

# Effects of pH on Potassium: New Explanations for Old Observations

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## ABSTRACT

Maintenance of extracellular K<sup>+</sup> concentration within a narrow range is vital for numerous cell functions, particularly electrical excitability of heart and muscle. Potassium homeostasis during intermittent ingestion of K<sup>+</sup> involves rapid redistribution of K<sup>+</sup> into the intracellular space to minimize increases in extracellular K<sup>+</sup> concentration, and ultimate elimination of the K<sup>+</sup> load by renal excretion. Recent years have seen great progress in identifying the transporters and channels involved in renal and extrarenal K<sup>+</sup> homeostasis. Here we apply these advances in molecular physiology to understand how acid-base disturbances affect serum potassium.

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The effects of acid-base balance on serum potassium are well known.<sup>1</sup> Maintenance of extracellular K<sup>+</sup> concentration within a narrow range is vital for numerous cell functions, particularly electrical excitability of heart and muscle.<sup>2</sup> However, maintenance of normal extracellular K<sup>+</sup> (3.5 to 5 mEq/L) is under two potential threats. First, as illustrated in Figure 1, because some 98% of the total body content of K<sup>+</sup> resides within cells, predominantly skeletal muscle, small acute shifts of intracellular K<sup>+</sup> into or out of the extracellular space can cause severe, even lethal, derangements of extracellular K<sup>+</sup> concentration. As described in Figure 1, many factors in addition to acid-base perturbations modulate internal K<sup>+</sup> distribution including insulin, catecholamines, and hypertonicity.<sup>3,4</sup> Rapid redistribution of K<sup>+</sup> into the intracellular space is essential for minimizing increases in extracellular K<sup>+</sup> concentration during acute K<sup>+</sup> loads. Second, as also illustrated in Figure 1, in steady state the typ-

ical daily K<sup>+</sup> ingestion of about 70 mEq/d would be sufficient to cause large changes in extracellular K<sup>+</sup> were it not for continuous renal K<sup>+</sup> excretion, because K<sup>+</sup> loss from the gastrointestinal tract is quite modest under normal conditions. Thus, plasma K<sup>+</sup> is at the mercy of the interplay between internal K<sup>+</sup> distribution and external K<sup>+</sup> balance mediated by renal K<sup>+</sup> excretion.

Recent years have seen remarkable advances in identifying the transport processes involved in renal and extrarenal K<sup>+</sup> balance and their regulation. Here we apply these advances in molecular physiology to understand the basis for longstanding observations of the effects of acid-base disturbances on serum potassium. We do not address the large spectrum of clinical syndromes that mutually affect K<sup>+</sup> and acid-base balance.

## Effects of Acid-Base Status on Internal K<sup>+</sup> Distribution

Most of the body K<sup>+</sup> content resides in the intracellular space of skeletal mus-

cle.<sup>2</sup> An overview of ion transport pathways that directly or indirectly mediate shifts of K<sup>+</sup> between muscle cells and the extracellular space in response to acid-base changes is shown in Figure 2.

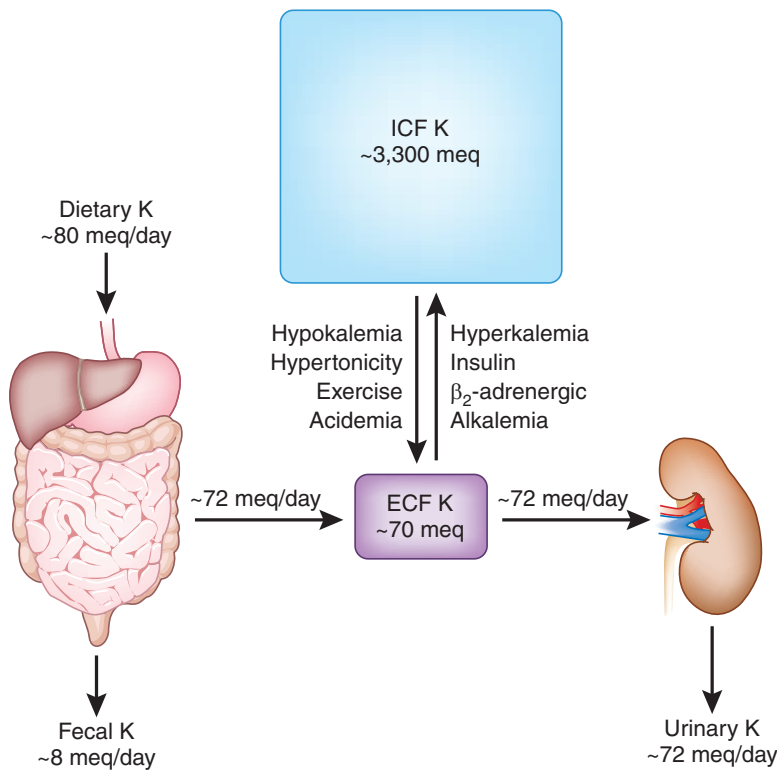
Muscle contraction is triggered by action potentials involving depolarizing Na<sup>+</sup> entry through Na<sup>+</sup> channels followed by membrane repolarization mediated by K<sup>+</sup> efflux through K<sup>+</sup> channels. Cl<sup>-</sup> channels play an important role in stabilizing the membrane potential and contributing to repolarization after action potentials. The electrochemical gradients of Na<sup>+</sup> and K<sup>+</sup> are restored by active Na<sup>+</sup> extrusion and K<sup>+</sup> uptake by the Na<sup>+</sup>,K<sup>+</sup>-ATPase.<sup>5</sup> Accordingly, cell ion content is determined by the balance between pump and leak pathways for Na<sup>+</sup> and K<sup>+</sup>.

However, muscle cells have additional pathways regulating intracellular pH homeostasis that can indirectly affect cellular Na<sup>+</sup> and K<sup>+</sup> balance.<sup>6</sup> Quantitatively, the most important pathway regulating intracellular pH in skeletal muscle is Na<sup>+</sup>-H<sup>+</sup> exchange,<sup>7</sup> as shown in Figure 2. Na<sup>+</sup>-H<sup>+</sup> exchange in skeletal muscle is highly dependent on intracellular pH, with marked activation by in-

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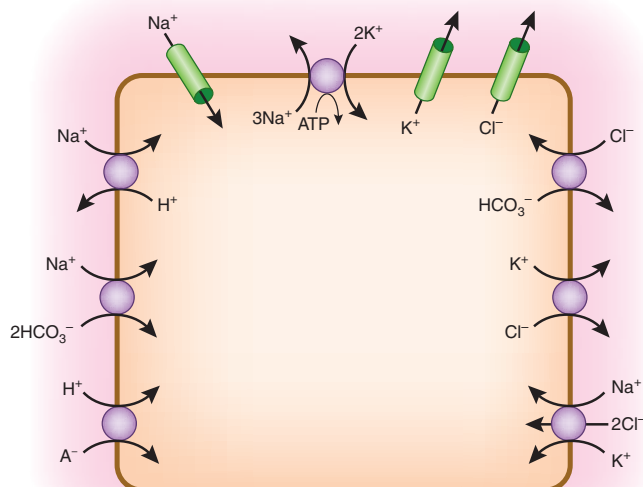


**Figure 1.**  $K^+$  concentration in the extracellular fluid (ECF) is affected by dietary intake, exchange with the intracellular fluid (ICF), and urinary excretion.

tracellular acidity and inhibition by alkalinity.<sup>8</sup> Activity of this pathway in response to acid-base perturbations strongly affects intracellular  $Na^+$  loading.<sup>7</sup>  $Na^+$ - $H^+$  exchanger isoform NHE1

is expressed in skeletal muscle and presumably accounts for most  $Na^+$ - $H^+$  exchange activity in this tissue.<sup>9</sup>

A lesser component of intracellular pH regulation in skeletal muscle is

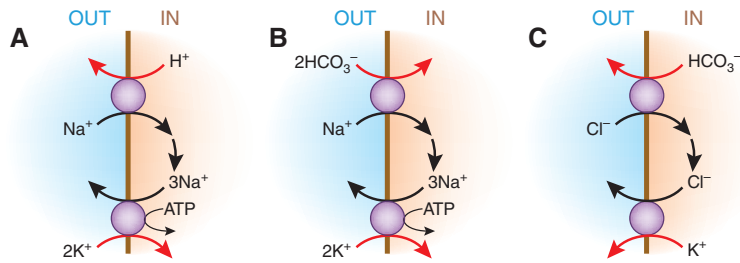


**Figure 2.** Multiple ion transport pathways directly or indirectly affect net  $K^+$  flux in skeletal muscle cells.

$HCO_3^-$ -dependent, because of  $Cl^-$ - $HCO_3^-$  exchange,<sup>7</sup> also shown in Figure 2. In addition, isoforms of the  $Na^+$ -bicarbonate cotransporter, NBCe1 and NBCe2, are expressed in muscle, raising the possibility that  $Na^+$ - $HCO_3^-$  cotransport contributes to intracellular pH regulation,<sup>10</sup> as also indicated in Figure 2. Another pathway of potential importance for cellular acid-base homeostasis is monocarboxylate cotransport that mediates coupled flux of  $H^+$  with such organic anions as lactate (Figure 2). Monocarboxylate cotransporters, MCT1 and MCT4, are expressed in skeletal muscle.<sup>11</sup> During conditions like lactic acidosis, this pathway will mediate influx of  $H^+$  and lactate, resulting in decreased intracellular pH. Cation-chloride cotransport pathways are also present. Expression of  $K^+$ - $Cl^-$  cotransporters KCC1, KCC3, and KCC4, as well as  $Na^+$ - $K^+$ - $Cl^-$  cotransporter NKCC1, has been detected in skeletal muscle.<sup>12–17</sup> Interaction of  $K^+$ - $Cl^-$  cotransport with acid-base transport will be discussed later.

Acute effects of acid-base disturbances on  $K^+$  redistribution have long been known.<sup>1,4</sup> In general, metabolic acidosis with acidemia causes a net shift of  $K^+$  from the intracellular to the extracellular space. Conversely, net cellular uptake of  $K^+$  is observed in metabolic alkalosis with alkalemia. The directional effects of acidemia and alkalemia on  $K^+$  redistribution are similar in respiratory acid-base disturbances as in metabolic derangements,<sup>4</sup> but the effects of respiratory disorders on  $K^+$  redistribution tend to be smaller than metabolic acid-base disturbances.<sup>4</sup>

How can these effects of acid-base disturbances on  $K^+$  redistribution be explained in terms of the underlying cellular transport mechanisms? The general effect of acidemia to cause  $K^+$  loss from cells is often attributed to membrane  $K^+$ - $H^+$  exchange. However, directly coupled  $K^+$ - $H^+$  exchange is undetected in skeletal muscle.<sup>7</sup> Nevertheless, reduction of extracellular pH results in net loss of  $K^+$  even from isolated muscle,<sup>18,19</sup> indicating this phenomenon is at least in part intrinsic to muscle and independent of changes in hormonal milieu as might



**Figure 3.** Apparent  $K^+$ - $H^+$  exchange (or  $K^+$ - $HCO_3^-$  cotransport) in skeletal muscle cells can arise from functional coupling between (A)  $Na^+$ - $H^+$  exchange and  $Na^+$ , $K^+$ -ATPase, (B)  $Na^+$ - $HCO_3^-$  cotransport and  $Na^+$ , $K^+$ -ATPase, or (C)  $Cl^-$ - $HCO_3^-$  exchange and  $K^+$ - $Cl^-$  cotransport.

occur *in vivo*. What then explains the apparent  $K^+$ - $H^+$  exchange?

As illustrated in Figure 3, the multiple acid-base transport pathways mentioned above may give rise to apparent  $K^+$ - $H^+$  exchange. In the case of the predominant pH regulatory pathway,  $Na^+$ - $H^+$  exchange,  $Na^+$  that enters by this route must be extruded by the  $Na^+$ , $K^+$ -ATPase (Figure 3A). Accordingly,  $K^+$  uptake by the  $Na^+$ , $K^+$ -ATPase will be greater when  $Na^+$ - $H^+$  exchange activity is stimulated and will be diminished when the rate of  $Na^+$ - $H^+$  exchange is reduced. In the case of acidosis with acidemia, the fall in extracellular pH would result in inhibition of the rate of  $Na^+$ - $H^+$  exchange, leading to the accumulation of intracellular  $H^+$  and a decline in intracellular  $Na^+$ . The latter would result in reduced  $Na^+$ , $K^+$ -ATPase activity, leading to decreased active cellular  $K^+$  uptake to counteract passive  $K^+$  efflux through  $K^+$  channels.<sup>20</sup> The final result would be as if  $H^+$  had entered the cell in exchange for  $K^+$ .

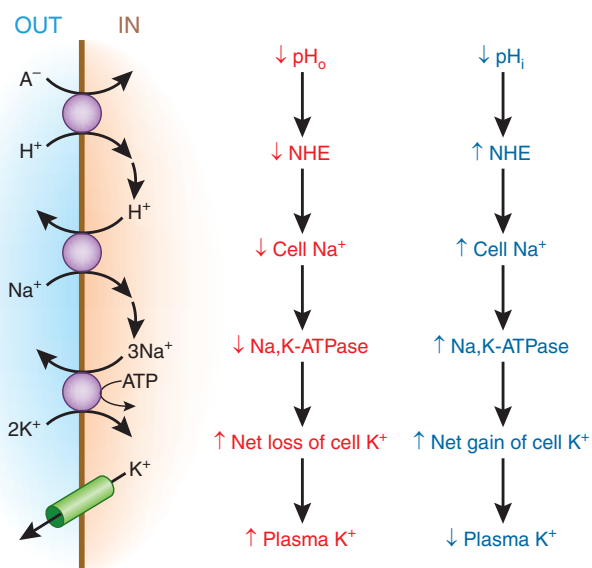
Similarly, as illustrated in Figure 3B,  $Na^+$ - $HCO_3^-$  cotransport operating in parallel with  $Na^+$ , $K^+$ -ATPase may result in  $K^+$ - $HCO_3^-$  cotransport, which is equivalent to  $K^+$ - $H^+$  exchange. For example, in the case of metabolic acidosis with acidemia, the fall in extracellular  $HCO_3^-$  results in inhibition of the inward rate of  $Na^+$ -bicarbonate cotransport, leading to a fall in intracellular  $Na^+$  and reduced  $Na^+$ , $K^+$ -ATPase activity. Lower  $Na^+$ , $K^+$ -ATPase activity would cause a net loss of cellular  $K^+$ . Again, the result would be as if  $H^+$  had entered the cell in exchange for  $K^+$ .

Finally,  $Cl^-$ - $HCO_3^-$  exchange also may contribute to apparent  $K^+$ - $H^+$  exchange if operating in parallel with  $K^+$ - $Cl^-$  cotransport, as shown in Figure 3C. Metabolic acidosis with a fall in extracellular  $HCO_3^-$  would increase the inward movement of  $Cl^-$  by  $Cl^-$ - $HCO_3^-$  exchange. The resulting rise in intracellular  $Cl^-$  would then promote  $K^+$  efflux by  $K^+$ - $Cl^-$  cotransport. The net result would be  $K^+$  efflux along with  $HCO_3^-$ , which is an equivalent process to exchanging intracellular  $K^+$  for extracellular  $H^+$ .

A striking observation has been that metabolic acidosis caused by mineral acid (hyperchloremic, nongap acidosis) causes a much larger shift of  $K^+$  into the extracellular fluid than does organic aci-

dosis (lactic acidosis).<sup>21</sup> The effect of hydrochloric acid but not organic acids to release  $K^+$  into the extracellular space had been observed using isolated muscle preparations, indicating this phenomenon can occur independently of systemic factors.<sup>22</sup> In the case of acidemia caused by an organic acidosis like lactic acidosis, there would again be the effect of both low extracellular pH and  $HCO_3^-$  tending to inhibit  $Na^+$ - $H^+$  exchange and  $Na^+$ -bicarbonate cotransport. This is illustrated for the case of  $Na^+$ - $H^+$  exchange in Figure 4, but in contrast to the situation with hyperchloremic acidosis, there would also be a strong inward flux of lactate and  $H^+$  through the monocarboxylate transporter, resulting in a larger fall in intracellular pH and  $HCO_3^-$ . The decrease in intracellular pH and  $HCO_3^-$  would tend to stimulate  $Na^+$  entry by  $Na^+$ - $H^+$  exchange and  $Na^+$ - $HCO_3^-$  cotransport, stimulating  $Na^+$ , $K^+$ -ATPase activity. The net effect would be to drive net cellular uptake of  $K^+$ .

Thus, as illustrated in Figure 4, extracellular and intracellular acidosis are predicted to have opposing effects on the distribution of  $K^+$  because of their differing effects on cellular  $Na^+$  loading. During organic acidosis, there will be greater cellular acidification and  $Na^+$  entry than during hyperchloremic acidosis, resulting in higher  $Na^+$ , $K^+$ -ATPase ac-



**Figure 4.** Opposing effects of extracellular and intracellular pH modify the influence of organic acidosis on plasma  $K^+$ .

tivity compared with hyperchloremic acidosis. However, in several tissues,  $\text{Na}^+, \text{K}^+$ -ATPase activity is affected by intracellular pH, with reduced activity when intracellular pH is lower than normal.<sup>23–25</sup> For intracellular acidification to stimulate net  $\text{K}^+$  uptake, it would require that the effect of low intracellular pH to inhibit  $\text{Na}^+, \text{K}^+$ -ATPase activity is less significant than the effect of intracellular  $\text{Na}^+$  loading to stimulate pump activity.

The acid-base mechanisms illustrated in Figures 2 and 3 also provide a possible explanation for the observation that bicarbonate can affect  $\text{K}^+$  redistribution independent of the effect of extracellular pH.<sup>26,27</sup>  $\text{Na}^+$  entry by  $\text{Na}^+/\text{HCO}_3^-$  cotransport would be enhanced whenever extracellular  $\text{HCO}_3^-$  is increased, resulting in increased cell  $\text{Na}^+$  uptake, stimulation of  $\text{Na}^+, \text{K}^+$ -ATPase activity, and net cellular  $\text{K}^+$  uptake (Figure 3B). Conversely, inhibition of  $\text{Na}^+/\text{HCO}_3^-$  cotransport when extracellular  $\text{HCO}_3^-$  is reduced leads to a net loss of cell  $\text{K}^+$ . Analogously, the rate of  $\text{Cl}^-$  entry by  $\text{Cl}^-/\text{HCO}_3^-$  exchange would be higher when extracellular  $\text{HCO}_3^-$  is reduced, increasing cell  $\text{Cl}^-$  and enhancing the exit of  $\text{K}^+$  by  $\text{K}^+/\text{Cl}^-$  cotransport (Figure 3C). Conversely,  $\text{Cl}^-$  entry by  $\text{Cl}^-/\text{HCO}_3^-$  exchange would be lower when extracellular  $\text{HCO}_3^-$  is increased, leading to reduced  $\text{K}^+$  efflux by  $\text{K}^+/\text{Cl}^-$  cotransport.

Similar considerations may also account for the smaller shifts in  $\text{K}^+$  observed with respiratory acidosis compared with metabolic acidosis.<sup>4</sup> In respiratory acidosis, there is a fall in extracellular pH, but extracellular bicarbonate is elevated. One would therefore expect that  $\text{Na}^+/\text{H}^+$  exchange is inhibited as in metabolic acidosis with equivalent acidemia, but  $\text{Na}^+/\text{bicarbonate}$  cotransport would not be reduced. Accordingly, as compared with metabolic acidosis, respiratory acidosis would be associated with a smaller decrement in intracellular  $\text{Na}^+$ , less inhibition of  $\text{Na}^+, \text{K}^+$ -ATPase activity, and reduced net  $\text{K}^+$  loss from the cell. In addition, during respiratory acidosis with elevated  $\text{pCO}_2$ , rapid cell entry of  $\text{CO}_2$  will acidify intracellular pH. As discussed above for the case of organic acidosis, acidification

of intracellular pH, by stimulating  $\text{Na}^+$  entry by  $\text{Na}^+/\text{H}^+$  exchange tends to enhance  $\text{Na}^+, \text{K}^+$ -ATPase activity and oppose a net loss of intracellular  $\text{K}^+$ .

In view of the longstanding observations discussed above on  $\text{K}^+$  redistribution in acid-base disorders, one would expect alkalization by  $\text{HCO}_3^-$  administration to be an effective modality for acute treatment of hyperkalemia. However, some investigators have failed to find an effect of  $\text{HCO}_3^-$  administration to lower plasma  $\text{K}^+$  in hyperkalemic patients.<sup>28–30</sup> An effect of  $\text{HCO}_3^-$  administration to lower plasma  $\text{K}^+$  has been more striking in patients with more severe degrees of pre-existing acidosis than in those with only minimal reductions of plasma  $\text{HCO}_3^-$ .<sup>31</sup> One possible factor modifying the effect of extracellular  $\text{HCO}_3^-$  and pH on  $\text{K}^+$  distribution is the level of intracellular pH and  $\text{HCO}_3^-$ . At any given extracellular pH and  $\text{HCO}_3^-$ ,  $\text{Na}^+$  entry by  $\text{Na}^+/\text{H}^+$  exchange and  $\text{Na}^+/\text{bicarbonate}$  cotransport is greater when intracellular pH and  $\text{HCO}_3^-$  are reduced, as discussed earlier. Patients with appreciable pre-existing metabolic acidosis would be expected to have lower intracellular pH and  $\text{HCO}_3^-$ . This may account for the fact that the effect of  $\text{HCO}_3^-$  administration to reduce plasma  $\text{K}^+$  has been more striking in patients with pre-existing acidosis.<sup>31</sup>

Effects of pH and  $\text{HCO}_3^-$  on internal  $\text{K}^+$  distribution may be modified by hormonal systems that affect cellular  $\text{K}^+$  uptake and release. For example, net cellular uptake of  $\text{K}^+$  is strongly stimulated by insulin because of increased  $\text{Na}^+, \text{K}^+$ -ATPase activity.<sup>32</sup> There is evidence that stimulation of insulin secretion by acidosis diminishes the hyperkalemia otherwise resulting from acidosis.<sup>32</sup> Moreover, differential effects of organic *versus* hyperchloremic acidosis on insulin and glucagon secretion may contribute to the differing effects of these forms of acidosis on plasma  $\text{K}^+$  as discussed earlier.<sup>33</sup> Although skeletal muscle is the predominant source of intracellular  $\text{K}^+$  content, there is evidence that the effect of organic acid-induced insulin secretion on plasma  $\text{K}^+$  is mediated at least in part by

hepatic  $\text{K}^+$  uptake.<sup>33</sup> The interactions of acid-base disturbances with other hormonal systems are at present incompletely defined.

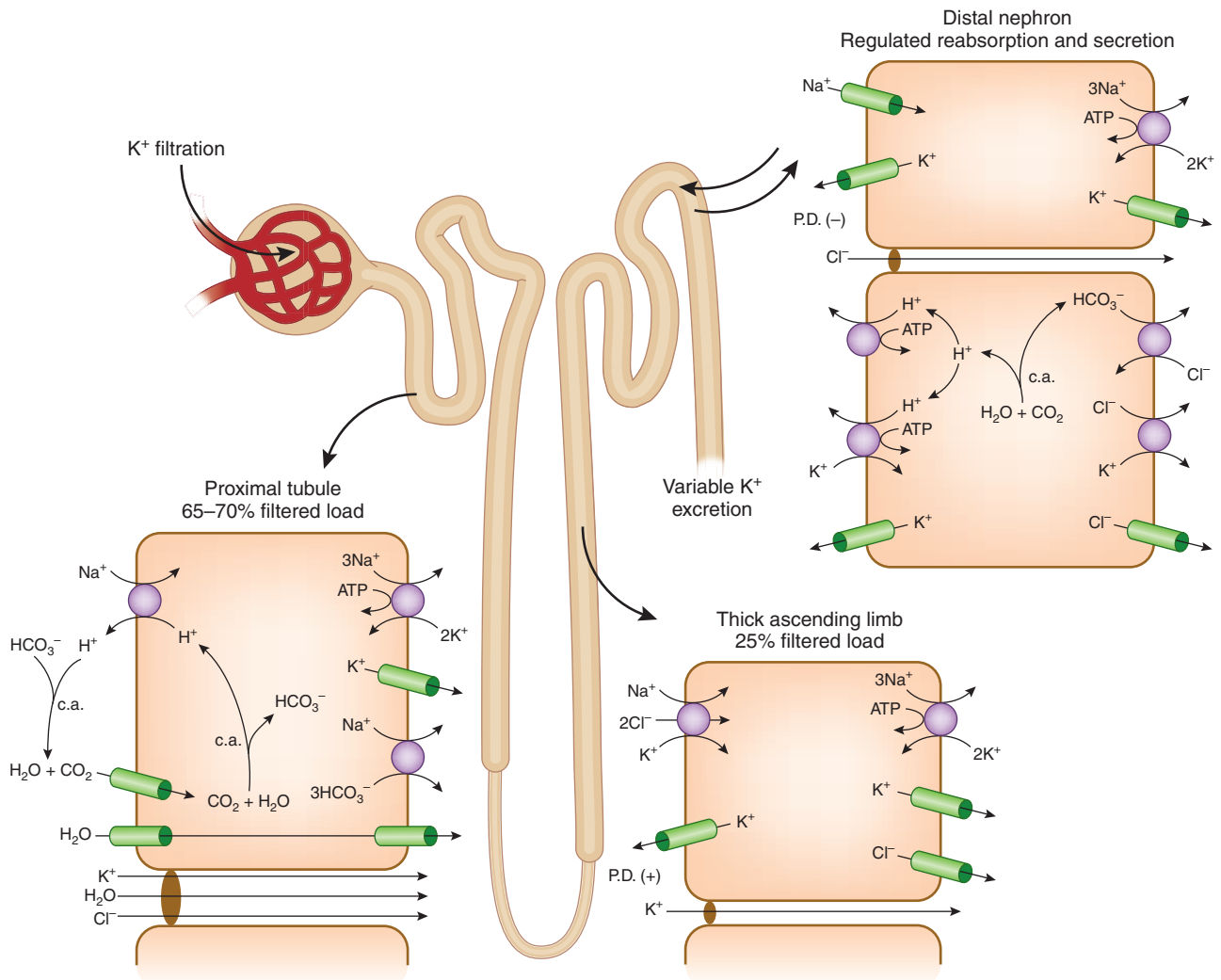
### Effects of Acid-Base on Renal $\text{K}^+$ Excretion

The geography of net  $\text{K}^+$  transport along the nephron is shown in Figure 5. The bulk of filtered potassium is reabsorbed in the proximal tubule, where its reabsorption is predominantly passive and paracellular.<sup>34</sup> Such passive reabsorption ultimately depends on and is driven by fluid reabsorption, which in turn is a function of  $\text{Na}^+$  reabsorption in this highly water-permeable nephron segment. Accordingly, the proximal tubule reabsorbs  $\text{K}^+$  in approximate proportion to reabsorption of filtered  $\text{Na}^+$  and water, accounting for retrieval of some 70% of the filtered load of  $\text{K}^+$ .

Most of the remaining fraction of filtered  $\text{K}^+$  (approximately 25%) is reabsorbed in the thick ascending limb by a transcellular route involving  $\text{Na}^+/\text{K}^+/\text{Cl}^-$  cotransport across the apical membrane and extrusion by the  $\text{Na}^+, \text{K}^+$ -ATPase across the basolateral membrane.<sup>34</sup> As a consequence, only about 5% of the filtered  $\text{K}^+$  reaches the distal tubule.

The critical sites in the nephron responsible for determining  $\text{K}^+$  excretion are the connecting tubule and collecting tubule.<sup>35</sup> Under conditions of normal or elevated dietary  $\text{K}^+$  intake, these segments mediate net secretion of  $\text{K}^+$ . However, under conditions of  $\text{K}^+$  depletion, net  $\text{K}^+$  reabsorption may occur in this portion of the nephron.

Cell models for  $\text{K}^+$  secretion and reabsorption are illustrated in Figure 6.  $\text{K}^+$  secretion, which takes place through connecting tubule cells and principal cells of the collecting tubule, involves active uptake across the basolateral membrane mediated by  $\text{Na}^+, \text{K}^+$ -ATPase, followed by passive exit across the apical membrane through  $\text{K}^+$  channels. The ENaC  $\text{Na}^+$  channel in the apical membrane indirectly plays a critical role in  $\text{K}^+$  secretion by providing intracellular  $\text{Na}^+$  as substrate for the  $\text{Na}^+, \text{K}^+$ -ATPase and by depolarizing the apical membrane, thereby increasing the driving force for



**Figure 5.** Urinary  $K^+$  excretion is the resultant of filtration, reabsorption and secretion along the nephron. Mechanisms of  $K^+$  transport in proximal tubule, thick ascending limb, and distal nephron (connecting and collecting tubule) are shown in the inset.

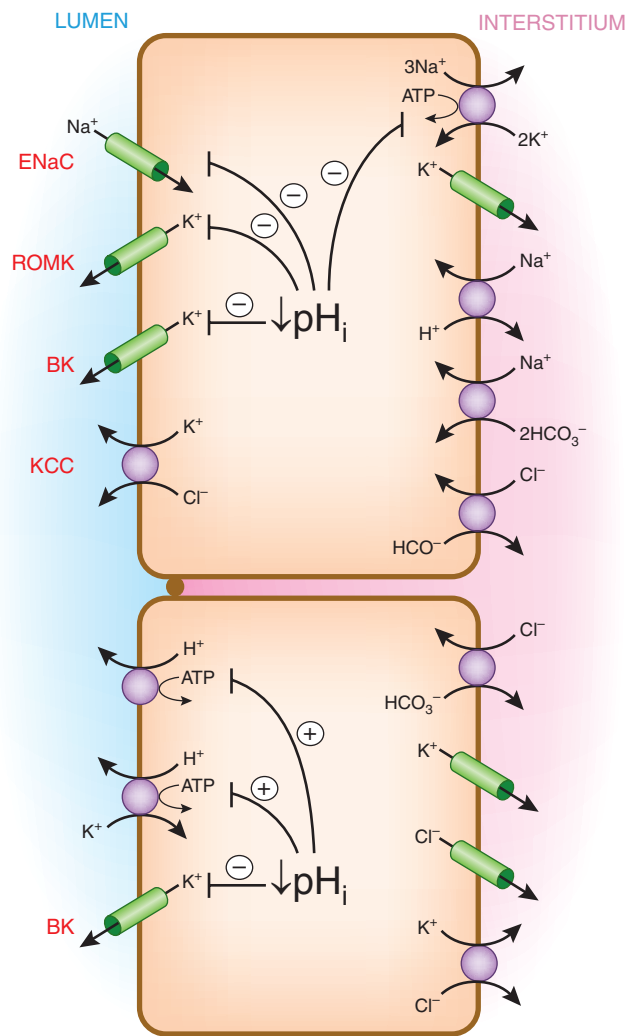
$K^+$  efflux into the tubule fluid. At least two types of apical  $K^+$  channels have been implicated in  $K^+$  secretion, ROMK and BK.<sup>36–39</sup> In addition, effects of luminal  $Cl^-$  on  $K^+$  secretion suggest that a component of apical membrane  $K^+$  efflux takes place by electroneutral  $K^+-Cl^-$  cotransport,<sup>40,41</sup> although its molecular mechanism is unclear.

Multiple factors modulate  $K^+$  secretion in these distal nephron segments. First, plasma  $K^+$  concentration is an important determinant of the rate of  $K^+$  secretion. External  $K^+$  is substrate for the  $Na^+,K^+-ATPase$ , and there is saturable dependence of  $K^+$  secretion on plasma  $K^+$  concentration.<sup>42</sup> In addition, the higher the plasma  $K^+$ , the less backflux

of  $K^+$  takes place through basolateral  $K^+$  channels. Second, aldosterone, whose secretion is stimulated by elevated plasma  $K^+$ , is a major regulator of  $K^+$  secretion in distal nephron segments. Aldosterone acts through several signaling pathways and effector mechanisms to bring about increased activities of apical  $Na^+$  and  $K^+$  channels and increased activity of basolateral  $Na^+,K^+-ATPase$ .<sup>35,43,44</sup> Third,  $K^+$  secretion is highly affected by luminal  $Na^+$  and fluid delivery to distal  $K^+$  secretory sites.<sup>45,46</sup>  $K^+$  secretion is dependent directly and indirectly on luminal  $Na^+$  entry through ENaC, as discussed above, which in turn is a function of luminal  $Na^+$  concentration.<sup>47</sup>  $K^+$  secretion is enhanced at higher luminal flow rates be-

cause of activation of BK channels.<sup>46</sup> Recent studies suggest that flow activation of BK channels involves intracellular  $Ca^{++}$  signaling most likely in response to mechanosensing by apical cilia.<sup>46</sup> Finally, net  $K^+$  secretion is modified by modulation of active  $K^+$  reabsorption mediated by apical  $H^+,K^+-ATPase$  in intercalated cells.<sup>48,49</sup> Chronic  $K^+$  depletion causes upregulation of  $K^+$  reabsorption mediated by apical  $H^+,K^+-ATPase$  in the collecting tubule.<sup>50,51</sup>

Acid-base disturbances have complex effects on renal  $K^+$  excretion. For example, acute acidemia caused by metabolic acidosis reduces renal  $K^+$  excretion, whereas metabolic acidosis at a later phase promotes urinary  $K^+$  excretion



**Figure 6.** Multiple ion transport pathways directly or indirectly affect  $K^+$  transport in the connecting tubule and cortical collecting tubule and are affected by pH.

and development of  $K^+$  deficiency.<sup>52</sup> How can these seemingly contradictory effects of acidemia be explained?

The principal cells of the cortical collecting tubule have transporters regulating intracellular pH that can indirectly affect  $K^+$  secretion. Specifically, activities of  $Na^+-H^+$  exchange,  $Na^+-HCO_3^-$  cotransport and  $Cl^- -HCO_3^-$  exchange at the basolateral membrane of principal cells have been observed.<sup>53–56</sup>  $Na^+-H^+$  exchanger isoform NHE1 is strongly expressed on the basolateral membrane of connecting tubule and principal cells.<sup>57</sup> As discussed earlier for the case of muscle, activities of  $Na^+-H^+$  exchange and  $Na^+-HCO_3^-$  cotransport can affect intracellular  $Na^+$  loading and  $Na^+$ ,  $K^+$ -ATPase activity. Changes in  $Na^+$ ,  $K^+$ -AT-

Pase activity would affect basolateral  $K^+$  uptake and transtubular  $K^+$  secretion. In fact, basolateral  $Na^+$  entry by  $Na^+-H^+$  exchange can support  $K^+$  secretion in the absence of luminal  $Na^+$ .<sup>58</sup> Thus, inhibition of  $Na^+-H^+$  exchange and  $Na^+-HCO_3^-$  cotransport during metabolic acidosis with acidemia reduces cell  $Na^+$  and thereby inhibits  $K^+$  secretion. Conversely, metabolic alkalosis with alkalemia would tend to stimulate  $K^+$  secretion.

Several of the pathways involved in distal nephron  $K^+$  secretion are directly affected by pH, as illustrated in Figure 6. A fall in intracellular pH reduces activity of  $Na^+$ ,  $K^+$ -ATPase on the basolateral membrane<sup>23–25</sup> and diminishes activities of ENaC, ROMK, and BK on the apical membrane.<sup>59–62</sup> These effects could con-

tribute to inhibition of active  $K^+$  secretion in response to acidemia to the extent that there is parallel intracellular acidosis. ENaC abundance is decreased when luminal or basolateral  $HCO_3^-$  (and pH) is reduced.<sup>63</sup> In addition, stimulation of electrogenic  $H^+$  secretion by the vacuolar ATPase in intercalated cells during acidemia would tend to reduce the lumen-negative transepithelial potential difference,<sup>64</sup> inhibiting  $K^+$  secretion. Furthermore, acidemia upregulates apical  $H^+$ ,  $K^+$ -ATPase in intercalated cells,<sup>65</sup> thereby enhancing  $K^+$  reabsorption and reducing net  $K^+$  secretion.

However, metabolic acidosis causes increased distal  $Na^+$  delivery and flow rate, as well as increased urinary  $Na^+$  excretion.<sup>42,52</sup> There are at least four possible factors contributing to this phenomenon. First,  $Na^+$ ,  $K^+$ -ATPase in all nephron segments may be inhibited by acidemia with reduced intracellular pH, leading to diminished renal tubular  $Na^+$  reabsorption upstream of the  $K^+$  secretory sites. Second, although acidosis causes upregulation of NHE3 activity in the proximal tubule,<sup>66,67</sup> the absolute amount of  $Na^+$  reabsorbed with  $HCO_3^-$  in the proximal tubule is actually reduced in metabolic acidosis because of the diminished filtered load of  $HCO_3^-$ .<sup>68</sup> Third, the reduction in absolute fluid reabsorption coupled to  $NaHCO_3$  reabsorption in metabolic acidosis results in a less than normal increment in luminal  $Cl^-$  concentration along the length of the proximal tubule, lowering the driving force for passive paracellular  $NaCl$  reabsorption.<sup>68</sup> Fourth, apical membrane  $Cl^-$ -base exchange activity is downregulated in metabolic acidosis, reducing transcellular  $NaCl$  reabsorption in the proximal tubule.<sup>69</sup>

These effects of metabolic acidosis to enhance distal  $Na^+$  delivery and flow rate and to augment urinary  $Na^+$  excretion tend to cause volume depletion, resulting in increased renin and aldosterone secretion.<sup>70</sup> In addition, acidosis may directly stimulate aldosterone secretion independent of renin secretion.<sup>70,71</sup> As discussed above,  $K^+$  secretion in the distal nephron is strongly stimulated by

aldosterone and by increased luminal  $\text{Na}^+$  delivery and flow rate. These factors eventually predominate over the local inhibitory effect of acidemia on the  $\text{K}^+$  secreting cells, resulting in increased  $\text{K}^+$  excretion and negative  $\text{K}^+$  balance. Ultimately, a new steady state is established as  $\text{K}^+$  balance is restored at the expense of hypokalemia.

This pathophysiology is well illustrated by the disorder of classic distal RTA. Although there are exceptions,<sup>72</sup> these patients generally have hypokalemia caused by renal  $\text{K}^+$  wasting during chronic acidosis, which is corrected by base administration.<sup>73,74</sup> Another clinical example in which the effect of distal  $\text{Na}^+$  delivery and flow to stimulate urinary  $\text{K}^+$  excretion predominates over the inhibitory effect of acidemia is diabetic ketoacidosis with osmotic diuresis.<sup>75</sup> Moreover, organic acidosis with high rates of excretion of nonchloride anions in the urine will enhance distal  $\text{K}^+$  secretion caused by the reduced luminal  $\text{Cl}^-$  concentration through presumed apical membrane  $\text{K}^+-\text{Cl}^-$  cotransport.<sup>40,41</sup>

Acidemia caused by respiratory acidosis results in similar directional changes in urinary  $\text{K}^+$  excretion as does metabolic acidosis. Acute acidemia caused by respiratory acidosis inhibits renal  $\text{K}^+$  secretion in the distal nephron<sup>76</sup> and reduces urinary  $\text{K}^+$  excretion.<sup>52</sup> Over the following days, respiratory acidosis results in urinary  $\text{K}^+$  wasting associated with increased  $\text{Na}^+$  excretion.<sup>52</sup> It is likely these effects are mediated by many of the same mechanisms discussed above for metabolic acidosis. However, the effects of chronic acidemia to augment  $\text{K}^+$  excretion caused by increased distal  $\text{Na}^+$  and fluid delivery will tend to be milder for respiratory acidosis than for metabolic acidosis.<sup>52</sup> First, renal compensation of chronic respiratory acid-base disorders restores blood pH much closer to normal than does respiratory compensation of metabolic acid-base disorders.<sup>77,78</sup> Thus, acidemia will be milder in chronic respiratory acidosis than in chronic metabolic acidosis, resulting in less inhibition of  $\text{Na}^+, \text{K}^+-\text{ATPase}$  and less augmentation of  $\text{Na}^+$  and fluid delivery to the distal nephron. Second, the role of low filtered

load of  $\text{HCO}_3^-$  in limiting absolute  $\text{NaHCO}_3$  reabsorption along the proximal tubule in metabolic acidosis as discussed above would not be a contributing factor in respiratory acidosis.

In general, alkalemia stimulates distal nephron  $\text{K}^+$  secretion and urinary  $\text{K}^+$  excretion.<sup>52</sup> These effects are larger for metabolic than for respiratory alkalosis and are in essence the converse of the effects of acidemia on the  $\text{K}^+$  secretory pathways, because elevated intracellular pH tends to increase activities of ENaC, ROMK, and BK.<sup>59–62</sup> ENaC abundance is increased when luminal or basolateral  $\text{HCO}_3^-$  (and pH) is elevated.<sup>63</sup> Moreover, in the case of acute metabolic alkalosis, there is inhibition of fractional  $\text{NaHCO}_3$  and fluid reabsorption in the proximal tubule, leading to increased distal delivery of  $\text{Na}^+$  and  $\text{HCO}_3^-$  and enhanced fluid flow.<sup>79</sup> As already discussed, increased  $\text{Na}^+$  delivery and fluid flow stimulate  $\text{K}^+$  secretion. In addition, increased luminal delivery of  $\text{HCO}_3^-$  as a nonchloride anion stimulates a component of distal  $\text{K}^+$  secretion because of the reduced luminal  $\text{Cl}^-$  concentration as mentioned earlier.<sup>41</sup> Of course, in the maintenance phase of metabolic alkalosis with volume contraction, distal  $\text{Na}^+$  and  $\text{HCO}_3^-$  delivery will decline, thereby mitigating the rate of  $\text{K}^+$  wasting.<sup>80</sup>

### Summary and Conclusions

We have used new information about the molecular physiology of extrarenal and renal potassium transport to explain longstanding observations of the complex effects of acid-base disturbances on serum potassium. It should be emphasized that many aspects of these explanations are somewhat speculative because the quantitative contributions of specific pathways to mediating effects of pH on extrarenal and renal  $\text{K}^+$  transport are not known with certainty. Moreover, we have not considered new developments concerning molecular sensors and signaling mechanisms that respond to changes in extracellular and intracellular pH,  $\text{CO}_2$ , and  $\text{HCO}_3^-$ .<sup>81</sup> The roles of these sensor and signaling mechanisms in mediating the effects of acid-base distur-

bances on extrarenal and renal  $\text{K}^+$  homeostasis are not yet known.

Nevertheless, it should be apparent that there is no simple relationship between pH and serum potassium because of the multiplicity of factors affecting internal and external  $\text{K}^+$  balance. For example, consider the patient with diabetic ketoacidosis. Acidemia will tend to shift  $\text{K}^+$  out of cells and cause hyperkalemia, but this effect is less pronounced in organic acidosis than in mineral acidosis. On the other hand, hypertonicity in the absence of insulin will promote  $\text{K}^+$  release into the extracellular space. Renal  $\text{K}^+$  excretion will be acutely inhibited by acidemia but ultimately enhanced by the increased distal  $\text{Na}^+$  delivery and flow rate caused by metabolic acidosis and osmotic diuresis in the setting of high aldosterone. Indeed, the patient may present with marked  $\text{K}^+$  depletion if osmotic diuresis has been going on for some time. Renal  $\text{K}^+$  excretion may later become reduced when GFR falls as volume depletion ensues. Accordingly, there will not be a straightforward relationship between serum potassium and acid-base status in such a patient. The clinician will need to be knowledgeable about the many factors affecting internal and external  $\text{K}^+$  balance to provide optimal patient care.

### DISCLOSURES

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### REFERENCES

1. Burnell JM, Scribner BH, Uyeno BT, Villamil MF: The effect in humans of extracellular pH change on the relationship between serum potassium concentration and intracellular potassium. *J Clin Invest* 35: 935–939, 1956
2. Youn JH, McDonough AA: Recent advances in understanding integrative control of potassium homeostasis. *Annu Rev Physiol* 71: 381–401, 2009
3. Bia MJ, DeFronzo RA: Extrarenal potassium

- homeostasis. *Am J Physiol* 240: F257–F268, 1981
4. Adrogue HJ, Madias NE: Changes in plasma potassium concentration during acute acid-base disturbances. *Am J Med* 71: 456–467, 1981
  5. Clausen T: Na<sup>+</sup>-K<sup>+</sup> pump regulation and skeletal muscle contractility. *Physiol Rev* 83: 1269–1324, 2003
  6. Juel C: Regulation of pH in human skeletal muscle: Adaptations to physical activity. *Acta Physiol* 193: 17–24, 2008
  7. Aickin CC, Thomas RC: An investigation of the ionic mechanism of intracellular pH regulation in mouse soleus muscle fibres. *J Physiol* 273: 295–316, 1977
  8. Juel C: Skeletal muscle Na<sup>+</sup>/H<sup>+</sup> exchange in rats: pH dependency and the effect of training. *Acta Physiol Scand* 164: 135–140, 1998
  9. Juel C: Expression of the Na<sup>+</sup>/H<sup>+</sup> exchanger isoform NHE1 in rat skeletal muscle and effect of training. *Acta Physiol Scand* 170: 59–63, 2000
  10. Kristensen JM, Kristensen M, Juel C: Expression of Na<sup>+</sup>/HCO<sub>3</sub><sup>-</sup> co-transporter proteins (NBCs) in rat and human skeletal muscle. *Acta Physiol Scand* 182: 69–76, 2004
  11. Juel C: Lactate-proton cotransport in skeletal muscle. *Physiol Rev* 77: 321–358, 1997
  12. Gillen CM, Brill S, Payne JA, Forbush B 3rd: Molecular cloning and functional expression of the K-Cl cotransporter from rabbit, rat, and human: A new member of the cation-chloride cotransporter family. *J Biol Chem* 271: 16237–16244, 1996
  13. Race JE, Makhlof FN, Logue PJ, Wilson FH, Dunham PB, Holtzman EJ: Molecular cloning and functional characterization of KCC3, a new K-Cl cotransporter. *Am J Physiol* 277: C1210–C1219, 1999
  14. Mount DB, Mercado A, Song L, Xu J, George AL Jr, Delpire E, Gamba G: Cloning and characterization of KCC3 and KCC4, new members of the cation-chloride cotransporter gene family. *J Biol Chem* 274: 16355–16362, 1999
  15. Hiki K, D'Andrea RJ, Furze J, Crawford J, Woollatt E, Sutherland GR, Vadas MA, Gamble JR: Cloning, characterization, and chromosomal location of a novel human K<sup>+</sup>-Cl<sup>-</sup> cotransporter. *J Biol Chem* 274: 10661–10667, 1999
  16. Wong JA, Fu L, Schneider EG, Thomason DB: Molecular and functional evidence for Na<sup>+</sup>-K<sup>+</sup>-2Cl<sup>-</sup> cotransporter expression in rat skeletal muscle. *Am J Physiol* 277: R154–R161, 1999
  17. Kristensen M, Hansen T, Juel C: Membrane proteins involved in potassium shifts during muscle activity and fatigue. *Am J Physiol Regul Integr Comp Physiol* 290: R766–R772, 2006
  18. Rogers TA: Tissue buffering in rat diaphragm. *Am J Physiol* 191: 363–366, 1957
  19. Fenn WO, Rogers TA, Ohr EA: Muscle electrolytes in acid and alkaline solutions. *Am J Physiol* 194: 373–378, 1958
  20. Kamel KS, Wei C: Controversial issues in the treatment of hyperkalemia. *Nephrol Dial Transplant* 18: 2215–2218, 2003
  21. Oster JR, Perez GO, Vaamonde CA: Relationship between blood pH and potassium and phosphorus during acute metabolic acidosis. *Am J Physiol* 235: F345–F351, 1978
  22. Rogers TA, Wachenfeld AE: Effect of physiologic acids on electrolytes in rat diaphragm. *Am J Physiol* 193: 623–626, 1958
  23. Russell JM, Boron WF, Brodwick MS: Intracellular pH and Na fluxes in barnacle muscle with evidence for reversal of the ionic mechanism of intracellular pH regulation. *J Gen Physiol* 82: 47–78, 1983
  24. Eaton DC, Hamilton KL, Johnson KE: Intracellular acidosis blocks the basolateral Na-K pump in rabbit urinary bladder. *Am J Physiol* 247: F946–F954, 1984
  25. Breitwieser GE, Altamirano AA, Russell JM: Effects of pH changes on sodium pump fluxes in squid giant axon. *Am J Physiol* 253: C547–C554, 1987
  26. Fraley DS, Adler S: Isohydric regulation of plasma potassium by bicarbonate in the rat. *Kidney Int* 9: 333–343, 1976
  27. Fraley DS, Adler S: Correction of hyperkalemia by bicarbonate despite constant blood pH. *Kidney Int* 12: 354–360, 1977
  28. Blumberg A, Weidmann P, Shaw S, Gnadinger M: Effect of various therapeutic approaches on plasma potassium and major regulating factors in terminal renal failure. *Am J Med* 85: 507–512, 1988
  29. Blumberg A, Weidmann P, Ferrari P: Effect of prolonged bicarbonate administration on plasma potassium in terminal renal failure. *Kidney Int* 41: 369–374, 1992
  30. Gutierrez R, Schlessinger F, Oster JR, Rietberg B, Perez GO: Effect of hypertonic versus isotonic sodium bicarbonate on plasma potassium concentration in patients with end-stage renal disease. *Miner Electrolyte Metab* 17: 297–302, 1991
  31. Schwarz KC, Cohen BD, Lubash GD, Rubin AL: Severe acidosis and hyperkalemia treated with sodium bicarbonate infusion. *Circulation* 19: 215–220, 1959
  32. Wiederseiner JM, Muser J, Lutz T, Hulter HN, Krampf R: Acute metabolic acidosis: Characterization and diagnosis of the disorder and the plasma potassium response. *J Am Soc Nephrol* 15: 1589–1596, 2004
  33. Adrogue HJ, Chap Z, Ishida T, Field JB: Role of the endocrine pancreas in the kalemic response to acute metabolic acidosis in conscious dogs. *J Clin Invest* 75: 798–808, 1985
  34. Giebisch G, Krampf R, Wagner C: Renal and extrarenal regulation of potassium. *Kidney Int* 72: 397–410, 2007
  35. Wang WH, Giebisch G: Regulation of potassium (K) handling in the renal collecting duct. *Pflugers Arch* 458: 157–168, 2009
  36. Palmer LG: Potassium secretion and the regulation of distal nephron K channels. *Am J Physiol* 277: F821–F825, 1999
  37. Hebert SC, Desir G, Giebisch G, Wang W: Molecular diversity and regulation of renal potassium channels. *Physiol Rev* 85: 319–371, 2005
  38. Welling PA, Ho K: A comprehensive guide to the ROMK potassium channel: Form and function in health and disease. *Am J Physiol Renal Physiol* 297: F849–F863, 2009
  39. Pluznick JL, Sansom SC: BK channels in the kidney: Role in K<sup>+</sup> secretion and localization of molecular components. *Am J Physiol Renal Physiol* 291: F517–F529, 2006
  40. Ellison DH, Velazquez H, Wright FS: Stimulation of distal potassium secretion by low lumen chloride in the presence of barium. *Am J Physiol* 248: F638–F649, 1985
  41. Amorim JB, Bailey MA, Musa-Aziz R, Giebisch G, Malnic G: Role of luminal anion and pH in distal tubule potassium secretion. *Am J Physiol Renal Physiol* 284: F381–F388, 2003
  42. Stanton BA, Giebisch G: Effects of pH on potassium transport by renal distal tubule. *Am J Physiol* 242: F544–F551, 1982
  43. Vinciguerra M, Mordasini D, Vandewalle A, Feraille E: Hormonal and nonhormonal mechanisms of regulation of the Na,K-pump in collecting duct principal cells. *Semin Nephrol* 25: 312–321, 2005
  44. Verrey F, Fakitsas P, Adam G, Staub O: Early transcriptional control of ENaC (de)ubiquitylation by aldosterone. *Kidney Int* 73: 691–696, 2008
  45. Khuri RN, Strieder WN, Giebisch G: Effects of flow rate and potassium intake on distal tubular potassium transfer. *Am J Physiol* 228: 1249–1261, 1975
  46. Satlin LM, Carattino MD, Liu W, Kleyman TR: Regulation of cation transport in the distal nephron by mechanical forces. *Am J Physiol Renal Physiol* 291: F923–F931, 2006
  47. Palmer LG, Sackin H, Frindt G: Regulation of Na<sup>+</sup> channels by luminal Na<sup>+</sup> in rat cortical collecting tubule. *J Physiol* 509: 151–162, 1998
  48. Codina J, DuBose TD Jr: Molecular regulation and physiology of the H<sup>+</sup>,K<sup>+</sup>-ATPases in kidney. *Semin Nephrol* 26: 345–351, 2006
  49. Gumz ML, Lynch IJ, Greenlee MM, Cain BD, Wingo CS: The renal H<sup>+</sup>-K<sup>+</sup>-ATPases: Physiology, regulation, and structure. *Am J Physiol Renal Physiol* 298: F12–F21, 2010
  50. Wingo CS: Active proton secretion and potassium absorption in the rabbit outer medullary collecting duct: Functional evidence for proton-potassium-activated adenosine triphosphatase. *J Clin Invest* 84: 361–365, 1989
  51. Cheval L, Barlet-Bas C, Khadouri C, Feraille E, Marsy S, Doucet A: K<sup>+</sup>-ATPase-mediated Rb<sup>+</sup> transport in rat collecting tubule:



- Modulation during K<sup>+</sup> deprivation. *Am J Physiol* 260: F800–F805, 1991
52. Gennari FJ, Cohen JJ: Role of the kidney in potassium homeostasis: Lessons from acid-base disturbances. *Kidney Int* 8: 1–5, 1975
  53. Chaillet JR, Lopes AG, Boron WF: Basolateral Na-H exchange in the rabbit cortical collecting tubule. *J Gen Physiol* 86: 795–812, 1985
  54. Wang X, Kurtz I: H<sup>+</sup>/base transport in principal cells characterized by confocal fluorescence imaging. *Am J Physiol* 259: C365–C373, 1990
  55. Weiner ID, Hamm LL: Regulation of intracellular pH in the rabbit cortical collecting tubule. *J Clin Invest* 85: 274–281, 1990
  56. Silver RB, Frindt G, Palmer LG: Regulation of principal cell pH by Na/H exchange in rabbit cortical collecting tubule. *J Membr Biol* 125: 13–24, 1992
  57. Biemesderfer D, Reilly RF, Exner M, Igarashi P, Aronson PS: Immunocytochemical characterization of Na(+)-H<sup>+</sup> exchanger isoform NHE-1 in rabbit kidney. *Am J Physiol* 263: F833–F840, 1992
  58. Muto S, Tsuruoka S, Miyata Y, Fujimura A, Kusano E, Wang W, Seldin D, Giebisch G: Basolateral Na<sup>+</sup>/H<sup>+</sup> exchange maintains potassium secretion during diminished sodium transport in the rabbit cortical collecting duct. *Kidney Int* 75: 25–30, 2009
  59. Palmer LG, Frindt G: Effects of cell Ca and pH on Na channels from rat cortical collecting tubule. *Am J Physiol* 253: F333–F339, 1987
  60. Wang WH, Schwab A, Giebisch G: Regulation of small-conductance K<sup>+</sup> channel in apical membrane of rat cortical collecting tubule. *Am J Physiol* 259: F494–F502, 1990
  61. Hirsch J, Leipziger J, Frobe U, Schlatter E: Regulation and possible physiological role of the Ca(2+)-dependent K<sup>+</sup> channel of cortical collecting ducts of the rat. *Pflugers Arch* 422: 492–498, 1993
  62. Schlatter E, Haxelmans S, Hirsch J, Leipziger J: pH dependence of K<sup>+</sup> conductances of rat cortical collecting duct principal cells. *Pflugers Arch* 428: 631–640, 1994
  63. Pech V, Pham TD, Hong S, Weinstein AM, Spencer KB, Duke BJ, Walp E, Kim YH, Sutliff RL, Bao HF, Eaton DC, Wall SM: Pendrin modulates ENaC function by changing luminal HCO<sub>3</sub>. *J Am Soc Nephrol* 21: 1928–1941, 2010
  64. Koeppen BM, Helman SI: Acidification of luminal fluid by the rabbit cortical collecting tubule perfused in vitro. *Am J Physiol* 242: F521–F531, 1982
  65. Silver RB, Mennitt PA, Satlin LM: Stimulation of apical H-K-ATPase in intercalated cells of cortical collecting duct with chronic metabolic acidosis. *Am J Physiol* 270: F539–F547, 1996
  66. Wu MS, Biemesderfer D, Giebisch G, Aronson PS: Role of NHE3 in mediating renal brush border Na<sup>+</sup>-H<sup>+</sup> exchange: Adaptation to metabolic acidosis. *J Biol Chem* 271: 32749–32752, 1996
  67. Ambuhl PM, Amemiya M, Danczkay M, Lotscher M, Kaissling B, Moe OW, Preisig PA, Alpern RJ: Chronic metabolic acidosis increases NHE3 protein abundance in rat kidney. *Am J Physiol* 271: F917–F925, 1996
  68. Cogan MG, Rector FC Jr: Proximal reabsorption during metabolic acidosis in the rat. *Am J Physiol* 242: F499–F507, 1982
  69. Wang T, Egbert AL Jr, Aronson PS, Giebisch G: Effect of metabolic acidosis on NaCl transport in the proximal tubule. *Am J Physiol* 274: F1015–F1019, 1998
  70. Schambelan M, Sebastian A, Katuna BA, Arteaga E: Adrenocortical hormone secretory response to chronic NH<sub>4</sub>Cl-induced metabolic acidosis. *Am J Physiol* 252: E454–E460, 1987
  71. Loon NR, Wilcox CS: Mild metabolic alkalosis impairs the natriuretic response to bu-
  - metanide in normal human subjects. *Clin Sci* 94: 287–292, 1998
  72. Sebastian A, McSherry E, Morris RC Jr: Renal potassium wasting in renal tubular acidosis (RTA): Its occurrence in types 1 and 2 RTA despite sustained correction of systemic acidosis. *J Clin Invest* 50: 667–678, 1971
  73. Pines KL, Mudge GH: Renal tubular acidosis with osteomalacia: Report of 3 cases. *Am J Med* 11: 302–311, 1951
  74. Gill JR Jr, Bell NH, Bartter FC: Impaired conservation of sodium and potassium in renal tubular acidosis and its correction by buffer anions. *Clin Sci* 33: 577–592, 1967
  75. Bergenstal RM: Diabetic ketoacidosis: How to treat and, when possible, prevent. *Postgrad Med* 77: 151–157, 161, 1985
  76. Malnic G, De Mello Aires M, Giebisch G: Potassium transport across renal distal tubules during acid-base disturbances. *Am J Physiol* 221: 1192–1208, 1971
  77. Brackett NC Jr, Wingo CF, Muren O, Solano JT: Acid-base response to chronic hypercapnia in man. *N Engl J Med* 280: 124–130, 1969
  78. Gennari FJ, Goldstein MB, Schwartz WB: The nature of the renal adaptation to chronic hypocapnia. *J Clin Invest* 51: 1722–1730, 1972
  79. Malnic G, De Mello Aires M, Giebisch G: Micropuncture study of renal tubular hydrogen ion transport in the rat. *Am. J. Physiol.* 222: 147–158, 1972
  80. Kassirer JP, Schwartz WB: The response of normal man to selective depletion of hydrochloric acid: Factors in the genesis of persistent gastric alkalosis. *Am J Med* 40: 10–18, 1966
  81. Tresguerres M, Buck J, Levin LR: Physiological carbon dioxide, bicarbonate, and pH sensing. *Pflugers Arch* 460: 953–964, 2010