Expression in HK-2 Cells In Vitro

Next, we used a human proximal tubular cell line (HK-2) to confirm albumin induction of caspase-12 expression. HK-2 cells were incubated with 0, 30, and 60 μg/ml BSA in serum-free medium for 24 hours. BSA at 30 μg/ml enhanced ROS generation (Figure 8A) but not caspase-12 activity (Figure 8B). Furthermore, increases in caspase-12 protein (combined both pro–caspase-12 (55 kD) and cleaved pro–caspase-12 (45 kD) expression level did not reach statistical significance (Figure 8C). At 60 μg/ml, BSA induced significantly higher ROS generation (Figure 8A) and enhanced caspase-12 activity (Figure 8B) and caspase-12 protein expression (Figure 8, C and D). Tiron effectively blocked the stimulatory effects of BSA on...
ROS generation, caspase-12 activity, and caspase-12 cleavage but had no effect on pro-caspase-12 expression (Figure 8, A through C). Culturing HK-2 cells in high-glucose medium also augmented total caspase-12 expression (Figure 8D).

**Albumin Initiates Caspase-3 Activation and Apoptosis in HK-2 cells**

Next, we investigated whether albumin could induce caspase-3 activation and apoptosis in HK-2 cells. Incubation of HK-2 cells with BSA at 60 μg/ml but not at 30 μg/ml in serum-free medium for 24 hours stimulated caspase-3 activity (Figure 9A), as well as poly-(ADP-ribose)polymerase (PARP) expression and cleavage (Figure 9B). Furthermore, albumin enhanced the number of apoptotic HK-2 cells in normal and high-glucose medium (Figure 9C, b and d) compared with untreated cells (Figure 9C, a and c). Interestingly, high glucose (25 mM) alone did not increase significantly the number of apoptotic cells in HK-2 cells (Figure 9D).

Finally, we investigated whether specific knockdown of caspase-12 expression would reduce the proapoptotic effect of albumin. Transfection of HK-2 cells with caspase-12 small interfering RNA (siRNA) but not a scrambled siRNA effectively reduced caspase-12 expression assessed as both the 55- and 42-kD bands in a dosage-dependent manner (Figure 10A). Incubation of HK-2 cells with 60 μg/ml BSA increased the number of apoptotic cells compared with HK-2 cells cultured with 40 pmol of scrambled siRNA alone in 5 or 25 mM d-glucose medium (Figure 10, B and C, b and e). By contrast, caspase-12 siRNA reduced the number of apoptotic HK-2 cells incubated with 60 μg/ml BSA (Figure 10, B and C, c and f). Of note, culture of HK-2 cells in high-glucose medium (25 mM d-glucose) did not significantly increase the number of apoptotic cells as compared with those cultured in normal glucose (Figure 10, B and C, a and d).

**DISCUSSION**

This study demonstrated increased caspase-12, GRP78/BiP, and CHOP expression in RPTCs of diabetic db/db mice. The kidneys of diabetic db/db mice also indicated a higher degree of apoptosis than db/m+ and db/db Cat-Tg mice, which co-localized with areas of increased caspase-12 expression. Albumin-induced ROS generation and stimulated caspase-12, GRP78/BiP, and CHOP expression in RPTCs of non-diabetic mice ex vivo and in HK-2 cells in vitro but not in RPTCs of Cat-Tg mice ex vivo. Furthermore, albumin increased the number of apoptotic HK-2 cells in vitro, and this was abrogated by treatment with caspase-12 siRNA. These findings indicate an important role for ROS in mediating albumin-induced RPTC apoptosis via ER stress and proapoptotic gene expression.

Caspase-12 immunostaining was markedly augmented in RPTCs of db/db mice with albuminuria at 20 weeks of age. This increase in caspase-12 expression in RPTCs was initially found by gene chip microarray analysis and subsequently validated by RT-qPCR. The increase in caspase-12 expres-
tion in RPTs coincided with the development of albuminuria in db/db mice. Thus, at 12 weeks of age, when db/db mice had no apparent albuminuria, caspase-12 expression in their RPTs seemed to be similar to that of db/m+ mice. In contrast, development of macroalbuminuria at the age of 20 weeks coincided with increased caspase-12 expression in the RPTs. Intriguingly, caspase-12 expression was dramatically decreased in RPTs of 20-week-old db/db CAT-Tg mice, suggesting that caspase-12 expression was mediated, at least in part, via ROS generation. Immunofluorescence staining for caspase-12 and the TUNEL assay in 20-week-old mice revealed that the kidneys of db/db mice contained higher numbers of apoptotic cells that also expressed caspase-12, suggesting a causal link between caspase-12 expression and RPTC apoptosis.

ER stress–mediated activation of caspase-12 suggests a role for caspase-12 in RPTC apoptosis, because it directly cleaves procaspase-9, leading to caspase-3 activation independent of the intrinsic (mitochondrial) pathway. Mice deficient in caspase-12 develop normally, and cells derived from caspase-12 null embryos are usually resistant to pharmacologic induction of ER stress, although some types of cells could undergo ER-induced apoptosis in the absence of caspase-12. Thus, the importance of caspase-12–mediated apoptosis in vivo remains unclear but could be cell specific; however, it is very likely that caspase-12 activation (mitochondrial independent) could act in concert with the mitochondrial pathway to promote RPTC apoptosis in diabetes. Indeed, our previous study demonstrated increased Bax expression in RPTCs of db/db mice.

To date, several pathways of ER stress have been reported (reviewed by Zhang and Kaufman). GRP78/BiP, a molecular chaperone localized in the ER, is involved in the folding and translocation of nascent proteins. CHOP is a proapoptotic gene activated by ER stress. We detected increases in GRP78/BiP and CHOP mRNA expression in the RPTs of 20-week-old db/db mice, indicating that caspase-12 upregulation in db/db mice was ER stress dependent. We also detected increased GRP78/BiP protein expression but not CHOP in the RPTs of 20-week-old db/db mice compared with db/m+ and db/db Cat-Tg mice (unpublished results, Supplemental Figure A). Although the reasons for undetectable CHOP expression by immunoblotting are not clear, it is possible that either the expression of endogenous mouse CHOP was below the detection limit of our assay, or the anti-CHOP antibody that works well in immunohistochemistry does not work in immunoblotting. Furthermore, increases in GRP78/BiP and CHOP expression coincided with the onset of albuminuria. Twelve-week-old db/db mice did not exhibit increased expression compared with db/m+ and db/db Cat-Tg, whereas 20-week-old animals did show augmented expression corresponding with an elevated ACR.

To explore the underlying mechanism(s) of albumin-induced caspase-12, GRP78/BiP, and CHOP expression, we studied mouse RPTs (mRPTs) freshly isolated from wt and CAT-Tg mice. We observed that albumin enhanced ROS production and caspase-12 activity and caspase-12 mRNA and protein expression and CHOP mRNA expression in freshly isolated RPTs from wt mice. No such enhancement or activation was noted, however, in RPTs from CAT-Tg mice. Thus, albumin stimulation of caspase-12 activation seems to be ROS dependent. Indeed, albumin induced ROS generation in RPTs. Thus, these findings support the notion that stimulation of caspase-12 and ER stress gene expression and activation by albumin is mediated, at least in part, via ROS generation in RPTs. Surprising, we did not observe an increase of GRP78/BiP protein and mRNA expression by albumin ex vivo. By contrast,
previous studies reported that high levels of albumin induced GRP78/BiP expression in RPTCs in vitro. Of note, we used BSA at the concentration range of 10 to 100 μg/ml, which were detected in the ultrafiltrate by Lindenmeyer et al., whereas other studies used albumin at 100- to 1000-fold higher concentrations (i.e., 10 to 30 mg/ml). Our results with mouse kidneys were further supported by in vitro studies on HK-2 cells. Intriguing, albumin also enhanced caspase-12 expression in HK-2 cells via ROS generation. Previous bioinformatics analyses predicted that a silencing stop codon is inserted in the caspase-12 sequence in 98% of the population, yielding a truncated caspase-12 (i.e., inactive caspase-12). Positive selection for the mutation inserting the stop codon presumably occurred more than 100,000 years ago and may have an advantage for survival because caspase-12 also functions as an inhibitor of caspase-1 to impair the inflammasome. Thus, the presence of the full-length caspase-12 (a 52- to 54-kD) protein might sensitize humans to sepsis, a disadvantage for survival. Recent findings suggest that full-length caspase-12 could be re-expressed in human tissues under certain pathologic conditions, including cancer. Intriguingly, our sequencing confirmed presence of a silencing stop codon in caspase-12; however, a full-length 55-kD caspase-12 protein continued to be expressed in our studies. Furthermore, caspase-12 siRNA silenced expression of both the 55- and 42-kD caspase-12 with concomitant reduction in the number of apoptotic cells. These findings strongly suggest that human caspase-12 may have a functional role in some cell types despite the notion that it may be a pseudogene lacking activity.

Cell-free in vitro transcription/translation experiments demonstrated that a single-nucleotide polymorphism (T125C) in human caspase-12 is translated into a 38-kD human caspase-12 protein but that this does not occur when there is a stop codon (TGA) in human caspase-12 at amino acid position 125. Other studies documented expression of a 50-kD
caspase-12 in human promyelocytic leukemia cells (HL-60) and a 60-kD caspase-12 in human gastrointestinal stromal tumors (STI571). It is not clear how a 52- or 60-kD caspase-12 protein can be expressed in the presence of a stop codon; however, these studies raise the possibility that the 52- or 60-kD pro–caspase-12 could be aberrantly expressed in cells undergoing pathologic changes.

Future studies are required to investigate whether human caspase-12 is glycosylated in HK-2 or cancer cells. Our culture media contained 100 μg/ml streptomycin, an aminoglycoside. Because aminoglycosides permit translational read-through of the stop codon in many genes (reviewed by Linde and Kerem), it remains to be investigated whether streptomycin might have promoted translational read-through of the stop codon in human caspase-12 gene.

Studies of rodent and human RPTCs reported albumin-induced caspase-12 activation and consequently RPTC apoptosis. In HK-2 cells, we also observed albumin-induced caspase-12 activation, caspase-3 activation, PARP cleavage, and apoptosis. Moreover, specific knockdown of caspase-12 with siRNA markedly reduced apoptosis in response to albumin. These findings lend further support to the hypothesis that elevated tubular fluid albumin levels could induce RPTC apoptosis via caspase-12 expression. Surprising, high glucose (25 mM) enhanced caspase-12 expression without inducing HK-2 cell apoptosis. A possible explanation is that high glucose enhances expression of caspase-12 expression and activity that did not lead to caspase-3 activation as suggested by our unpublished data (Supplemental Figure B).

Our findings may have important clinical implications. Because tubular apoptosis is detectable in various renal diseases with proteinuria, including DN, and tubular atrophy seems to be a better indicator of disease progression than glomerular pathology, we postulate that albuminuria-induced RPTC apoptosis may be an initial step leading to tubular atrophy and that ROS is one of the key mediators of this process. ROS may induce ER stress in RPTCs and stimulate caspase-12, GRP78/Bip, and CHOP expression and activation, triggering the initiation and amplification of the apoptotic cascade leading to tubular apoptosis.

In summary, our results indicate an important role for ROS and ER stress in albuminuria-induced RPTC apoptosis and suggest that caspase-12 activation may contribute to RPTC apoptosis and nephropathy progression in diabetes.

### Concise Methods

**Chemicals and Constructs**

- Glucose, mannitol, lipid-free BSA, tiron, and mAbs against β-actin were purchased from Sigma-Aldrich Canada Ltd. (Oakville, Ontario, Canada).
- Normal glucose (5 mM D-glucose), DMEM, and FBS were bought from Invitrogen (Burlington, Ontario, Canada).
- Polyclonal anti-GRP78/BiP, anti-PARP, and anti-CHOP antibodies were obtained from Santa Cruz Biotechnology (Santa Cruz, CA).
- Polyclonal anti-human/mouse caspase-12 antibody was procured from Cell Signaling (New England Biolabs Ltd., Pickering, Ontario, Canada) and (eBioscience, Cedarlane Laboratories, Hornby, Ontario, Canada).
- Oligonucleotides were synthesized by Invitrogen.

**Generation of db/db CAT-Tg Mice**

We have reported the generation of db/db CAT-Tg mice overexpressing rat CAT in their RPTCs. Non-Tg, gender-matched db/m+ littermates (controls), db/db CAT-Tg mice, and db/db mice were studied at 12 and 20 weeks of age. All animals received standard mouse chow and water ad libitum. Animal care met the standards set forth by the Canadian Council on Animal Care, and all procedures were approved by the Animal Care Committee of the Centre hospitalier de l’Université de Montréal.
Physiologic Parameters
BP was monitored with a BP-2000 tail-cuff pressure instrument (Visitech Systems, Apex, NC) every 2 weeks for a period of 20 weeks, starting at the age of 8 weeks as described previously. Blood glucose level was quantified weekly with an ACCU-CHEK Compact Plus glucose meter (Roche Diagnostics, Laval, Que ´bec, Canada). The mice were housed in metabolic cages to obtain 24-hour urine samples for the assessment of albuminuria by ELISA (Albuwell and Creatinine companion; Exocell, Philadelphia, PA).

Mouse RPT Isolation and DNA Microarray Analysis
Animals were killed at 12 and 20 weeks of age, and kidneys were removed immediately. The left kidney was used for histology and immunohistochemistry. The right kidney was used for proximal tubule isolation by Percoll gradient as described previously. Aliquots of freshly isolated mRPTs from individual animals (three mice in each group) at 20 weeks of age were immediately used for proximal tubule isolation and subjected to Affymetrix Mouse Genome 430 2.0 microarray chips analysis (Microarray Centre of the Centre de recherche du Centre hospitalier de l’Université de Montréal). Data were analyzed on a computer with the R statistical language (version 2.51). The affylmGUI (version 1.10.5) software and the LIMMA package (version 2.10.5) of the Bioconductor library (release 2.0) were used for analysis.

PCR Assays for Gene Expression
Total RNA was used in RT-PCR and RT-qPCR to quantify the amount of caspase-12, GRP78/BiP, and CHOP mRNA expressed in mRPTs and human RPTCs (HK-2).

Immunohistochemistry
Immunostaining for mouse caspase-12, GRP78/BiP, and CHOP was performed by standard avidin-biotin-peroxidase complex method (ABC Staining System; Santa Cruz Biotechnology). Slides from six animals per group were analyzed visually under a light microscope by two investigators who were unaware of the treatments.

Cell Culture
HK-2 cells were cultured as described elsewhere. Briefly, cells were cultured in 5 mM D-glucose DMEM containing 5% FBS. For experiments, cells were synchronized for 24 hours in serum-free 5 mM D-glucose DMEM at 60 to 70% confluence. Then, cells were cultured in serum-free 5 mM D-glucose DMEM plus 20 mM D-mannitol or 25 mM D-glucose in the absence or presence of 60 µg/ml albumin. *P < 0.05; **P < 0.01; ***P < 0.005.

In Vivo Fluorescence Staining
Kidney sections of 20-week-old animals were incubated initially with the TUNEL fluorescein kit (Roche Diagnostics, Indianapolis, IN) as indicated by the manufacturer’s protocol. After the last PBS wash, sections were blocked with goat serum followed by overnight incubation with anti-caspase-12 antibody (eBioscience). Sections were then incubated with anti-rabbit AlexaFluor 555 (Invitrogen), and nuclei were stained with DAPI.

Caspase-12 and Caspase-3 Activity Assay and Immunoblotting
Freshly isolated mRPTs and HK-2 cells were incubated for 24 hours as described previously and then harvested for caspase-12 and caspase-3 activity assay as well as for immunoblotting of caspase-12, PARP, GRP78/BiP, and β-actin mRNA and protein quantification, respectively.

Caspase-12 and Caspase-3 Activity in HK-2 cells
(A) ROS generation. (B) Caspase-12 activity assay. (C) Caspase-12 protein expression assessed by immunoblotting and quantification. (D) Immunoblotting and quantification of caspase-12 protein expression in HK-2 cells in 5 mM D-glucose plus 20 mM D-mannitol or 25 mM D-glucose in the presence or absence of 60 µg/ml albumin. *P < 0.05; **P < 0.01; ***P < 0.005.
Albumin affects caspase-3 activity and apoptosis in HK-2 cells in vitro. HK-2 cells were incubated in 5 mM D-glucose serum-free DMEM in the absence or presence of albumin (0, 30, or 60 μg/ml) with or without tiron (10^{-4} M) for 16 hours. (A) Caspase-3 activity assay. (B) Immunoblotting for PARP expression and cleavage in HK-2 cells in 5 mM D-glucose plus 20 mM D-mannitol or 25 mM D-glucose in the presence or absence of 60 μg/ml albumin. (C) HK-2 cell apoptosis analyzed by TUNEL staining assay. HK-2 cells were incubated in 5 mM D-glucose plus 20 mM D-mannitol medium in the absence (a) or in the presence (b) of 60 μg/ml albumin or in 25 mM D-glucose medium in the absence (c) or presence (d) of 60 μg/ml albumin. Arrows indicate the TUNEL-positive cells. (D) Quantification of TUNEL-positive cells in C. Data are means ± SD; n = 3. ***P < 0.005. Magnification, ×200 in C.

Table 4. Primers used for conventional RT-PCR and RT-qPCR

Mouse caspase-12
- sense N + 778 to N + 797 (5'-GAA GGA ATC TGT GGG GTG AA-3')
- antisense N + 971 to N + 952 (5'-TCA GCA GTG GCT AT CCC TTT-3') (NM 009808)

Mouse CHOP
- sense N + 248 to N + 268 (5'-GTC CCT AGC TTG GCT GAC AGA-3')
- antisense N + 413 to N + 396 (5'-TGG AGA GAG ACC GGT TG-3') (X67083.1)

Mouse GRP78/BiP
- sense N + 1269 to N + 1290 (5'-AAG GTG AAC GAC CCC TAA CAA A-3')
- antisense N + 1404 to N + 1378 (5'-GTC ACT CGG AGA ATA CCA TTA ACA TCT-3') (BC005785.1)

β-actin
- sense N + 155 to N + 179 (5'-ATG CCA TCC TGC GTC TGG ACC TGG C-3')
- antisense N + 115 to N + 139 (5'-AGC ATT TGC GGT GCA CGA TGG AGG G-3') (NM 031144)

or ATAD-AFC (for caspase-12; 15 μg/ml) was added to the mixture. After incubation at 37°C for 1 hour, the amount of AFC liberated from ATAD-AFC was measured using a spectrofluorometer with excitation of 380 nm and emission wavelength of 460 nm. The relative densities of caspase-12 and PARP and their respective cleaved forms and β-actin bands in immunoblotting were quantified by computerized laser densitometry (ImageQuant 5.1 software; Molecular Dynamics).

siRNA Dosage-Dependence Analysis
HK-2 cells were plated at a density of 2.5 × 10^4 in a 12-well plate. The next day, cells were transfected using lipofectamine (Invitrogen) according to the manufacturer’s instructions using 20, 40, and 80 pmol of caspase-12 siRNA (Santa Cruz Biotechnology) or scrambled siRNA (Bio-Rad). Low-glucose medium plus 5% FBS was added after 4 to 6 hours of incubation. Whole-cell lysates were collected for immunoblotting 48 hours after transfection.

TUNEL Assay
HK-2 cells were plated at a density of 1.2 × 10^4 in four-well chamber slides coated with poly-D-lysine (Sigma). The next day, cells were transfected with 40 pmol of siRNA or scrambled siRNA using lipofectamine (Invitrogen). Low-glucose medium plus 5% FBS was placed after 4 to 6 hours of incubation with lipofectamine. Twenty-four hours after transfection, the medium was replaced with serum-free medium containing either 5 mM D-glucose plus 20 mM D-mannitol or 25 mM D-glucose with or without 60 μg/ml albumin and incubated for an additional 24 hours. The chamber slides were then processed for staining with TUNEL fluorescein kit according to the manufacturer’s protocol (Roche Diagnostics, Indianapolis, IN). Nuclei were visualized using DAPI staining. Slides were examined using a fluorescence microscope, and the percentage of TUNEL-positive cells was determined using Image J software (http://rsb.info.nih.gov/ij/).

Sequencing of Genomic DNA of HK-2 Cells
Genomic DNA of HK-2 cells were isolated and subjected to PCR using primers framing a region of 300 bp surrounding the T125C polymorphism (sense 5'-GTCATTCTGTGGTATATTGC-3'; antisense 5'-CCATAATATCATACATCCTTGCTC-3') to amplify the genomic DNA.49 The PCR product was sequenced by Genome Quebec (Montréal, Québec, Canada).

Statistical Analysis
Experimental data were expressed as means ± SD. The data were analyzed by one-way ANOVA using Bonferroni correction. P < 0.05 was considered statistically significant.
Figure 10. Caspase-12 siRNA affects the number of TUNEL-positive apoptotic cells. (A) HK-2 cells were incubated in 5 mM d-glucose medium with increasing dosages of either caspase-12 siRNA or scrambled siRNA. (B) HK-2 cells were incubated in normal-glucose (5 mM d-glucose plus 20 mM d-mannitol) or high-glucose (25 mM d-glucose) medium in the absence or presence of albumin (60 µg/ml) and with or without caspase-12 siRNA or scrambled siRNA (40 pmol). Quantification of TUNEL-positive stained cells. Data are means ± SD. *P < 0.05; **P < 0.01. (C) Apoptosis was assessed by the TUNEL assay. TUNEL-positive apoptotic cells fluoresced green, and DAPI-stained nuclei fluoresced blue. HK-2 cells, plated at a density of 2.5 × 10^4 in 12-well microplates, were incubated in 5 mM d-glucose plus 20 mM d-mannitol in the absence of albumin and presence of 40 pmol of scrambled siRNA (a) or in the presence of 60 µg/ml albumin and 40 pmol of scrambled siRNA (b) or in the presence of 60 µg/ml albumin and 40 pmol of caspase-12 siRNA (c). HK-2 cells were incubated in 25 mM d-glucose medium in the absence of albumin and presence of 40 pmol of scrambled siRNA (d) or in the presence of 60 µg/ml albumin and 40 pmol of scrambled siRNA (e) or in the presence of 60 µg/ml albumin and 40 pmol of caspase-12 siRNA (f). Magnification, ×100 in C.

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DISCLOSURES

None.

REFERENCES


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