Absence of MyD88 Signaling Induces Donor-Specific Kidney Allograft Tolerance

Huiling Wu,*† Gerda A. Noordmans,*† Maya R. O’Brien,*† Jin Ma,*† Cathy Y. Zhao,*† Geoff Y. Zhang,† Tony K.T. Kwan,*† Stephen I. Alexander,†‡ and Steven J. Chadban*†

*Renal Medicine and Transplant Research Group, Royal Prince Alfred Hospital, Sydney, New South Wales, Australia; †Faculty of Medicine, University of Sydney, Sydney, New South Wales, Australia; and ‡Children Hospital at Westmead, Westmead, New South Wales, Australia

ABSTRACT
Toll-like receptors (TLRs) play a fundamental role in innate immunity and provide a link between innate and adaptive responses to an allograft; however, whether the development of acute and chronic allograft rejection requires TLR signaling is unknown. Here, we studied TLR signaling in a fully MHC-mismatched, life-sustaining murine model of kidney allograft rejection. Mice deficient in the TLR adaptor protein MyD88 developed donor antigen-specific tolerance, which protected them from both acute and chronic allograft rejection and increased their survival after transplantation compared with wild-type controls. Administration of an anti-CD25 antibody to MyD88-deficient recipients depleted CD4+CD25+FoxP3+ cells and broke tolerance. In addition, defective development of Th17 immune responses both in vitro and in vivo occurred, resulting in an increased ratio of Tregs to Th17 effectors. Thus, MyD88 deficiency was associated with an altered balance of Tregs over Th17 cells, promoting tolerance instead of rejection. This study provides evidence that targeting innate immunity may be a clinically relevant strategy to facilitate transplantation tolerance.

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Correspondence: Dr. Huiling Wu, Transplant Research Group, Room W601, Blackburn Building D06, University of Sydney, Sydney, NSW 2006, Australia. Email: huiling.wu@sydney.edu.au

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RESULTS

Absence of MyD88 Signaling Prolongs Kidney Allograft Survival

We transplanted BALB/c.MyD88−/− (H2-Kb) kidneys into C57BL/6.MyD88−/− recipients (H2-Kb) as MyD88−/− allografts. Survival to 100 days was observed in all isografts. WT mice rejected their WT allografts with a mean graft survival of 40 days, whereas MyD88−/− allografts displayed prolonged survival (Figure 1A). When MyD88 was absent from recipients, five of five allografts survived to 100 days (Figure 1B). When only the allograft was MyD88-deficient, survival was modestly prolonged over WT allografts; however, only three of seven recipients survived to 100 days (Figure 1B).

Absence of MyD88 Signaling Protects against Acute Kidney Rejection at Day 14

Significant kidney dysfunction in WT allografts was evident and reflected by a tripling of serum creatinine compared with isografts (Figure 2A). Kidney function was preserved in MyD88−/− and recipient MyD88−/− donor MyD88+/− allografts equally, whereas only partial preservation of function was seen in recipient MyD88+/− donor MyD88−/− allografts compared with WT allografts (Figure 2A and Supplemental Figure 1A).

WT allografts exhibited severe tubulitis compared with isografts. Tubulitis was present in MyD88−/− allografts and allografts where either donor or recipient was MyD88−/−, although it was significantly diminished compared with WT allografts (Figure 2B and Supplemental Figure 1B).

Substantial infiltration of CD4+ and CD8+ cells was evident in WT allografts versus isografts (P<0.001) (Figure 2C). CD8+, although not CD4+, cell infiltration was attenuated in MyD88−/− allografts (Figure 2C). Extensive macrophage (CD68+) accumulation was evident in WT allografts compared with isografts; however, this accumulation was markedly diminished in MyD88−/− allografts (Figure 2C).

Reduced Expression of Proinflammatory and Th1 Cytokines and Cytotoxic Molecules in MyD88−/− Allografts at Day 14

WT allografts showed strong upregulation of proinflammatory cytokines (TNF-α, IL-6, and TGF-β) and cytokines (IFN-γ, IL-4, and IL-17) compared with isografts (Figure 2D). All proinflammatory cytokines and IFN-γ were reduced in MyD88−/− allografts compared with WT allografts (Figure 2D), whereas upregulation of IL-4 was of a similar magnitude to the magnitude in WT allografts. MyD88−/− allografts showed a nonsignificant trend to reduced expression of IL-17 compared with WT allografts (P=0.06) (Figure 2D). Cytotoxic molecules (perforin, granzyme, and inducible nitric oxide synthase) are mediators of kidney damage during acute rejection. All were upregulated in WT allografts compared with isografts but attenuated in the absence of MyD88 (Figure 2D).

MyD88 Deficiency Is Protective against Chronic Allograft Injury at Day 100 Post-Transplantation

By day 100 post-transplantation, surviving WT mice had developed significant kidney allograft dysfunction, which was evidenced by increased serum creatinine and proteinuria for WT allografts versus isografts (Figure 3, A and B). Kidney function remained normal in MyD88−/− allografts and was comparable with kidney function of isografts. Proteinuria in MyD88−/− allograft mice was similar to isografts and significantly less than WT allografts (Figure 3B).

WT allografts developed severe glomerulosclerosis, interstitial fibrosis, and tubular atrophy, whereas these changes were absent in isografts and attenuated in MyD88−/− allografts (Figure 3). To verify interstitial fibrosis, we stained collagen and α-smooth muscle actin (α-SMA). The expression of interstitial collagen and α-SMA was significantly higher in WT allografts versus isografts and significantly reduced in MyD88−/− allografts (Figure 3C).

Significant CD4+ and CD8+ cell accumulation was evident in WT allografts compared with isografts (P<0.05), with a modest reduction in MyD88−/− mice compared with WT (Figure 3C). There was a significant increase in CD68+ in WT allografts compared with isografts (P<0.01), which was attenuated in MyD88−/− allografts (Figure 3C). There was no significant difference between isografts and MyD88−/− mice.

IL-12, and IL-23). After organ transplantation, these TLR-driven innate immune responses are important for the promotion of ischemia reperfusion injury (IRI) and the initiation of adaptive alloimmunity, which causes acute and chronic rejection.

Significant and nonredundant roles have been shown for TLR2 and TLR4 signaling in kidney IRI. We previously reported that IRI resulted in upregulation of TLR4 and its endogenous ligands high-mobility group protein B1 (HMGB1), hyaluronan, and biglycan within the kidney and that TLR4 activation through the MyD88 pathway is a key mediator of kidney IRI. We subsequently showed potential for blockade of the endogenous TLR ligand HMGB1 through TLR4 signaling to prevent kidney IRI.

In humans, the presence of a hypofunctioning TLR4 polymorphism has been associated with protection from IRI, manifesting as a reduced incidence of delayed graft function after kidney transplantation.

Given the importance of TLR/MyD88 signaling in kidney IRI, the association between reduced TLR4 function and improved outcomes in human kidney transplantation, and the emerging roles for innate immunity in modulating adaptive immunity, we hypothesized that TLR signaling through MyD88 is required for the development of kidney allograft rejection. In the current set of experiments, we compared MyD88−/− and wild-type (WT) mice after receipt of life-sustaining fully MHC-mismatched kidney allografts. We herein present data that show how the absence of TLR signaling by MyD88 protected mice from acute and chronic rejection through the development of donor-specific tolerance.

BASIC RESEARCH
BASIC RESEARCH

Cytokines, Chemokines, and Fibrosis-Related Genes

MyD88

WT allografts compared with isografts (\(P \leq 0.01\)) and these results were attenuated in MyD88\(^{-/-}\) allografts (Figure 4).

Donor Antigen-Specific Immune Tolerance in the Absence of MyD88

To test for immune tolerance, a subset of both WT and MyD88\(^{-/-}\) kidney allograft acceptors received skin grafts from BALB/c (donor-matched allograft), C57BL/6 (isograft), and B10Br (third-party allograft) mice at day 150 after kidney transplant. WT mice rejected skin allografts from both donor-matched BALB/c and third-party B10Br mice at 12–14 days (Figure 5, A and B). MyD88\(^{-/-}\) recipients accepted donor-matched skin allografts for over 75 days after skin transplant but rejected B10Br third-party allografts by day 15 (Figure 5, A and B). Skin isografts on WT and MyD88\(^{-/-}\) kidney allograft recipients remained intact. Placement of skin grafts disrupted kidney graft acceptance in WT recipients; all of the recipients died between 34 and 107 days after skin allograft challenge, whereas MyD88\(^{-/-}\) recipients retained normal kidney function, which was evidenced by normal serum creatinine (13.3±3.1 μmol/L) when euthanized at day 180 after skin grafting. Consistent with donor antigen-specific tolerance, MyD88\(^{-/-}\) splenocytes harvested from an MyD88\(^{-/-}\)–allograft acceptor at day 100 after kidney transplantation stimulated with donor-matched allogeneic spleen cells produced significantly less IFN-γ than similarly stimulated WT splenocytes from a WT allograft acceptor as assessed by ELISPOT (Figure 5C).

Absence of MyD88 Signaling Results in Reduced mRNA Expression of Proinflammatory and Th1 Cytokines, Chemokines, and Fibrosis-Related Genes

Expression of proinflammatory cytokines (TNF-α, IL-6, and IL-1β) was upregulated in WT allografts compared with isografts (\(P < 0.01\)) and partially reduced in MyD88\(^{-/-}\) allografts compared with WT (Figure 4). IFN-γ mRNA was increased in WT allografts compared with isografts (\(P < 0.001\)) but not MyD88\(^{-/-}\) allografts (Figure 4). IL-4 mRNA expression in WT and MyD88\(^{-/-}\) allografts was higher than in isografts (Figure 4). IL-17 expression was upregulated in WT allografts compared with isografts but not MyD88\(^{-/-}\) allografts (\(P = 0.08\)) (Figure 4). Expression of chemokines (CCL2, chemokine [C-X-C motif] ligand 9 [CXCL9], and CXCL10) was significantly higher in WT mice compared with isografts (\(P < 0.05\)) (Figure 4). MyD88\(^{-/-}\) mice showed significantly reduced expression of CCL2 and CXCL9 compared with WT mice (Figure 4), consistent with our findings by immunohistology of a reduction in infiltrating CD68+ cells in MyD88\(^{-/-}\)–allografts. WT allografts showed a higher expression of fibrosis-related genes (TGF-β, matrix metalloproteinase 2 [MMP2], and tissue inhibitor of metalloproteinase 1 [TIMP1]) compared with isografts (\(P < 0.01\)).

Allograft Tolerance in the Absence of MyD88 Signaling Is Dependent on a CD4+CD25+FoxP3+ Regulatory Mechanism

We hypothesized that long-term allograft tolerance in the absence of MyD88 signaling may be dependent on immune regulation mediated by CD4+CD25+ regulatory cells (Tregs). To test this hypothesis, we treated MyD88\(^{-/-}\) mice with an anti-CD25 mAb. This depleted CD4+CD25+FoxP3+ cells by >90% and abrogated graft acceptance (Figure 6A). Administration of anti-CD25 mAb to WT allograft recipients did not alter rejection kinetics (Figure 6B).

We next sought to determine whether tolerance was transferrable. CD4+CD25+ Tregs were isolated and purified from long-term C57BL/6.MyD88\(^{-/-}\)–acceptors and adoptively co-transferred with WT CD4+CD25+ T effectors (Teffs) into C57BL/6.RAG\(^{-/-}\) recipients that were previously transplanted with a BALB/c skin graft. Skin grafts showed long-term survival in four of five mice compared with 100% graft loss from mice that received either WT CD4+CD25+ Teffs alone or with C57BL/6.MyD88\(^{-/-}\)–CD4+CD25+ Teffs (Figure 6C).

Figure 1. Absence of MyD88 signaling prolongs kidney allograft survival. (A) Absence of MyD88 signaling in both donor and recipient prolongs kidney allograft survival. Survival to 100 days was observed in all isografts (n=6) and 15 of 17 MyD88\(^{-/-}\) allografts (n=17) but only 12 of 31 WT allografts (n=31; ***\(P < 0.001\) WT allografts versus MyD88\(^{-/-}\) allografts and isografts). (B) When MyD88 was absent from recipients, five of five allografts survived to 100 days (n=5; \(P = 0.05\), WT allografts versus MyD88\(^{-/-}\) recipients). When only the allograft was MyD88-deficient, three of seven WT recipients survived to 100 days (n=7).

In human kidney transplantation, complement split product C4d deposition in peritubular capillaries (PTCs) is a marker of antibody-mediated rejection and is associated with development of transplant glomerulopathy. Prominent C4d staining was present diffusely in almost all cross-sections of PTCs, with a bright, broad linear pattern in WT allografts (Figure 3C). Fluorescence intensity of C4d deposition in PTCs was significantly reduced in MyD88\(^{-/-}\) allografts. Consistent with this result, WT allograft recipients exhibited higher titers of donor-specific antibody, both IgG2a and IgG1 subclasses, compared with MyD88\(^{-/-}\) allografts (Supplemental Figure 2).

Absence of MyD88 Signaling Results in Reduced mRNA Expression of Proinflammatory and Th1 Cytokines, Chemokines, and Fibrosis-Related Genes

Expression of proinflammatory cytokines (TNF-α, IL-6, and IL-1β) was upregulated in WT allografts compared with isografts (\(P < 0.01\)) and partially reduced in MyD88\(^{-/-}\) allografts compared with WT (Figure 4). IFN-γ mRNA was increased in WT allografts compared with isografts (\(P < 0.001\)) but not MyD88\(^{-/-}\) allografts (Figure 4). IL-4 mRNA expression in WT and MyD88\(^{-/-}\) allografts was higher than in isografts (Figure 4). IL-17 expression was upregulated in WT allografts compared with isografts but not MyD88\(^{-/-}\) allografts (\(P = 0.08\)) (Figure 4). Expression of chemokines (CCL2, chemokine [C-X-C motif] ligand 9 [CXCL9], and CXCL10) was significantly higher in WT mice compared with isografts (\(P < 0.05\)) (Figure 4). MyD88\(^{-/-}\) mice showed significantly reduced expression of CCL2 and CXCL9 compared with WT mice (Figure 4), consistent with our findings by immunohistology of a reduction in infiltrating CD68+ cells in MyD88\(^{-/-}\)–allografts. WT allografts showed a higher expression of fibrosis-related genes (TGF-β, matrix metalloproteinase 2 [MMP2], and tissue inhibitor of metalloproteinase 1 [TIMP1]) compared with isografts (\(P < 0.01\)), and these results were attenuated in MyD88\(^{-/-}\) allografts (Figure 4).
Figure 2. Absence of MyD88 signaling was protective against acute rejection. (A) Loss of kidney function, as indicated by a significant elevation of serum creatinine, was evident in WT allografts (n=12) but not MyD88−/− allografts (n=9) at day 14 post-transplant compared with isografts (n=5; ***P<0.001). (B) Tubulitis was present in MyD88−/− allografts (n=7), although it was significantly diminished compared with WT allografts (n=9; ***P<0.001). (C) Interstitial infiltrates in the allografts at day 14 after kidney transplant.
Increased Ratio of Tregs/Th17 Teffs Cells in MyD88−/− Allografts

There was a significant increase in CD4+FoxP3+ cells as a percentage of total CD4+ splenocytes obtained from WT allograft recipients, and to a greater extent, there was a significant increase in MyD88−/− allograft recipients compared with isograft recipients at day 14 post-transplant (Figure 7A). There was no significant difference in the percentages of CD4+ and CD8+ splenocytes producing either IL-17 or IFN-γ obtained from WT or MyD88−/− allograft recipients (Supplemental Figure 3).

We isolated CD4+ and CD8+ lymphocytes from kidney grafts at day 14. CD4+FoxP3+ Tregs were significantly increased in MyD88−/− allografts compared with WT allografts (Figure 7A). The percentages of CD4+ and CD8+ cells producing IL-17 obtained from WT allografts were significantly higher than from MyD88−/− allografts (Figure 7B), whereas the percentage of T cells producing IFN-γ was similar in both (Figure 7C).

To confirm the role of Th17 in kidney allograft rejection in our model, we transplanted BALB/c.IL17−/− (H-2Kb) kidneys into C57BL/6.IL17−/− recipients (H2-Kb) as IL17−/− allografts. IL17−/− allografts displayed prolonged survival compared with WT allografts (P<0.05; data not shown).

Absence of MyD88 Impairs Allroactive Th17 Development, Resulting in an Increased Ratio of Tregs/Th17 Teffs Cells in Mixed Lymphocyte Cultures

We examined whether the absence of MyD88 would produce less IL-6 and consequently, an increased ratio of Treg/Th17 cells in an alloreactive mixed lymphocyte culture under conditions promoting either iTreg or Th17 differentiation.

CD4+ T cells obtained from naive C57BL/6.WT or C57BL/6.MyD88−/− mice, stimulated with BALB/c.WT or BALB/c. MyD88−/− allogenic splenocytes in the absence of exogenous TGF-β and IL-6, did not result in the induction of either Foxp3+ Tregs or IL-17 Teffs (Figure 8A). The addition of TGF-β to WT and MyD88−/− CD4+ T cells in culture resulted in the generation of Foxp3+ Tregs but not Th17 cells, whereas the addition of IL-6 in the presence of TGF-β to WT CD4+ T cells promoted Th17 differentiation (Figure 8, B and C). C57BL/6.MyD88−/−/CD4+ T cells stimulated with BALB/c. MyD88−/− allogenic splenocytes in the presence of exogenous TGF-β generated a comparable number of Foxp3+ Tregs compared with WT (Figure 8B), whereas MyD88−/−/CD4+ T cells in the presence of TGF-β plus IL-6 generated significantly smaller numbers of IL-17–producing T cells compared with WT (Figure 8C). These results suggest that absence of MyD88 impairs alloreactive Th17 differentiation, resulting in an increased ratio of Tregs/Th17 Teffs in the alloreactive culture system in vitro.

MyD88−/− Mice Exhibit Fewer Donor-Specific Th17 Immune Responders as Assessed by ELISPOT

To investigate whether MyD88−/−-tolerant mice generated fewer donor-specific Th17 cells, we measured T cell production of IFN-γ, IL-4, and IL-17 in response to allostimulation by ELISPOT assay. WT and MyD88−/− splenocytes, primed and stimulated with allogeneic cells, produced higher numbers of donor-reactive IFN-γ spots than splenocytes similarly stimulated with syngenic or third-party (B10Br) cells (Figure 9A). IL-4 production was not different between WT and MyD88−/− spleen cells (Figure 9B). The number of IL-17–producing cells was significantly greater in WT compared with MyD88−/− splenocytes after donor allostimulation (Figure 9C), with no significant difference after stimulation by either isogenic or third-party control cells.

DISCUSSION

IRI is an inevitable consequence of kidney implantation, and it leads to rapid activation of an innate immune response causing kidney injury. In the present study, we found that expression of both TLR2 and -4 and their endogenous ligands, including HMGB1, hyaluronan, and biglycan, was similarly increased in kidney grafts after transplantation (Supplemental Figure 4). These results, taken together with observational studies showing TLR4 and ligand upregulation within hours of human transplantation,16 suggest that TLR activation is a mediator of the innate immune response within the newly transplanted kidney.

Modulation of the graft microenvironment by this early innate response may be a prerequisite for the full development of adaptive allograft immunity and subsequent allograft rejection. In an allosensitized mouse model, allografts were acutely rejected in the presence of inflammation caused by ischemia reperfusion but not in its absence.6 In humans, observational studies have shown improved outcomes of lung and kidney allografts in patients with hypofunctioning TLR4 polymorphisms.17–19 Experimental work has shown that, although the absence of MyD88 signaling was sufficient to prevent minor MHC mismatched skin graft rejection,20 survival of fully mismatched allogeneic skin and heart transplants was not

Representative sections of kidney grafts at day 14 post-transplant were stained for CD4, CD8, CD68, and CD11c by immunochemistry. Substantial infiltration of CD4+ cells was evident in WT and MyD88−/− allografts versus isografts (P<0.001). CD8+ cell infiltration was attenuated in MyD88−/−/allografts versus WT allografts (P<0.05). Significant macrophage (CD68+) accumulation was evident in the WT allografts compared with isografts but significantly reduced in MyD88−/−/allografts (P<0.01). (D) Absence of MyD88 signaling resulted in reduced miRNA expression of proinflammatory and Th1 cytokines and cytotoxic molecules at day 14 after kidney transplant. *P<0.05; **P<0.01. Data are presented as mean ± SD. Isografts are n=5, WT allografts are n=9, and MyD88−/− allografts are n=7.


MyD88 and Kidney Allograft Tolerance
Figure 3. MyD88 deficiency prevents the allograft from chronic rejection at day 100 after kidney transplant. (A) Serum creatinine was significantly lower in MyD88^{-/-} allografts than WT allografts (**P<0.01). (B) Proteinuria was significantly diminished in MyD88^{-/-} allografts compared to WT allografts (**P<0.01).
prolonged.\textsuperscript{21} A recent study reported that recipient MyD88 contributes to chronic graft dysfunction.\textsuperscript{22}

Against this background, the importance of MyD88 signaling in the development of kidney allograft rejection was not fully understood. The current study found that mice geneti-
cally deficient in MyD88 developed donor-specific tolerance across a full MHC mismatch that was robust, which was shown by acceptance of donor-matched skin but rejection of third-
party grafts. Our observations that CD25 depletion was suf-
cient to break tolerance and that tolerance was associated with impaired Th17 responses to alloantigen provide new evidence highlighting the extent to which adaptive alloimmunity may be controlled by the innate immune system.

TLRs can trigger activation of several arms of the adaptive immune response, generating effectors including Ig, Th1 and Th17 CD4\textsuperscript{+} effector T cells, and CD8\textsuperscript{+} effector T cells. The MyD88 pathway is known to be important in generating Th1 responses.\textsuperscript{12} Studies in allotransplant models seem par-
tially consistent with this data. Gene expression of IFN-\gamma but not IL-4 was diminished in draining lymph nodes retrieved from MyD88\textsuperscript{-/-} skin allograft recipients compared with WT recipients.\textsuperscript{20} In the present study, we found that IFN-\gamma mRNA expression was significantly reduced in MyD88\textsuperscript{-/-} allografts compared with WT allografts. How-
ev, IL-4 mRNA expression was comparable, and CD4\textsuperscript{+} and CD8\textsuperscript{+} T cells isolated from WT and MyD88\textsuperscript{-/-} kidney allografts at day 14 post-transplant contained comparable numbers of IFN-\gamma-producing T cells. Consistent with this result, recent experimental data in transplantation has pro-
voked evidence that MyD88 signaling is not critical for allo-
geneic dendritic cells (DCs) or APCs to activate alloprimed T cells,\textsuperscript{21} although MyD88 deficiency led to a reduction in allo-
geneic T cell priming and consequent Th1 immunity in a skin transplant model.\textsuperscript{21}

Although IFN-\gamma is an essential component of Th1-driven immune responses, it is also a critical cytokine in tolerance. IFN-\gamma has been shown to act as a protective factor against the onset of experimental autoimmune encephalomyelitis,\textsuperscript{23} and IFN-\gamma is specifically required for the induction of tolerance to alloantigens in several transplantation models.\textsuperscript{24–27} IFN-\gamma seems to be required for the generation and function of allo-
specific Tregs during the development of operational tolerance to donor alloantigens in vivo.\textsuperscript{28} IFN-\gamma conditioning ex vivo generates CD25\textsuperscript{+}CD62L\textsuperscript{-}FoxP3\textsuperscript{+} Tregs that prevent allograft rejection\textsuperscript{29} and shapes the alloreactive T cell repertoire by inhibiting Th17 responses and generating functional FoxP3\textsuperscript{+} Tregs.\textsuperscript{30}

Activation of TLRs may regulate the development of Th17 cells. TLR signaling through MyD88 leads to the production of proinflammatory cytokines including IL-6, a potent Th1 differentiation cytokine. Naïve T cells in the presence of IL-6 combined with TGF-\beta differentiate into Th17. Th17 cells are important for the clearance of pathogens but have also been found to have pathogenic roles in autoimmune and inflam-
matory diseases.\textsuperscript{31–35} Evidence has emerged that Th17 cells are active in the process of IRI and allograft rejection. IL-17A\textsuperscript{-/-} and IL-17R\textsuperscript{-/-} mice were found to be protected from kidney IRI\textsuperscript{36} and elevation of IL-17 mRNA, and protein has been identified in the early stages of rat and human kidney allograft rejection, suggesting a role for Th17 cells, particularly in the initiation of rejection.\textsuperscript{37–39} It has been shown, using mice deficient in the Th1-specific transcription factor T-bet, that Th17 cells can mediate cardiac allograft rejection\textsuperscript{40} and are responsible for costimulation blockade-resistant allograft re-
jection.\textsuperscript{41,42} In the current study, we found that WT allografts expressed significantly higher amounts of IL-6 and TGF-\beta than MyD88\textsuperscript{-/-} allografts, and IL-17 mRNA was significantly increased in WT allografts (Supplemental Figure 5). CD4\textsuperscript{+} and CD8\textsuperscript{+} cells isolated from WT allografts contained higher percent-
ges of IL-17\textsuperscript{+} producing cells than those cells from MyD88\textsuperscript{-/-} allografts. WT splenocytes primed and stimulated with donor-matched allogeneic cells produced significantly higher numbers of IL-17\textsuperscript{+} producing cells than similarly stimulated MyD88\textsuperscript{-/-} splenocytes. Finally, we confirmed that IL-17\textsuperscript{+} mice displayed prolonged kidney allograft survival, suggesting that Th17 cells contribute to rejection in our model.

Th17 cells share a complex relationship with Tregs and may modulate Treg-induced transplant tolerance. Naïve T cells in the presence of TGF-\beta may differentiate into either Th17 or induced Tregs, with Th17 development promoted and in-
duced Treg generation blocked by the concomitant presence of IL-6.\textsuperscript{43–45} IL-6, produced in response to TLR engagement by agonists, rendered activated T cells refractory to the suppres-
sive activity of Tregs,\textsuperscript{46} and nTregs, in the presence of IL-6, can themselves differentiate into Th17 cells.\textsuperscript{47} In transplantation, the activation of innate immunity through TLR signaling
Figure 4. Absence of MyD88 signaling resulted in reduced mRNA expression of proinflammatory and Th1 cytokines, chemokines, and fibrosis-related genes. IL-6, TNF-α, and IL-1β expression was attenuated in MyD88−/− allografts compared with WT allografts. IFN-γ mRNA expression was reduced in MyD88−/− allografts compared with WT allografts. IL-4 mRNA expression in WT and MyD88−/− allografts was higher than in isografts, with no significant difference between MyD88−/− and WT mice. Upregulated IL-17 expression was evident in WT allografts compared with isografts and reduced in MyD88−/− allografts (*P=0.08). CCL2 and CXCL9 expression were significantly reduced in MyD88−/− allografts compared with WT allografts. Expression of the fibrosis-related genes TGF-β, MMP2, and TIMP1 were reduced in MyD88−/− allografts versus WT (*P<0.05). Data are presented as mean ± SD (n=5 for isografts; n=8 for each allograft group).
triggered by IRI leads to an inflammatory milieu characterized by increased production of key cytokines, including IL-6. The resultant increase in IL-6 may lead to induction of Th17 cells but also inhibition of Tregs, promoting graft rejection over tolerance. Administration of the exogenous TLR9 ligand CpG at the time of transplant abrogated costimulation blockade-induced cardiac allograft acceptance in WT mice by the induction of IFN-γ and IL-17 production by intragraft CD4+ T cells and the prevention of induced Treg development. Consistent with this concept, our data show that IL-6 expression was reduced in MyD88−/− versus WT allografts. We also found a significantly higher number of CD4+ and CD8+ IL-17− producing T cells in rejecting WT allografts versus tolerant MyD88−/− allografts, whereas CD4+FoxP3+ Tregs were significantly increased in tolerant MyD88−/− spleen and allografts compared with rejecting WT counterparts. Our results suggest that absence of MyD88 impairs Th17 development, resulting in an increased ratio of Tregs over Th17. A recent study showed that absence of TLR/MyD88 signaling in DCs impairs Th17 cell development in response to phagocytosis of infected apoptotic cells, and instead, it supports Treg development. Indeed, our in vitro data also showed that absence of MyD88 impairs Th17 development and results in an increased ratio of Tregs/Th17effs in an alloreactive culture in vitro.

TLR activation may prevent experimental transplantation tolerance. Peritransplant infection triggers TLR signaling, which abrogates transplantation tolerance induced by costimulatory blockade. Administration of TLR agonists abrogated costimulation blockade-induced prolongation of skin allograft survival and prevented cardiac allograft acceptance, which was associated with impaired intragraft recruitment and function of CD4+FoxP3+ regulatory T cells. Conversely, impaired innate responses in MyD88-deficient recipients promoted costimulation blockade-induced allograft acceptance by the impairment of DC production of IL-6, reduced T cell activation, and enhanced T cell suppression mediated by CD4+CD25+ Tregs. We found a high frequency of CD4+FoxP3+ cells within spleens and kidney allografts of tolerant

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**Figure 5.** Donor antigen-specific immune tolerance in the absence of MyD88. WT and MyD88−/− kidney allograft acceptors received skin grafts from BALB/c (donor-matched allograft), C57BL/6 (isograft), and B10Br (H-2k; third-party allograft) mice at day 150 after kidney transplant. (A) Skin graft survival curves for donor-matched BALB/c skin challenges in WT and MyD88−/− acceptors. (B) WT acceptors rejected skin allografts from both donor-matched BALB/c and third-party B10Br mice. In contrast, MyD88−/− recipients accepted donor-matched skin allografts for over 100 days after skin transplant but rejected B10Br third-party allografts. Skin isografts on WT and MyD88−/− kidney allograft recipients remained intact. (C) MyD88−/− splenocytes from an MyD88−/− kidney allograft acceptor at day 100 post-transplant stimulated with donor-matched allogeneic spleen cells produced significantly less IFN-γ than similarly stimulated WT splenocytes from a WT allograft acceptor (**P<0.001). Data are mean ± SD (n=4 for each group).

**Figure 6.** Allograft tolerance in the absence of MyD88 signaling was dependent on a CD4+CD25+FoxP3+ regulatory mechanism. (A) Administration of anti-CD25 Ab to deplete CD4+CD25+ Tregs abrogated graft acceptance in MyD88−/− recipients (**P<0.01). (B) Administration of anti-CD25 Ab did not alter allograft rejection kinetics in WT recipients. (C) Transferable tolerance. CD4+CD25+ Tregs isolated from long-term C57BL/6, MyD88−/− acceptors were adoptively cotransferred with WT CD4+CD25+ T effectors into C57BL/6, RAG−/− recipients that were previously transplanted with a BALB/c skin graft; skin grafts remained intact, whereas transfer of WT Teffs either alone or with Teffs from long-term MyD88−/− acceptors caused graft loss (**P<0.05).
Figure 7. An increased ratio of T regulatory/Th17 effector cells in MyD88<sup>-/-</sup> allografts. (A) CD4<sup>+</sup>FoxP3<sup>+</sup> regulatory T cells were significantly increased in MyD88<sup>-/-</sup> spleen (n=6 for WT isograft, n=3 for MyD88<sup>-/-</sup> isograft, and n=10 for each allograft group) and MyD88<sup>-/-</sup> allografts (n=5) compared with WT allografts (n=4) by flow cytometry analysis (*P<0.05, **P<0.01). (B) IL-17 and IFN-γ production by T cells isolated from allografts at day 14 after kidney transplant. The percentages of CD4<sup>+</sup> and CD8<sup>+</sup> cells producing IL-17 obtained from WT allografts (n=5) were significantly higher than percentages from MyD88<sup>-/-</sup> allografts (n=4; **P<0.01). (C) The percentage of T cells producing IFN-γ was similar in WT and MyD88<sup>-/-</sup> allografts (n=8 each). Data are presented as mean ± SD.
Figure 8. Absence of MyD88 signaling-impaired alloreactive Th17 development results in an increased ratio of T regulatory/Th17 effector cells in mixed lymphocyte cultures. (A) A representative flow cytometry analysis for reciprocal expression of FoxP3+ and IL-17+ in CD4+ T cells. (B) CD4+ T cells obtained from naïve C57BL/6.WT or C57BL/6.MyD88−/− mice, stimulated with BALB/c.WT or BALB/c.MyD88−/− allogeneic splenocytes in the absence of exogenous TGF-β and IL-6, did not result in the induction of either Foxp3+ or IL-17+ T effectors. Addition of TGF-β to WT and MyD88−/− CD4+ T cells in culture generated a comparable number of Foxp3+ Tregs. (C) Addition of IL-6 in the presence of TGF-β to WT CD4+ T cells during differentiation increased the yield of Th17 cells, whereas MyD88−/− CD4+ T cells under the same conditions differentiated into significantly fewer numbers of IL-17-producing T cells compared with WT CD4+ T cells (**P<0.001). Combined data from four independent experiments are presented as mean ± SD (n=5–9 each).

MyD88−/− recipients, whereas depletion of CD4+CD25+ FoxP3+ T cells in tolerant MyD88−/− recipients broke tolerance. We were also able to transfer graft acceptance by CD4+CD25+ Tregs. Collectively, our data identify CD4+CD25+ FoxP3+ Tregs as critical mediators of tolerance development and maintenance in MyD88−/− recipients in this model.

One limitation of our data is the lack of specificity of MyD88 for the TLRs, because both IL-1 and -18 can also signal through MyD88. Studies using caspase-1–deficient recipients that cannot activate MyD88 through IL-1 and -18 have shown that TLR-MyD88 signaling is critical for the rejection of minor mismatched skin allografts and costimulation blockade-induced skin allograft acceptance.53 We previously showed that IL-18 does not impact on the development of kidney allograft rejection in our model.54 Thus, we believe that TLR-mediated activation of the MyD88 pathway is critical in our model, and the findings are unlikely to be attributable to either IL-1 or -18.

In conclusion, our study has showed that, in the absence of MyD88, mice developed donor-specific kidney allograft tolerance, which was associated with impairment of Th17 and Th1 effectors, resulting in an increased ratio of Tregs over Th17 cells. Our data imply that inhibiting innate immunity may be a potential clinically relevant strategy to facilitate transplantation tolerance.

CONCISE METHODS

Animals
Male WT:BALB/c (H2-Kd) donor, C57BL/6 (H2-Kb) recipient, and C57BL/6.RAG−/− mice were obtained from the Animal Resource Center, Perth, Australia. MyD88−/− on BALB/c or C57BL/6 backgrounds (designated as BALB/c. MyD88−/− and B6.MyD88−/− mice) was provided by the Animal Service of The Australian National University with permission from Professor S. Akira (Osaka University, Osaka, Japan). The mice were housed in a specific pathogen-free facility within the University of Sydney. Male mice aged 10–14 weeks were used in all experiments. Experiments were conducted using established guidelines for animal care and approved by the animal ethics committee of the University of Sydney.

Experimental Design
Five groups of mice received a kidney transplant: (1) WT allografts (C57BL/6/mice that received a kidney from a BALB/c donor); (2) MyD88−/− allografts (C57BL/6/MyD88−/− mice that received a BALB/c.MyD88−/− donor kidney); (3) isografts (C57BL/6 recipients of a C57BL/6 kidney); (4) donor MyD88−/− allografts (C57BL/6/WT mice that received a kidney from a BALB/c. MyD88−/−/donor); and (5) recipient MyD88−/−/allografts (C57BL/6. MyD88−/−/mice that received a kidney from a BALB/c WT donor).

Survival Experiment
To determine the importance of MyD88 signaling in allograft rejection, MyD88−/−/allografts (n=17), donor MyD88−/− allografts (n=7), and recipient MyD88−/−/allografts (n=5) were compared with WT allografts (n=31) and isografts (n=6). For those recipients surviving to day 150, a proportion of MyD88−/−/allografts (n=4) and WT allografts (n=4) received full-thickness skin grafts from three sources: BALB/c (kidney donor-matched skin allograft), C57BL/6 (syngeneic), and B10Br (third-party allograft). Grafts were observed regularly for signs of rejection to at least day 100.
**Time Course Study**
To examine intragraft events and mechanisms, groups of WT allografts and MyD88<sup>−/−</sup> allografts were killed early (day 14) and late (day 100) to compare differences in acute and chronic rejection, respectively.

**Mouse Kidney Transplantation**
Orthotopic kidney transplants were performed on the animals as previously described. Briefly, the left kidney of the donor animal was flushed with heparinized saline and removed together with ureter and vessels en mass, including a small (1–2 mm) bladder cuff attached to the distal ureter. The recipient animal underwent a left-sided nephrectomy, and the transplanted kidney was placed orthotopically on day 0. Urinary tract reconstruction was established by either inserting the ureter into the bladder (day 14 experiment only) or suturing the bladder patch to a cystostomy located on the bladder dome<sup>55</sup> (a bladder-to-bladder anastomosis) for survival study and dome<sup>55</sup>, respectively. The total tubulitis score for each animal was the sum of these scores. Mild and severe tubulitis was multiplied by two or three, respectively. A score for the degree of tubulitis was calculated for each animal as per a modified method described previously, where each normal tubule was assigned a value of one and the number of tubules affected with mild and severe tubulitis was multiplied by two or three, respectively. The total tubulitis score for each animal was the sum of these scores.

**Skin Transplantation**
Full-thickness tail skin grafts were placed on graft beds prepared on recipients at day 150 after kidney allograft transplantation. Grafts were covered with protective bandages for 7 days. Rejection was defined as graft necrosis of the entire graft area. Recipients that accepted their grafts and MyD88<sup>−/−</sup> recipients generated a diminished donor-specific Th17 immune response as assessed by ELISPOT. (A) MyD88<sup>−/−</sup> splenocytes primed and stimulated with allogeneic cells produced more donor-reactive IFN-γ-producing cells than similarly stimulated WT controls (**P**<0.05). (B) IL-4 production was not different in allogstimulated WT compared with MyD88<sup>−/−</sup> spleen cells. (C) MyD88<sup>−/−</sup> splenocytes primed and stimulated with allogeneic cells produced significantly less IL-17 than similarly stimulated WT splenocytes (**P**<0.05). Representative data from two independent experiments are presented (mean ± SD, n=4 or n=8 each).

**Assessment of Kidney Function**
Serum creatinine was measured using the modified Jaffe rate reaction by the Biochemistry Department of The Royal Prince Alfred Hospital, Sydney, Australia.

**Histology**
All histologic analysis was performed in a blinded manner. Periodic acid-Schiff (PAS) staining was performed on 3 μm paraffin kidney sections to assess tubulitis, glomerulosclerosis, and interstitial fibrosis. Glomerulosclerosis was quantitated by the presence of PAS-positive staining material involving >30% of each glomerulus as described previously.<sup>26</sup> All glomeruli per section were scored to determine the percentage of glomeruli displaying glomerulosclerosis.

Interstitial fibrosis and tubular atrophy were graded using the Banff 97 scoring criteria on a scale of one to three: 1, mild interstitial fibrosis and tubular atrophy (<25% of cortical area); 2, moderate interstitial fibrosis and tubular atrophy (26%–50% of cortical area); 3, severe interstitial fibrosis and tubular atrophy/loss (>50% of cortical area). If changes were minimal but not absent, the score of 0.5 was applied. Using an ocular grid, the score of each sample was counted at embedding, frozen in OCT compound (Sakura Finetek Inc., Torrance, CA), or snap frozen in liquid nitrogen for subsequent mRNA extraction.
least in 15–25 consecutive fields across a full section (×200 magnification) and averaged for each graft.

Picro-Sirius red (PSR) staining was performed on 5-μm paraffin sections. Interstitial collagen was assessed by point counting using an ocular grid as described in the work by McWhinnie et al. in at least 12 consecutive fields (×200 magnification). Only interstitial collagen was counted, and vessels and gromeruli were excluded. The result was expressed as the percentage of positive staining area per field.

**Immunohistochemistry Staining**

Acetone-fixed frozen sections (7 μm) were exposed to 0.06% H2O2 in PBS, blocked with a biotin blocker system (DAKO Corporation), and then blocked in 10% normal horse serum. Primary antibody, rat anti-mouse CD68 antibody (ABD Serotec), CD4 (BD Biosciences, San Diego, CA), CD8 (BD Biosciences), and hamster anti-mouse CD11c (BD Biosciences) were applied to the sections for 60 minutes. Formalin-fixed paraffin-embedded sections of 5 μm thickness were deparaffinized and boiled for 15 minutes in 10 mM sodium citrate buffer (pH 6.0) for α-SMA (Abcam, MA), HMGB1 (Abcam, MA), and hyaluronan detection. Sections were blocked in 20% normal goat serum and incubated with rabbit anti-α-SMA or anti-HMGB1 for 1 hour. Endogenous peroxidase activity was then quenched in 3% H2O2 in methanol. Concentration-matched Ig was used as an isotype negative control. Sections were incubated with the biotinylated secondary antibody: anti-rat IgG, anti-hamster IgG, or anti-rabbit IgG (BD Biosciences). Instead of a primary antibody, a biotinylated hyaluronan-binding protein (Seikagaku Corp. Japan) at 5 μg/ml was used for hyaluronan staining. Vector stain ABC kit (Vector Laboratories Inc.) was applied to the tissue and followed by 3,3′-diaminobenzidine substrate–chromogen solution (DAKO, Carpinteria, CA). The slides were counterstained with Harris' hematoxylin.

**Quantification of Immunohistochemistry**

Analysis of the cellular infiltrates for CD4 and CD8 was performed in a blinded manner by assessing 20 consecutive high-power fields (HPFs; ×400 magnification) of the cortex in each section. Using an ocular grid, the number of cells staining positively for each antibody was counted and expressed as cells per HPF. Analysis of CD68⁺ and CD11c⁻ infiltrates was performed using a digital image analysis program (Automated Cellular Imaging System ACIS III; Dako, Aliso Viejo, CA). An area of cortex was analyzed for interstitial cellular-positive staining versus counter-stained area. The results were expressed as percentage of positive staining. Analysis of α-SMA staining was assessed by point counting using an ocular grid as described above. Staining around the vessels was not included. The result was expressed as the percentage of positive staining area per HPF.

**Immunofluorescence**

For C4d immunofluorescent staining, frozen sections were blocked with 1% BSA in PBS for 20 minutes and incubated with rat anti-mouse antibodies to C4d (Abcam plc, Cambridge, MA) for 60 minutes followed by anti-rat IgG conjugated with AlexaFluor 488 (Molecular Probes, Eugene, OR). Staining for C4d was considered positive when the peritubular capillaries were diffusely (all HPFs) and brightly stained. Scoring of C4d staining was based on the percentage of stained tissue on immunofluorescence that had a linear, circumferential staining pattern in PTCs following the Banff 07 scoring criteria on a scale of zero to three: 0, negative (0%); 1, minimal C4d stain/ detection (1<10%); 2, focal C4d stain/positive (10%–50%); 3, diffuse C4d stain/positive (>50%).

**RNA Extraction and cDNA Synthesis**

Total RNA was extracted from kidney tissue and cells using TRIzol (Invitrogen). cDNA was synthesized using oligo d (T)16 (Applied Biosystems, Foster City, CA) and the SuperScript III Reverse transcription kit (Invitrogen).

**Real-Time PCR**

Specific Taqman primers and probes for TLR2, TLR4, HMGB1, biglycan, HAS1, HAS2, hsp60, hsp70, IL-6, TNF-α, IL-1β, IL-4, IFN-γ, CCL2, CXCL10, inducible nitric oxide synthase, and glyceraldehyde-3-phosphate dehydrogenase were previously described. Specific Taqman primers and probes for Granzyme B (Mm00442834_m1), Perforin1 (Mm00812512_m1), TGF-β1 (Mm01178820_m1), IL-17 (Mm00439619_m1), CXCL9 (Mm01345157_m1), MMP2 (Mm00439506_m1), and TIMP1 (Mm01341361_m1) were obtained from Applied Biosystems (Foster City, CA). cDNA was amplified in iX Universal Master Mix (Applied Biosystems) with gene-specific primers and probes in the Rotor-Gene 6000 system (Corbett Life Science). Data were analyzed using Rotor-Gene 6000 Analysis Software (Corbett Life Science). All results are expressed as ratio to glyceraldehyde-3-phosphate dehydrogenase.

**Cell Isolation from Allografts**

Cell suspensions from the kidney grafts were isolated as previously described. Kidney cortex was minced and underwent digestion with collagenase and deoxyribonuclease (Sigma-Aldrich); then, they passed through a 250-μm sieve. Cells were collected by centrifugation, resuspended in 40% Percoll (Sigma-Aldrich) in RPMI-1640 medium, and overlayed on a 66.7% Percoll solution. Gradient separation was carried out at 2100 rpm for 20 minutes at room temperature. Spleen and blood cells were also isolated from mice using standard methods.

**Flow Cytometric Analysis**

Donor-specific alloantibody titers were measured by flow cytometry. Dilutions (1/10 and 1/20) of mouse serum from transplanted mice were incubated with BALB/c (donor) splenocytes for 1 hour at 4°C. After washing three times, the donor cells were incubated with either rat FITC-conjugated anti-mouse IgG1 or IgG2a (BD Biosciences). The median fluorescence intensity of the stained samples was determined by flow cytometry. All analyses were performed on a FACS CALIBUR flow cytometer, and data were analyzed with Cellequest Pro software (BD Biosciences). Results were read as median fluorescent intensity.

FITC-conjugated anti-mouse CD4 (clone RM4–5), phycoerythrin (PE)-conjugated anti-mouse IFN-γ (clone XMG1.2), PE-Cy7–conjugated anti-mouse CD8 (clone 53–6.7), APC-Cy7–conjugated anti-mouse CD45 (clone 30–F11), and isotype controls were purchased from BD Biosciences. APC-conjugated anti-mouse IL-17A (clone TC11–18H10) was purchased from Biolegend (San Diego, CA).
and PE–conjugated anti-mouse FoxP3 (clone FJK-16a) was purchased from eBioscience (San Diego, CA). IFN-γ and IL-17A intracytoplasmic stainings were performed after cell incubation with 10 µg/ml PMA and 500 ng/ml ionomycin in the presence of GolgiPlug (BD Biosciences) for 4 hours; then, the cells were stained for surface markers for 40 minutes, washed, fixed with CytoFix/CytoPerm (BD Biosciences), permeabilized with Perm/Wash buffer (BD Biosciences), and labeled with anticytokine antibodies. FoxP3 staining was performed after cell surface marker labeling using eBioscience fixation/permeabilization and permeabilization buffers according to the manufacturer’s instructions. Cytometry analysis was performed on a Canto cytometer using FACS Diva software (BD Biosciences), and data were analyzed with FLOWJO software (Tree Star, Inc).

CD25+ Cell Depletion

To deplete CD4+CD25+ cells in vivo, mice received rat anti-mouse CD25 mAb (clone PC-61, rat IgG1; BioXcell, NH) or isotype rat IgG (0.5 mg/mouse) intraperitoneally on days $-2, 0$, and $2$ and 2 weeks post-transplantation ($n=9$). CD25+ cell depletion was confirmed between days 14 and 21 after kidney transplantation by staining blood white cells with anti-CD25 (clone 7D4 or 3C7; BD Biosciences, Phar-mingen, CA).

CD4+ T Cell Enrichment and Cell Sorting

To purify CD4+CD25+ (Tregs) and CD4+CD25− (Teffs), spleen cells were enriched for CD4+ T cells using a mouse CD4+ T cell enrichment kit (EasySep; StemCell Technologies) according to the manufacturer’s instructions. The enriched CD4+ T cells were stained with FITC-conjugated anti-mouse CD4 and APC-conjugated anti-mouse CD25 (clone PC-61; BD Biosciences), and then, they were sorted for CD4+CD25+ or CD4+CD25− cells (>95% purity) by FACS.

Adoptive Transfer

C57BL/6.RAG−/− mice received full-thickness BALB/c skin grafts 2 weeks before receiving adoptive transfers through tail vein injection of either $2 \times 10^5$ CD4+CD25+ (Tregs) or CD4+CD25− (Teffs) harvested from C57BL/6.MyD88−/− recipients that had accepted their BALB/c.MyD88−/− kidney allografts for >200 days; $1 \times 10^5$ CD4+CD25− T effector cells harvested from naive WT C57BL/6 mice were coinfused.

In Vitro T Cell Differentiation

CD4+ T cells from the spleens of WT and MyD88−/− mice on C57BL/6 backgrounds were purified using anti-CD4 beads (Miltenyi) according to the manufacturer’s instructions. Purified CD4+ T cells were stimulated with allogeneic, irradiated splenocytes from BALB/c or BALB/c.MyD88−/− mice in a 48-well plate under iTreg differentiation conditions (TGF-β; 10 ng/ml) in the presence or absence of IL-6 (20 ng/ml) in complete RPMI-10 medium for 5 days. Cells were supplemented with recombinant IL-2 (50 U/ml) at 24 and 72 hours.

ELISPOT Analysis of Donor-Specific IFNγ, IL4, and IL17 Production

ELISpotPLUS kits for IFNγ, IL4, and IL17 (Mabtech AB, Stockholm, Sweden) were used according to the manufacturer’s instructions as previously described.54 Responder splenocytes were isolated from mice (C57BL/6 WT and C57BL/6 MyD88−/−) primed 14 days prior with $1.5 \times 10^5$ BALB/c splenocytes administered by tail vein injection or from WT and MyD88−/− kidney allograft acceptors at 100 days after kidney transplant. Stimulator splenocytes from naïve BALB/c, C57BL/6, or B10.BR mice were isolated and irradiated. Responder and irradiated stimulator splenocytes were each added at $3 \times 10^5$ cells/well to a final volume 100 µl and incubated for 24 hours with 5% CO2 at 37°C. The plates were washed, and the appropriate biotinylated detection antibody (Mabtech) (diluted to 1 µg/ml in PBS containing 1.5% FCS) was added and incubated overnight at 4°C. After washings, streptavidin-alkaline phosphatase (1:1000) was added for 60 minutes at room temperature. The wells were washed and then developed with filtered substrate solution BCIP/NBT-plus (Mabtech) until distinct spots emerged. Concanavalin A was added to some wells with responder cells only as a positive, technical control; negative control wells with stimulator only and medium only were also performed on each plate. All combinations were performed at least in triplicate. Spots were counted using an automated AID ELISPOT reader (Autoimmum Diagnostika, Strasberg, Germany).

Statistical Analyses

All data are expressed as mean ± SD. Comparisons between graft survival times were calculated using Kaplan–Meier survival curves with the log-rank test. Statistical differences between the two groups were analyzed by unpaired two-tailed t tests, and multiple groups were compared using one- or two-way ANOVA with posthoc Bonferroni correction (Graph Pad Prism 4.0 software). $P$ values <0.05 were considered significant.

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DISCLOSURES

None.

REFERENCES


**Supplemental figure 1** Absence of MyD88 signaling on either donor or recipients was protective against acute rejection as evidenced by better kidney function (A) and less tubulitis (B) compared to WT allografts (*P<0.05; **P<0.01; ***P<0.001).
Supplemental figure 2. Absence of MyD88 signaling led to impaired donor specific alloantibody production. MyD88−/− allograft recipients (n=7) had less IgG2a allo-specific antibody in serum at day 14 post transplant versus WT recipients (n= 8, *p < 0.05). By 100 days, WT allograft recipients (n=8) maintained higher concentrations of both IgG2a and IgG1 donor-antigen specific antibodies compared MyD88−/− recipients (n=7) and isografts (n=5, *p<0.05; ** p<0.01). Data are mean ± SD.
Supplemental figure 3  IL-17 and IFN-γ production by splenocytes isolated from allograft recipients at day 14 post kidney transplant by flow cytometry analysis. There were no significant differences in the percentages of CD4⁺ and CD8⁺ splenocytes obtained from WT and MyD88⁻/⁻ allograft recipients producing either IL-17 or IFN-γ (n=7-10 per group).
Supplemental figure 4. Upregulation of TLR2&4 and their endogenous ligands in WT allografts. A. mRNA expression of TLR2 and TLR4 were significantly upregulated in WT allografts (*p<0.05 to **p<0.001). B. HMGB1, biglycan and HAS1&2 mRNA levels were significantly increased in allografts from day 3 (*p<0.05 to **p<0.001). C. Elevated expression of HMGB1 and hyaluronan in WT allografts at day 14 post transplant were confirmed by immunostaining.
Supplemental figure 5. IL-17 mRNA was significantly increased in WT allografts from days 5 to 14 post-transplantation.