Effects of pH on Potassium: New Explanations for Old Observations

Peter S. Aronson*† and Gerhard Giebisch†

*Department of Internal Medicine and †Department of Cellular and Molecular Physiology, Yale University School of Medicine, New Haven, Connecticut

ABSTRACT

Maintenance of extracellular K⁺ concentration within a narrow range is vital for numerous cell functions, particularly electrical excitability of heart and muscle. Potassium homeostasis during intermittent ingestion of K⁺ involves rapid redistribution of K⁺ into the intracellular space to minimize increases in extracellular K⁺ concentration, and ultimate elimination of the K⁺ load by renal excretion. Recent years have seen great progress in identifying the transporters and channels involved in renal and extrarenal K⁺ homeostasis. Here we apply these advances in molecular physiology to understand how acid-base disturbances affect serum potassium.

Effects of Acid-Base Status on Intracellular K⁺ Distribution

Most of the body K⁺ content resides in the intracellular space of skeletal muscle.² An overview of ion transport pathways that directly or indirectly mediate shifts of K⁺ between muscle cells and the extracellular space in response to acid-base changes is shown in Figure 2.

Muscle contraction is triggered by action potentials involving depolarizing Na⁺ entry through Na⁺ channels followed by membrane repolarization mediated by K⁺ efflux through K⁺ channels. Cl⁻ channels play an important role in stabilizing the membrane potential and contributing to repolarization after action potentials. The electrochemical gradients of Na⁺ and K⁺ are restored by active Na⁺ extrusion and K⁺ uptake by the Na⁺,K⁺-ATPase.⁵ Accordingly, cell ion content is determined by the balance between pump and leak pathways for Na⁺ and K⁺.

However, muscle cells have additional pathways regulating intracellular pH homeostasis that can indirectly affect cellular Na⁺ and K⁺ balance.⁶ Quantitatively, the most important pathway regulating intracellular pH in skeletal muscle is Na⁺-H⁺ exchange,⁷ as shown in Figure 2. Na⁺-H⁺ exchange in skeletal muscle is highly dependent on intracellular pH, with marked activation by in-
tracellular acidity and inhibition by alkalinity. Activity of this pathway in response to acid-base perturbations strongly affects intracellular Na\(^{+}\)/H\(^{+}\) loading. Na\(^{+}\)/H\(^{+}\)-HCO\(_3\)-/H\(^{2+}\) exchanger isoform NHE1 is expressed in skeletal muscle and presumably accounts for most Na\(^{+}\)/H\(^{+}\)-HCO\(_3\)-/H\(^{2+}\) exchange activity in this tissue. A lesser component of intracellular pH regulation in skeletal muscle is HCO\(_3\)-dependent, because of Cl\(^{-}\)-HCO\(_3\) exchange, also shown in Figure 2. In addition, isoforms of the Na\(^{+}\)-bicarbonate cotransporter, NBCe1 and NBCe2, are expressed in muscle, raising the possibility that Na\(^{+}\)-HCO\(_3\) cotransport contributes to intracellular pH regulation, as also indicated in Figure 2. Another pathway of potential importance for cellular acid-base homeostasis is monocarboxylate cotransport that mediates coupled flux of H\(^{+}\) with such organic anions as lactate (Figure 2). Monocarboxylate cotransporters, MCT1 and MCT4, are expressed in skeletal muscle. During conditions like lactic acidosis, this pathway will mediate influx of H\(^{+}\) and lactate, resulting in decreased intracellular pH. Cation-chloride cotransport pathways are also present. Expression of K\(^{+}\)-Cl\(^{-}\) cotransporters KCC1, KCC3, and KCC4, as well as Na\(^{+}\)-K\(^{+}\)-Cl\(^{-}\) cotransporter NKCC1, has been detected in skeletal muscle. Interaction of K\(^{+}\)-Cl\(^{-}\) cotransport with acid-base transport will be discussed later.

Acute effects of acid-base disturbances on K\(^{+}\) redistribution have long been known. In general, metabolic acidosis with acidemia causes a net shift of K\(^{+}\) from the intracellular to the extracellular space. Conversely, net cellular uptake of K\(^{+}\) is observed in metabolic alkalosis with alkalemia. The directional effects of acidemia and alkalosis on K\(^{+}\) redistribution are similar in respiratory acid-base disturbances as in metabolic derangements, but the effects of respiratory disorders on K\(^{+}\) redistribution tend to be smaller than metabolic acid-base disturbances.

How can these effects of acid-base disturbances on K\(^{+}\) redistribution be explained in terms of the underlying cellular transport mechanisms? The general effect of acidemia to cause K\(^{+}\) loss from cells is often attributed to membrane K\(^{+}\)-H\(^{+}\) exchange. However, directly coupled K\(^{+}\)-H\(^{+}\) exchange is undetected in skeletal muscle. Nevertheless, reduction of extracellular pH results in net loss of K\(^{+}\) even from isolated muscle, indicating this phenomenon is at least in part intrinsic to muscle and independent of changes in hormonal milieu as might

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**Figure 1.** K\(^{+}\) concentration in the extracellular fluid (ECF) is affected by dietary intake, exchange with the intracellular fluid (ICF), and urinary excretion.

**Figure 2.** Multiple ion transport pathways directly or indirectly affect net K\(^{+}\) flux in skeletal muscle cells.
occur in vivo. What then explains the apparent K\(^{+}\)-H\(^{+}\) exchange?

As illustrated in Figure 3, the multiple acid-base transport pathways mentioned above may give rise to apparent K\(^{+}\)-H\(^{+}\) exchange. In the case of the predominant pH regulatory pathway, Na\(^{+}\)-H\(^{+}\) exchange, Na\(^{+}\) that enters by this route must be extruded by the Na\(^{+}\),K\(^{+}\)-ATPase (Figure 3A). Accordingly, K\(^{+}\) uptake by the Na\(^{+}\),K\(^{+}\)-ATPase will be greater when Na\(^{+}\)-H\(^{+}\) exchange activity is stimulated and will be diminished when the rate of Na\(^{+}\)-H\(^{+}\) exchange is reduced. In the case of acidosis with acidemia, the fall in extracellular pH would result in inhibition of the rate of Na\(^{+}\)-H\(^{+}\) exchange, leading to the accumulation of intracellular H\(^{+}\) and a decline in intracellular Na\(^{+}\). The latter would result in reduced Na\(^{+}\),K\(^{+}\)-ATPase activity, leading to decreased active cellular K\(^{+}\) uptake to counteract passive K\(^{+}\) efflux through K\(^{+}\) channels.\(^{20}\) The final result would be as if H\(^{+}\) had entered the cell in exchange for K\(^{+}\).

Similarly, as illustrated in Figure 3B, Na\(^{+}\)-HCO\(_{3}^{-}\) cotransport operating in parallel with Na\(^{+}\),K\(^{+}\)-ATPase may result in K\(^{+}\)-HCO\(_{3}^{-}\) cotransport, which is equivalent to K\(^{+}\)-H\(^{+}\) exchange. For example, in the case of metabolic acidosis with acidemia, the fall in extracellular HCO\(_{3}^{-}\) results in inhibition of the inward rate of Na\(^{+}\)-bicarbonate cotransport, leading to a fall in intracellular Na\(^{+}\) and reduced Na\(^{+}\),K\(^{+}\)-ATPase activity. Lower Na\(^{+}\),K\(^{+}\)-ATPase activity would cause a net loss of cellular K\(^{+}\). Again, the result would be as if H\(^{+}\) had entered the cell in exchange for K\(^{+}\).

Finally, Cl\(^{-}\)-HCO\(_{3}^{-}\) exchange also may contribute to apparent K\(^{+}\)-H\(^{+}\) exchange if operating in parallel with K\(^{+}\)-Cl\(^{-}\) cotransport, as shown in Figure 3C. Metabolic acidosis with a fall in extracellular HCO\(_{3}^{-}\) would increase the inward movement of Cl\(^{-}\) by Cl\(^{-}\)-HCO\(_{3}^{-}\) exchange. The resulting rise in intracellular Cl\(^{-}\) would then promote K\(^{+}\) efflux by K\(^{+}\)-Cl\(^{-}\) cotransport. The net result would be K\(^{+}\) efflux along with HCO\(_{3}^{-}\), which is an equivalent process to exchanging intracellular K\(^{+}\) for extracellular H\(^{+}\).

A striking observation has been that metabolic acidosis caused by mineral acid (hyperchloremic, nongap acidosis) causes a much larger shift of K\(^{+}\) into the extracellular fluid than does organic acidosis (lactic acidosis).\(^{21}\) The effect of hydrochloric acid but not organic acids to release K\(^{+}\) into the extracellular space had been observed using isolated muscle preparations, indicating this phenomenon can occur independently of systemic factors.\(^{22}\) In the case of acidemia caused by an organic acidosis like lactic acidosis, there would again be the effect of both low extracellular pH and HCO\(_{3}^{-}\) tending to inhibit Na\(^{+}\)-H\(^{+}\) exchange and Na\(^{+}\)-bicarbonate cotransport. This is illustrated for the case of Na\(^{+}\)-H\(^{+}\) exchange in Figure 4, but in contrast to the situation with hyperchloremic acidosis, there would also be a strong inward flux of lactate and H\(^{+}\) through the monocarboxylate transporter, resulting in a larger fall in intracellular pH and HCO\(_{3}^{-}\). The decrease in intracellular pH and HCO\(_{3}^{-}\) would tend to stimulate Na\(^{+}\) entry by Na\(^{+}\)-H\(^{+}\) exchange and Na\(^{+}\)-HCO\(_{3}^{-}\) cotransport, stimulating Na\(^{+}\),K\(^{+}\)-ATPase activity. The net effect would be to drive net cellular uptake of K\(^{+}\).

Thus, as illustrated in Figure 4, extracellular and intracellular acidosis are predicted to have opposing effects on the distribution of K\(^{+}\) because of their differing effects on cellular Na\(^{+}\) loading. During organic acidosis, there will be greater cellular acidification and Na\(^{+}\) entry than during hyperchloremic acidosis, resulting in higher Na\(^{+}\),K\(^{+}\)-ATPase ac-

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**Figure 4.** Opposing effects of extracellular and intracellular pH modify the influence of organic acidosis on plasma K\(^{+}\).
tivity compared with hyperchloremic acidosis. However, in several tissues, Na⁺,K⁺-ATPase activity is affected by intracellular pH, with reduced activity when intracellular pH is lower than normal. For intracellular acidification to stimulate net K⁺ uptake, it would require that the effect of low intracellular pH to inhibit Na⁺,K⁺-ATPase activity is less significant than the effect of intracellular Na⁺ loading to stimulate pump activity.

The acid-base mechanisms illustrated in Figures 2 and 3 also provide a possible explanation for the observation that bicarbonate can affect K⁺ redistribution independent of the effect of extracellular pH. Na⁺ entry by Na⁺-HCO₃⁻ cotransport would be enhanced when extracellular HCO₃⁻ is increased, resulting in increased cell Na⁺ uptake, stimulation of Na⁺,K⁺-ATPase activity, and net cellular K⁺ uptake (Figure 3B). Conversely, inhibition of Na⁺-HCO₃⁻ cotransport when extracellular HCO₃⁻ is reduced leads to a net loss of cell K⁺. Analogously, the rate of Cl⁻ entry by Cl⁻-HCO₃⁻ exchange would be higher when extracellular HCO₃⁻ is reduced, increasing cell Cl⁻ and enhancing the exit of K⁺ by K⁺-Cl⁻ cotransport (Figure 3C). Conversely, Cl⁻ entry by Cl⁻-HCO₃⁻ exchange would be lower when extracellular HCO₃⁻ is increased, leading to reduced K⁺ efflux by K⁺-Cl⁻ cotransport.

Similar considerations may also account for the smaller shifts in K⁺ observed with respiratory acidosis compared with metabolic acidosis. In respiratory acidosis, there is a fall in extracellular pH, but extracellular bicarbonate is elevated. One would therefore expect that Na⁺-H⁺ exchange is inhibited as in metabolic acidosis with equivalent acidemia, but Na⁺-bicarbonate cotransport would not be reduced. Accordingly, as compared with metabolic acidosis, respiratory acidosis would be associated with a smaller decrement in intracellular Na⁺, less inhibition of Na⁺,K⁺-ATPase activity, and reduced net K⁺ loss from the cell. In addition, during respiratory acidosis with elevated pCO₂, rapid cell entry of CO₂ will acidify intracellular pH. As discussed above for the case of organic acidosis, acidification of intracellular pH, by stimulating Na⁺ entry by Na⁺-H⁺ exchange tends to enhance Na⁺,K⁺-ATPase activity and oppose a net loss of intracellular K⁺.

In view of the longstanding observations discussed above on K⁺ redistribution in acid-base disorders, one would expect alkalinization by HCO₃⁻ administration to be an effective modality for acute treatment of hyperkalemia. However, some investigators have failed to find an effect of HCO₃⁻ administration to lower plasma K⁺ in hyperkalemic patients. An effect of HCO₃⁻ administration to lower plasma K⁺ has been more striking in patients with more severe degrees of pre-existing acidosis than in those with only minimal reductions of plasma HCO₃⁻. One possible factor modifying the effect of extracellular HCO₃⁻ and pH on K⁺ distribution is the level of intracellular pH and HCO₃⁻. At any given extracellular pH and HCO₃⁻, Na⁺ entry by Na⁺-H⁺ exchange and Na⁺-bicarbonate cotransport is greater when intracellular pH and HCO₃⁻ are reduced, as discussed earlier. Patients with appreciable pre-existing metabolic acidosis would be expected to have lower intracellular pH and HCO₃⁻. This may account for the fact that the effect of HCO₃⁻ administration to reduce plasma K⁺ has been more striking in patients with pre-existing acidosis.

Effects of pH and HCO₃⁻ on internal K⁺ distribution may be modified by hormonal systems that affect cellular K⁺ uptake and release. For example, net cellular uptake of K⁺ is strongly stimulated by insulin because of increased Na⁺,K⁺-ATPase activity. There is evidence that stimulation of insulin secretion by acidosis diminishes the hyperkalemia otherwise resulting from acidosis. Moreover, differential effects of organic versus hyperchloremic acidosis on insulin and glucagon secretion may contribute to the differing effects of these forms of acidosis on plasma K⁺ as discussed earlier. Although skeletal muscle is the predominant source of intracellular K⁺ content, there is evidence that the effect of organic acid-induced insulin secretion on plasma K⁺ is mediated at least in part by hepatic K⁺ uptake. The interactions of acid-base disturbances with other hormonal systems are at present incompletely defined.

**Effects of Acid-Base on Renal K⁺ Excretion**

The geography of net K⁺ transport along the nephron is shown in Figure 5. The bulk of filtered potassium is reabsorbed in the proximal tubule, where its reabsorption is predominantly passive and paracellular. Such passive reabsorption ultimately depends on and is driven by fluid reabsorption, which in turn is a function of Na⁺ reabsorption in this highly water-permeable nephron segment. Accordingly, the proximal tubule reabsorbs K⁺ in approximate proportion to reabsorption of filtered Na⁺ and water, accounting for retrieval of some 70% of the filtered load of K⁺.

Most of the remaining fraction of filtered K⁺ (approximately 25%) is reabsorbed in the thick ascending limb by a transcellular route involving Na⁺-K⁺-Cl⁻ cotransport across the apical membrane and efflux by the Na⁺,K⁺-ATPase across the basolateral membrane. As a consequence, only about 5% of the filtered K⁺ reaches the distal tubule.

The critical sites in the nephron responsible for determining K⁺ excretion are the connecting tubule and collecting tubule. Under conditions of normal or elevated dietary K⁺ intake, these segments mediate net secretion of K⁺. However, under conditions of K⁺ depletion, net K⁺ reabsorption may occur in this portion of the nephron.

Cell models for K⁺ secretion and reabsorption are illustrated in Figure 6. K⁺ secretion, which takes place through connecting tubule cells and principal cells of the collecting tubule, involves active uptake across the basolateral membrane mediated by Na⁺,K⁺-ATPase, followed by passive exit across the apical membrane through K⁺ channels. The ENaC Na⁺ channel in the apical membrane indirectly plays a critical role in K⁺ secretion by providing intracellular Na⁺ as substrate for the Na⁺,K⁺-ATPase and by depolarizing the apical membrane, thereby increasing the driving force for

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K⁺ efflux into the tubule fluid. At least two types of apical K⁺ channels have been implicated in K⁺ secretion, ROMK and BK.

In addition, effects of luminal Cl⁻ on K⁺ secretion suggest that a component of apical membrane K⁺ efflux takes place by electroneutral K⁺-Cl⁻ cotransport,

although its molecular mechanism is unclear.

Multiple factors modulate K⁺ secretion in these distal nephron segments. First, plasma K⁺ concentration is an important determinant of the rate of K⁺ secretion. External K⁺ is substrate for the Na⁺,K⁺-ATPase, and there is saturable dependence of K⁺ secretion on plasma K⁺ concentration.

In addition, the higher the plasma K⁺, the less backflux of K⁺ takes place through basolateral K⁺ channels. Second, aldosterone, whose secretion is stimulated by elevated plasma K⁺, is a major regulator of K⁺ secretion in distal nephron segments. Aldosterone acts through several signaling pathways and effector mechanisms to bring about increased activities of apical Na⁺ and K⁺ channels and increased activity of basolateral Na⁺,K⁺-ATPase.

Third, K⁺ secretion is highly affected by luminal Na⁺ and fluid delivery to distal K⁺ secretory sites. K⁺ secretion is dependent directly and indirectly on luminal Na⁺ entry through ENaC, as discussed above, which in turn is a function of luminal Na⁺ concentration. K⁺ secretion is enhanced at higher luminal flow rates because of activation of BK channels. Recent studies suggest that flow activation of BK channels involves intracellular Ca²⁺ signaling most likely in response to mechanosensing by apical cilia. Finally, net K⁺ secretion is modified by modulation of active K⁺ reabsorption mediated by apical H⁺,K⁺-ATPase in intercalated cells.

Chronic K⁺ depletion causes upregulation of K⁺ reabsorption mediated by apical H⁺,K⁺-ATPase in the collecting tubule.

Acid-base disturbances have complex effects on renal K⁺ excretion. For example, acute acidemia caused by metabolic acidosis reduces renal K⁺ excretion, whereas metabolic acidosis at a later phase promotes urinary K⁺ excretion.
As discussed earlier for the case of muscle activity of Na\(^+\)-H\(^+\) exchange and Na\(^+\)-HCO\(_3\)\(^-\) cotransport can affect intracellular Na\(^+\) loading and Na\(^+\), K\(^+\)-AT-Pase activity. Changes in Na\(^+\), K\(^+\)-AT-Pase activity would affect basolateral K\(^+\) uptake and transtubular K\(^+\) secretion. In fact, basolateral Na\(^+\) entry by Na\(^+\)-H\(^+\) exchange can support K\(^+\) secretion in the absence of luminal Na\(^+\).\(^{58}\) Thus, inhibition of Na\(^+\)-H\(^+\) exchange and Na\(^+\)-HCO\(_3\)\(^-\) cotransport during metabolic acidosis with acidemia reduces cell Na\(^+\) and thereby inhibits K\(^+\) secretion. Conversely, metabolic alkalosis with alkalemia would tend to stimulate K\(^+\) secretion.

Several of the pathways involved in distal nephron K\(^+\) secretion are directly affected by pH, as illustrated in Figure 6. A fall in intracellular pH reduces activity of Na\(^+\), K\(^+\)-ATPase on the basolateral membrane\(^{23-25}\) and diminishes activities of ENaC, ROMK, and BK on the apical membrane.\(^{59-62}\) These effects could contribute to inhibition of active K\(^+\) secretion in response to acidemia to the extent that there is parallel intracellular acidosis. ENaC abundance is decreased when luminal or basolateral HCO\(_3\)\(^-\) (and pH) is reduced.\(^{63}\) In addition, stimulation of electrogenic H\(^+\) secretion by the vacuolar ATPase in intercalated cells during acidemia would tend to reduce the lumen-negative transepithelial potential difference,\(^{64}\) inhibiting K\(^+\) secretion. Furthermore, acidemia upregulates apical H\(^+\),K\(^+\)-ATPase in intercalated cells,\(^{65}\) thereby enhancing K\(^+\) reabsorption and reducing net K\(^+\) secretion.

However, metabolic acidosis causes increased distal Na\(^+\) delivery and flow rate, as well as increased urinary Na\(^+\) excretion.\(^{42,52}\) There are at least four possible factors contributing to this phenomenon. First, Na\(^+\),K\(^+\)-ATPase in all nephron segments may be inhibited by acidemia with reduced intracellular pH, leading to diminished renal tubular Na\(^+\) reabsorption upstream of the K\(^+\) secretory sites. Second, although acidosis causes upregulation of NHE3 activity in the proximal tubule,\(^{66,67}\) the absolute amount of Na\(^+\) reabsorbed with HCO\(_3\)\(^-\) in the proximal tubule is actually reduced in metabolic acidosis because of the diminished filtered load of HCO\(_3\)\(^-\).\(^{68}\) Third, the reduction in absolute fluid reabsorption coupled to NaHCO\(_3\) reabsorption in metabolic acidosis results in a less than normal increment in luminal Cl\(^-\) concentration along the length of the proximal tubule, lowering the driving force for passive paracellular NaCl reabsorption.\(^{69}\) Fourth, apical membrane Cl\(^-\)-base exchange activity is downregulated in metabolic acidosis, reducing tranacellular NaCl reabsorption in the proximal tubule.\(^{69}\) These effects of metabolic acidosis to enhance distal Na\(^+\) delivery and flow rate and to augment urinary Na\(^+\) excretion tend to cause volume depletion, resulting in increased renin and aldosterone secretion.\(^{70}\) In addition, acidosis may directly stimulate aldosterone secretion independent of renin secretion.\(^{70,71}\) As discussed above, K\(^+\) secretion in the distal nephron is strongly stimulated by...
aldosterone and by increased luminal Na⁺ delivery and flow rate. These factors eventually predominate over the local inhibitory effect of acidemia on the K⁺ secreting cells, resulting in increased K⁺ excretion and negative K⁺ balance. Ultimately, a new steady state is established as K⁺ balance is restored at the expense of hypokalemia.

This pathophysiology is well illustrated by the disorder of classic distal RTA. Although there are exceptions, these patients generally have hypokalemia caused by renal K⁺ wasting during chronic acidosis, which is corrected by base administration. Another clinical example in which the effect of distal Na⁺ delivery and flow to stimulate urinary K⁺ excretion predominates over the inhibitory effect of acidemia is diabetic ketoacidosis with osmotic diuresis. Moreover, organic acidosis with high rates of excretion of nonchloride anions in the urine will enhance distal K⁺ delivery and flow to stimulate urinary K⁺ excretion as does metabolic acidosis. Acute acidemia caused by respiratory acidosis inhibits renal K⁺ secretion in the distal nephron and reduces urinary K⁺ excretion. Over the following days, respiratory acidosis results in urinary K⁺ wasting associated with increased Na⁺ excretion. It is likely these effects are mediated by many of the same mechanisms discussed above for metabolic acidosis. However, the effects of chronic acidemia to augment K⁺ excretion caused by increased distal Na⁺ and fluid delivery will tend to be milder for respiratory acidosis than for metabolic acidosis. First, renal compensation of chronic respiratory acid-base disorders restores blood pH much closer to normal than does respiratory compensation of metabolic acid-base disorders. Thus, acidemia will be milder in chronic respiratory acidosis than in chronic metabolic acidosis, resulting in less inhibition of Na⁺, K⁺-ATPase and less augmentation of Na⁺ and fluid delivery to the distal nephron. Second, the role of low filtered load of HCO₃⁻ in limiting absolute NaHCO₃ reabsorption along the proximal tubule in metabolic acidosis as discussed above would not be a contributing factor in respiratory acidosis.

In general, alkalemia stimulates distal nephron K⁺ secretion and urinary K⁺ excretion. These effects are larger for metabolic than for respiratory alkalosis and are in essence the converse of the effects of acidemia on the K⁺ secretory pathways, because elevated intracellular pH tends to increase activities of ENaC, ROMK, and BK. ENaC abundance is increased when luminal or basolateral HCO₃⁻ (and pH) is elevated. Moreover, in the case of acute metabolic alkalosis, there is inhibition of fractional NaHCO₃ and fluid reabsorption in the proximal tubule, leading to increased distal delivery of Na⁺ and HCO₃⁻ and enhanced fluid flow. As already discussed, increased Na⁺ delivery and fluid flow stimulate K⁺ secretion. In addition, increased luminal delivery of HCO₃⁻ as a nonchloride anion stimulates a component of distal K⁺ secretion because of the reduced luminal Cl⁻ concentration as mentioned earlier. Of course, in the maintenance phase of metabolic alkalosis with volume contraction, distal Na⁺ and HCO₃⁻ delivery will decline, thereby mitigating the rate of K⁺ wasting.

Summary and Conclusions
We have used new information about the molecular physiology of extrarenal and renal potassium transport to explain longstanding observations of the complex effects of acid-base disturbances on serum potassium. It should be emphasized that many aspects of these explanations are somewhat speculative because the quantitative contributions of specific pathways to mediating effects of pH on extrarenal and renal K⁺ transport are not known with certainty. Moreover, we have not considered new developments concerning molecular sensors and signaling mechanisms that respond to changes in extracellular and intracellular pH, CO₂, and HCO₃⁻. The roles of these sensor and signaling mechanisms in mediating the effects of acid-base disturbances on extrarenal and renal K⁺ homeostasis are not yet known.

Nevertheless, it should be apparent that there is no simple relationship between pH and serum potassium because of the multiplicity of factors affecting internal and external K⁺ balance. For example, consider the patient with diabetic ketoacidosis. Acidemia will tend to shift K⁺ out of cells and cause hyperkalemia, but this effect is less pronounced in organic acidosis than in mineral acidosis. On the other hand, hypotonicity in the absence of insulin will promote K⁺ release into the extracellular space. Renal K⁺ excretion will be acutely inhibited by acidemia but ultimately enhanced by the increased distal Na⁺ delivery and flow rate caused by metabolic acidosis and osmotic diuresis in the setting of high aldosterone. Indeed, the patient may present with marked K⁺ depletion if osmotic diuresis has been going on for some time. Renal K⁺ excretion may later become reduced when GFR falls as volume depletion ensues. Accordingly, there will not be a straightforward relationship between serum potassium and acid-base status in such a patient. The clinician will need to be knowledgeable about the many factors affecting internal and external K⁺ balance to provide optimal patient care.

DISCLOSURES
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