AMPK Mediates the Initiation of Kidney Disease Induced by a High-Fat Diet

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ABSTRACT

The mechanisms underlying the association between obesity and progressive renal disease are not well understood. Exposure to a high-fat diet decreases levels of the cellular energy sensor AMPK in many organs, including the kidney, but whether AMPK contributes to the pathophysiology of kidney disease induced by a high-fat diet is unknown. In this study, we randomly assigned C57BL/6J mice to a standard or high-fat diet. After 1 week, mice fed a high-fat diet exhibited an increase in body weight, renal hypertrophy, an increase in urine H₂O₂ and urine MCP-1, and a decrease in circulating adiponectin levels and renal AMPK activity. Urine ACR progressively increased after 4 weeks of a high-fat diet. After 12 weeks, kidneys of mice fed a high-fat diet demonstrated a marked increase in markers of fibrosis and inflammation, and AMPK activity remained significantly suppressed. To determine whether inhibition of AMPK activity explained these renal effects, we administered an AMPK activator along with a high-fat diet for 1 week. Although AMPK activation did not abrogate the weight gain, it reduced the renal hypertrophy, urine H₂O₂, and urine and renal MCP-1. In vitro, AMPK activation completely inhibited the induction of MCP-1 by palmitic acid in mesangial cells. In conclusion, these data suggest that the energy sensor AMPK mediates the early renal effects of a high-fat diet.

Table 1. Characterization of mice fed STD or HFD

<table>
<thead>
<tr>
<th></th>
<th>STD Week 1</th>
<th>HFD Week 1</th>
<th>STD Week 12</th>
<th>HFD Week 12</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight, g</td>
<td>20.89 ± 0.78</td>
<td>25.26 ± 0.94a</td>
<td>27.68 ± 0.80</td>
<td>41.68 ± 0.77b</td>
</tr>
<tr>
<td>Plasma glucose, mg/dl</td>
<td>156.5 ± 8.5</td>
<td>172.3 ± 6.5</td>
<td>147.3 ± 8.3</td>
<td>203.3 ± 12.1b</td>
</tr>
<tr>
<td>Kidney weight, g</td>
<td>0.285 ± 0.01</td>
<td>0.343 ± 0.01a</td>
<td>0.300 ± 0.008</td>
<td>0.410 ± 0.012b</td>
</tr>
<tr>
<td>Kidney weight, g/mm of tibia length</td>
<td>0.0162 ± 0.0007</td>
<td>0.0194 ± 0.0006a</td>
<td>0.0151 ± 0.0005</td>
<td>0.0212 ± 0.0010b</td>
</tr>
<tr>
<td>Adiponectin, <em>μg/ml</em></td>
<td>13.53 ± 1.36</td>
<td>9.75 ± 1.36a</td>
<td>14.14 ± 0.93</td>
<td>17.97 ± 1.98</td>
</tr>
<tr>
<td>Plasma insulin, ng/ml</td>
<td>0.53 ± 0.07</td>
<td>1.03 ± 0.22</td>
<td>0.45 ± 0.02</td>
<td>13.53 ± 0.22b</td>
</tr>
</tbody>
</table>

Values are means ± SEM. _n_ = 5 in each group. Statistical analyses were performed by unpaired _t_ test.

*P ≤ 0.05 versus mice on STD week 1.

bP ≤ 0.05 versus mice on STD week 12.

In a prior study, our group identified that low levels of circulating adiponectin, an adipocyte-specific hormone that is reduced in insulin-resistant obese patients, correlate with low-grade albuminuria in obese African Americans and that the adiponectin knockout mouse developed low-grade albuminuria without co-existent obesity, hypertension, or hyperglycemia. Adiponectin has a protective effect on podocytes primarily via stimulating the enzyme AMP-activated protein kinase (AMPK). AMPK is an ubiquitous heterotrimeric enzyme that is considered to be the master energy sensor in all eukaryotic cells. AMPK has been reported to be reduced in various organs, including the kidney after exposure to a high-fat diet; however, the role of AMPK in mediating HFD-induced kidney disease has not been evaluated.

To define the renal response in relation to obesity we utilized a high-fat diet (HFD) induced obesity model and tested the hypothesis that AMPK is a key pathway by which a HFD contributes to the initial manifestations of kidney disease.

RESULTS

HFD Induces Obesity, Hyperglycemia, Insulin Resistance, and Kidney Hypertrophy

Table 1 illustrates the changes in body weight, blood glucose, kidney weight, plasma insulin, and adiponectin levels in mice fed a HFD compared with mice fed a standard diet (STD) at week 1 and 12 of the experimental protocol. We observed a significant increase in body weight and kidney weight as early as 1 week on the HFD. The level of plasma glucose and insulin were unchanged after 1 week but were significantly higher in HFD mice at week 12. In contrast, the circulating adiponectin level was significantly lower at week 1 in HFD mice, whereas at week 12 circulating adiponectin was similar with HFD and STD.

Effect of HFD on Glomerular Histology

At week 12, the HFD mice demonstrated an overall increase of glomerular area (Figure 1, A, B, and D). The increase in glomerular area was contributed by an accumulation of Periodic-Acid-Schiff (PAS)-positive matrix in the mesangium, an increase of nuclei, and an increase of glomerular filtration surface area (Figure 1D). There was an overall increase in vacuolated cells and some were positively stained by the oil-red O method (Figure 1, B and C). There was a robust increase in glomerular matrix because fibronectin, collagen type I, and collagen type IV were all greatly increased in the glomeruli of mice fed a HFD (Figure 1, F, H, and J). However, no morphologic changes were observed in the kidney after 1 week on a HFD (data not shown).

Urinary Markers of Inflammation Precede Albuminuria with HFD

To determine the time course by which HFD may affect markers of renal inflammation, we examined urine albumin and urine hydrogen peroxide (H2O2) levels in mice at weeks 1, 4, 8, and 12 with HFD compared with STD (Figure 2). As illustrated, the degree of albuminuria in HFD mice increased 3-fold at 4 and 8 weeks and increased to 10-fold at week 12 compared with STD (245.2 ± 13.0 _versus_ 24.4 ± 5.3 μg albumin/mg creatinine). At 1 week, however, the degree of albuminuria was the same in HFD and STD mice (Figure 2A). In contrast, the urinary H2O2 level was significantly higher as early as 1 week after HFD (Figure 2B). The increase in urine H2O2 was maintained and remained significantly increased throughout the 12-week period. In addition, at 1 week, the urine excretion of monocyte chemoattractant protein-1 (MCP-1) was significantly higher in HFD mice compared with the STD mice (Figure 2C).

Effect of HFD on Renal Gene Expression

To evaluate the effect of a HFD on candidate genes involved in renal inflammation and fibrosis, the mRNA expression of proinflammatory and fibrotic markers was measured by real-time PCR. As observed in Table 2, renal cortical mRNA levels of the proinflammatory markers (TNFα, IL-1β, IL-6, MCP-1, macrophage inflammatory protein [MIP]-1α, NADPH oxidase 2 [Nox2], p22phox [neutrophil cytochrome B, 22 kDa polypeptide]) as well as the profibrotic markers (TGF-β1, fibronectin, collagen type I and IV) were all significantly increased after 12 weeks on the HFD. TGF-β1 mRNA expression was the most highly stimulated of the profibrotic markers. The greatest increases of the inflammatory markers were IL-6 and the macrophage chemoattractants (MCP-1 and MIP).

AMPK Activity Is Reduced with HFD

To determine whether renal AMPK activity is affected by the HFD, the degree of AMPKα Thr 172 phosphorylation was performed by immunostaining and confocal analysis, and overall...
AMPK activity was measured by ELISA in the renal cortex (Figure 3). The phosphorylated AMPK (p-AMPK) staining of glomerular sections in the two groups revealed that the HFD caused a marked decrease of p-AMPK in glomeruli compared with the STD at week 12 (Figure 3A through 3C). There was also a quantitative reduction of AMPK activity in the renal cortices of the HFD mice (Figure 3D).

Role of AMPK in the Early Renal Changes of Mice Fed a HFD

On the basis of our time-course studies, it was apparent that markers of inflammation (urine H2O2 and urine MCP-1) preceded development of albuminuria. We hypothesized that AMPK inhibition would occur within days after institution of a high caloric diet and therefore we performed a second study to determine the role of AMPK in the early renal effects of HFD. Mice on the HFD were treated with a specific activator of AMPK, 5-aminoimidazole-4-carboxamide-1-β-D-ribo nucleoside (AICAR) for 1 week. As illustrated in Figure 4, AMPK activity was reduced in the kidney and glomeruli after 1 week of HFD (Figure 4A through 4D) and was increased with AICAR treatment. AICAR treatment did not prevent the increase in body weight with HFD (Table 3). Despite the lack of change in body weight gain, kidney hypertrophy was completely prevented in HFD mice treated with AICAR at week 1 (Table 3). Interestingly, the reduced plasma adiponectin level with HFD was also completely prevented by AICAR treatment. As shown in Table 3, AICAR administration in mice fed a STD did not affect the body weight, kidney weight, or plasma adiponectin level.

AMPK activation prevented the increase in urine H2O2 and urine MCP-1 with HFD (Figure 5, A and B). The increased macrophage infiltration observed in mice fed a HFD (Figure 5, C and D) was prevented by the AICAR treatment (Figure 5E). Of the genes that were stimulated with prolonged HFD, only MCP-1 gene expression was significantly increased in mice fed HFD for 1 week (Table 4). Localization of MCP-1 in glomeruli was assessed using confocal analysis. MCP-1 protein was increased in the glomeruli after 1 week of HFD and reduced with AICAR (Figure 6A through 6D). Of note, colocalization of MCP-1 with the podocyte marker podocin (Figure 6A through 6C) was less than 30% (0.29 ± 0.06 for STD mice; 0.21 ± 0.04 for HFD mice, and 0.24 ± 0.08 for HFD + AICAR mice, P = NS). To determine whether MCP-1 was colocalized...
AMPK Activation Prevents MCP-1 Production in Mesangial Cells

To determine if AMPK mediates high-fat induced MCP-1 production, mesangial cells in culture were stimulated with palmitic acid (PA) and MCP-1 was measured in the conditioned media at 4, 8, 16, 24, and 40 hours after stimulation (Figure 7). Mesangial cells exhibited a dramatic increase in MCP-1 expression with PA treatment in a dose- and time-dependent pattern. The peak effect of MCP-1 production was demonstrated between 24 and 40 hours. At each time point, treatment with AICAR completely prevented the PA-stimulated production of MCP-1.

DISCUSSION

Obesity is a major risk factor for renal disease progression and can cause de novo glomerulopathy. Obesity-related renal disease has been characterized by proteinuria, glomerulomegaly, and glomerulosclerosis. Increased body mass index and high fat intake have been associated with onset and progression of renal disease.6,15 In this study we show that high fat feeding has dramatic effects in a time-dependent manner. Even after just 1-week exposure to HFD, there was an increase in body weight, a reduction of plasma adiponectin, and renal inflammation. The early renal inflammatory response was limited to elevated levels of MCP-1 and urinary H2O2 and preceded the progressive increase in urine albumin. AMPK activity was decreased within 1 week of high fat exposure, and AMPK activation with AICAR was able to completely block the early onset of the renal inflammatory response to HFD. Taken together, the study likely has relevance to features of the human metabolic syndrome and provides insight into the mechanisms underlying obesity-related low-grade renal inflammation and albuminuria.16

AMPK is a ubiquitous heterotrimeric kinase consisting of a catalytic α subunit and regulatory β and γ subunits, and it is abundantly expressed in the kidney.17,18 As a cellular energy sensor, its activity is highly linked to the change in the intracellular...
AMPK activity whereas an increase of ATP level results in its inhibition. Hence, its activation results in a change of energy utilization involving the stimulation of energy-producing pathways to restore energy balance. Metabolic stress conditions such as diabetes or obesity modulate the activity of AMPK. The mechanism of AMPK reduction in renal cells exposed to HFD has not been evaluated, but it could be due to a change in AMP/ATP ratios or other upstream modulators of AMPK activity, such as LKB-1 (liver kinase B-1), and CaMKK (calcium calmodulin-dependent protein kinase kinase). Renal AMPK activity has previously been reported to be reduced after a HFD for 16 weeks; however, its role in mediating renal inflammation was not evaluated. We had previously found that AMPK activity was reduced in the glomeruli in adiponectin knockout mice and that glomerular AMPK was stimulated by adiponectin. In the study presented here, we identified that renal AMPK activity was reduced in association with adiponectin reduction during the early stage of HFD-induced kidney disease; however, AMPK reduction persisted despite an increase in adiponectin during the chronic stage of HFD-induced kidney disease. It is possible that the initial increase in visceral adiposity and inflammation contributes to the reduced adiponectin level, but the mouse is able to compensate and increases adiponectin levels with chronic HFD.

To demonstrate the functional role of AMPK, AICAR was administered with HFD concurrently for 1 week. AICAR is taken up into the cells where it is phosphorylated to 5-aminimidazole-4-carboxamide-1-β-D-furanosyl 5’-monophosphate and mimics the effect of AMP to activate AMPK. AICAR administration increased AMPK activity in the kidney and reduced renal hypertrophy despite the persistent increase in body weight with HFD. Moreover, the early reduction in plasma adiponectin level with HFD was completely prevented by AICAR treatment, demonstrating a potential role of AMPK on adipocyte production or release of adiponectin. In support of our in vivo observation, a prior study found that AICAR was able to directly increase gene expression of adiponectin in human adipocytes in vitro. Nevertheless, the beneficial effects of AMPK activation are likely independent of adiponectin because we found that AICAR treatment of mice with adiponectin deficiency conferred a renoprotective effect associated with stimulation of glomerular AMPK.

An important and early consequence of HFD-induced AMPK inhibition in the kidney is upregulation of MCP-1. Renal MCP-1 was increased as early as 1 week after HFD at the gene expression level, in the glomerulus by immunostaining, and in the urine. Interestingly, of the proinflammatory markers measured, only MCP-1 was increased at the early stage of HFD-induced kidney disease. The role of MCP-1 and its receptor, chemokine (C-C motif) receptor 2 (CCR2), has received greater recognition for its role in mediating chronic kidney disease. Recently, MCP-1 was found to regulate nephrin expression via chemokine receptor 2 in human podocytes, and mice lacking MCP-1 had resistance to diabetes-induced albuminuria. The role of MCP-1 as a mediator of insulin resistance has also been considered, although there are conflicting reports whether MCP-1 deficiency protects against insulin resistance with a HFD.

The major cellular source of glomerular MCP-1 with HFD remains unclear. Although podocytes are a source of MCP-1, our results suggest that mesangial and glomerular endothelial cells are also an important source of MCP-1 production with HFD. Studies in mesangial cells demonstrate a marked stimulation of MCP-1 secretion by PA, suggesting that exposure of circulating saturated fatty acids, such as PA, may be a trigger of glomerular MCP-1 production in the setting of HFD and obesity. We showed that the early increase in glomerular MCP-1 would contribute to the subsequent recruitment of macrophages that would then contribute to enhanced release of proinflammatory (TNFα, IL-6, IL-1, Nox2) and profibrotic factors (TGF-β). An additional pathway by which MCP-1 may enhance albuminuria is by a local paracrine effect on podocytes. MCP-1-induced podocyte motility may affect podocyte function and contribute to nephrin reduction.
and subsequent albuminuria. Finally, we demonstrated that AMPK activation is able to inhibit MCP-1 stimulation in vivo with HFD and in vitro in response to palmitic acid. The mechanism by which AMPK activation inhibits MCP-1 in renal cells is unclear, but it may be due to inhibition of nuclear factor-κB activation. AMPK has recently been shown to affect the proteolysis of inhibitor of κB in endothelial cells and regulate nuclear factor-κB.32 Although systemic AMPK activation has multiple beneficial effects to reduce systemic inflammation and improve insulin resistance, there may well be renal-specific responses to AMPK activation that are renoprotective. Hypertension, stimulation of the renin-angiotensin-aldosterone system, and reduced endothelial nitric oxide synthase may all contribute to the renal alterations with HFD; however, these parameters were not evaluated in the study presented here. Of note, systolic BP was increased only after 8 weeks of HFD10 and may therefore may not play a major role in the very early changes noted after 1 week of HFD.

In summary, our studies demonstrate that the kidney is an early responder to the challenge of high fat feeding. The kidney exhibits an initial programmed inflammatory response, initially characterized by increased MCP-1 and urine H2O2, followed by a subsequent robust inflammatory and fibrotic response at the gene expression, histologic, and functional level. The central energy sensor, AMPK, plays a critical role in regulating the initial response in the kidney. Specific AMPK activation in the kidney would therefore be a potential approach to protect the kidney from the early effects of obesity and insulin resistance.

Table 3. Characterization of mice with 1 week of STD, HFD, and STD, HFD + AICAR

<table>
<thead>
<tr>
<th></th>
<th>STD</th>
<th>STD-AICAR</th>
<th>HFD</th>
<th>HFD-AICAR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight, g</td>
<td>21.30</td>
<td>22.50</td>
<td>24.77</td>
<td>23.75</td>
</tr>
<tr>
<td></td>
<td>± 0.70</td>
<td>± 0.35</td>
<td>± 0.54</td>
<td>± 0.52</td>
</tr>
<tr>
<td>Plasma glucose level, mg/dl</td>
<td>175.7</td>
<td>151.3</td>
<td>190.7</td>
<td>178.9</td>
</tr>
<tr>
<td></td>
<td>± 9.6</td>
<td>± 8.4</td>
<td>± 8.6</td>
<td>± 10.5</td>
</tr>
<tr>
<td>Kidney weight, g</td>
<td>0.259</td>
<td>0.2665</td>
<td>0.290</td>
<td>0.265</td>
</tr>
<tr>
<td></td>
<td>± 0.013</td>
<td>± 0.005</td>
<td>± 0.005</td>
<td>± 0.004</td>
</tr>
<tr>
<td>Kidney weight, g/mm of tibia length</td>
<td>0.0138</td>
<td>0.0136</td>
<td>0.0160</td>
<td>0.0138</td>
</tr>
<tr>
<td></td>
<td>± 0.009</td>
<td>± 0.0003</td>
<td>± 0.004</td>
<td>± 0.0003</td>
</tr>
<tr>
<td>Adiponectin, μg/ml</td>
<td>15.12</td>
<td>16.13</td>
<td>8.54</td>
<td>18.45</td>
</tr>
<tr>
<td></td>
<td>± 2.02</td>
<td>± 1.66</td>
<td>± 0.89</td>
<td>± 3.94</td>
</tr>
</tbody>
</table>

Values are means ± SEM. n = 6 in each group. Statistical analyses were performed by one-way ANOVA followed by Newman–Keuls test.

*P ≤ 0.05 versus mice on STD

bP ≤ 0.05 versus mice on STD-AICAR.

cP ≤ 0.05 versus mice on HFD.
Animals
C57BL/6 mice were purchased from Jackson Laboratory (Bar Harbor, ME). All animal procedures were approved by the Institutional Animal Care and Use Committee of the University of California San Diego. Six-week-old mice were fed a HFD (60% of total calories from fat, 20% protein, 20% carbohydrate) (Research Diets, New Brunswick, NJ) or STD (5% fat, 24.5% protein) for 1 and 12 weeks. Mice were placed in metabolic cages for 24-hour urine collection at baseline and after week 1, 4, 8, and 12, and body weight and the blood glucose were also measured at those time points. Mice were sacrificed after 1 or 12 weeks on the diet. Blood samples were collected and portions of kidneys were snap-frozen in liquid nitrogen for RNA and protein isolation. An additional portion was frozen in OCT or fixed in 4% paraformaldehyde for immunostaining.

Animal Study
Mice were treated with AICAR (Toronto Chemicals) with a single daily intraperitoneal dose of 0.5 mg/g body weight for 7 days and then sacrificed. The control animals were given PBS alone. Twenty-four-hour urines from each mouse were collected before treatment and on the last day of the treatment period.

Cell Culture Design
Murine mesangial cells were grown in DMEM (glucose 5.5 mM), 10% heat-inactivated FBS (Atlanta Biologicals, Lawrenceville, GA), 100 U/ml penicillin G, 100 µg/ml streptomycin, and 2 mM L-glutamine (Life Technologies, Invitrogen). One day before treatment...
Figure 6. Increased glomerular MCP-1 with HFD is reduced with AMPK activation. Representative photomicrographs of glomeruli from mice on a (A) STD, (B) HFD, or (C) HFD + AICAR, and (D) semiquantitative analysis of MCP-1-positive staining per glomerular area at week 1. Colocalization of glomerular MCP-1 in mesangial cells. Representative photomicrographs of glomeruli from mice on a (E) STD or (F) HFD at week 12. Megsin (red)/MCP-1 (green); nuclei (blue). Values are means ± SEM. n = 6 in each group. Statistical analyses were performed by one-way ANOVA followed by Newman–Keuls test. *P ≤ 0.05 versus mice on STD; #P ≤ 0.05 versus mice on HFD.
with AICAR, BSA, or PA, cells were plated in 12-well plates (0.2 × 10^6 cells/well) in DMEM (5.5 mM glucose) and 0.5% FBS. PA-BSA was prepared as described previously.34 The next day, cells were treated with AICAR, BSA, or PA as described.34

Urine and Plasma Data
The urine albumin and creatinine were measured with a mouse Albuwell ELISA kit and a Creatinine Companion kit (Exocell). As an index of oxidative stress, urine samples were also analyzed for H_2O_2 by Amplex red assay (Invitrogen) following the manufacturer’s protocol. Plasma insulin and adiponectin levels were determined by ELISA kits (Millipore). The level of MCP-1 was measured with an ELISA kit (BD OptEIA Mouse MCP-1, BD Biosciences) following the manufacturer’s protocol.

p-AMPKα1 Activity
The level of activated AMPK-α1 in renal cortex tissue was determined by a sandwich ELISA to measure AMPK-α1 phosphorylated at T174 (R&D Systems, Minneapolis, MN).

Quantitative Real-Time PCR
Real-time PCR was performed on cortex homogenates as described previously35 using the primers for Nox2, p22phox, TNFα, MCP-1, IL-6, MIP-1α, IL-1β, α1-type I collagen, α1-type IV collagen, fibronectin, TGF-β1, and β-actin as a housekeeping gene.

Morphologic Analysis
Morphometry of sections of kidneys was carried out as previously reported.36 Briefly, 25 randomly selected glomeruli in the outer cortex of each kidney section were evaluated in a blinded manner. An image of each glomerulus was overlaid with grids. Each grid intersection was determined for the following features: capillary lumen, periodic-acid-Schiff-positive mesangial area, and nuclear number. The accumulation of all of the grid intersections was calculated for overall glomerular area.

Immunofluorescence and Oil-Red O Staining
Immunofluorescence was performed with the following primary antibodies: goat polyclonal anti-fibronectin (BD Biosciences), goat anti-type I collagen (SouthernBiotech), goat anti-type IV collagen (SouthernBiotech), and rabbit monoclonal anti-p-AMPKα (Thr172) (Cell Signaling Technology); overlay studies were carried out with the anti-MCP-1 rabbit polyclonal antibody (Abcam) using the podocyte-specific mouse anti-podocin antibody or the mesangial cell-specific marker anti-megsin antibody (Santa Cruz). Slides were observed with a confocal LSM 510 microscope (Zeiss), and representative photographic images were performed at a magnification of 63×. For each antibody, there were at least five mice per group. Neutral lipid accumulation was evaluated on frozen section using Oil-red O staining.

Statistical Analysis
Results are presented as mean values ± SEM. The level for statistical significance was defined as P < 0.05. Analyses were carried out using Graph Pad Prism Software version 4.03. Difference between data groups were evaluated for significance using independent t test of data or one-way ANOVA and Newman–Keuls post hoc test for multiple comparisons.

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DISCLOSURES
None.

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