

- myocardial infarction reveals a role of miR-29 in cardiac fibrosis. *Proc Natl Acad Sci USA* 105: 13027–13032, 2008
15. Roderburg C, Urban GW, Bettermann K, Vucur M, Zimmermann H, Schmidt S, Janssen J, Koppe C, Knolle P, Castoldi M, Tacke F, Trautwein C, Luedde T: Micro-RNA profiling reveals a role for miR-29 in human and murine liver fibrosis. *Hepatology* 53: 209–218, 2011
 16. Long J, Wang Y, Wang W, Chang BH, Danesh FR: MicroRNA-29c is a signature microRNA under high glucose conditions that targets Sprouty homolog 1, and its in vivo knockdown prevents progression of diabetic nephropathy. *J Biol Chem* 286: 11837–11848, 2011
 17. Reddy MA, Natarajan R: Epigenetics in diabetic kidney disease. *J Am Soc Nephrol* 22: 2182–2185, 2011

See related article, "Suppression of microRNA-29 Expression by TGF- β 1 Promotes Collagen Expression and Renal Fibrosis," on pages 252–265.

Sphingosine Lipids in the Resolution of Renal Ischemia and Reperfusion Injury

Almut Grenz

Mucosal Inflammation Program, Department of Anesthesiology, University of Colorado, Anschutz Medical Campus, Aurora, Colorado

J Am Soc Nephrol 23: 187–189, 2012.
doi: 10.1681/ASN.2011121234

Despite all research endeavors in the field of AKI, its severity and incidence are still increasing without substantial improvements in its prevention or therapy.¹ Thus, AKI is still associated with a dramatic increase in morbidity and mortality in patients; a recent study in hospitalized patients indicated that a rise of serum creatinine of only 0.3 mg/dl is associated with a 70% increase in the risk of death.² The increase in morbidity and mortality in conjunction with the lack of available therapies and the incredible costs associated with renal failure make it an area of intense investigations.

One of the leading causes of AKI is renal ischemia with attenuated blood supply to the kidneys associated with sepsis.^{1,3–7} Ischemic tissue damage has multifaceted effects on renal tissues, including renal inflammation, direct tubular damage, and alterations of vascular responses. AKI caused by ischemia can occur in different clinical settings such as surgical procedures, where cross-clamping of the aorta and renal vessels is associated with a renal failure rate of up to 30%.⁸ Similarly, acute renal failure after cardiac surgery occurs in up to 10% of patients under normal circumstances and is associated

with dramatic increases in mortality.⁹ Therefore, new potential therapeutic targets are of urgent need to prevent renal injury caused by ischemia.^{10–15} Luckily, mouse models of AKI are very well established, and studies in gene-targeted or conditional mice offer great potential for investigating interesting targets protective of AKI.

Lee and colleagues^{16–21} have intensively investigated the role of adenosine receptors, which are G-protein-coupled receptors, in the pathophysiology of AKI and have opened novel and promising potential therapeutic pathways in treating AKI caused by ischemia. In this issue of *JASN*, Park *et al.*²² investigated the impact of sphingosine 1-phosphate (S1P) and its G-protein-coupled receptor, so called S1P receptor 2 (S1P₂R), in renal ischemia reperfusion (IR) injury.

Sphingosine and its receptors belong to the sphingolipid family. One characteristic of lipids compared with other messenger molecules is they can freely diffuse across membranes. Thus, they cannot be stored in vesicles but are biosynthesized on demand. Sphingolipids are a class of lipids containing a backbone of sphingosine bases and aliphatic amino alcohols that includes sphingosine. Sphingosine is generated from *N*-deacylation of ceramide by ceramidase. It can be phosphorylated by sphingosine kinases (SK1 and SK2) to S1P. Once phosphorylated to S1P, it can activate all of the five known S1P receptors.

Five S1P receptors, discovered in the early 1990s, have been cloned thus far (S1P₁R, S1P₂R, S1P₃R, S1P₄R, and S1P₅R).²³ S1P receptors and their mediators have been recently shown to play an important role as potent bioactive messengers in cell differentiation, proliferation, apoptosis, migration, and angiogenesis.²⁴ Moreover, modulators of S1P receptor attenuated vascular leak during acute lung injury, attenuated ischemia reperfusion injury in the heart and the kidneys,^{25,26} and improved graft survival²⁷ in animal models. Furthermore, synthetic S1P receptors show therapeutic efficacy in clinical trials in multiple sclerosis.²⁸ The S1P₁ receptor is the most extensively studied receptor in immune-modulatory processes, whereas the role of the S1P₂ receptor in IR injury is largely unknown. Genetic deletion of the S1P₁ receptor in mice causes embryonic lethality due to incomplete vascular maturation,²⁹ whereas genetic deletion of S1P₂ leads to deafness.³⁰ In the kidney, S1P receptors are expressed on proximal tubules, endothelial cells, and immune cells.^{31,32}

Thus, the present study of Park *et al.*²² is of interest, based on recent findings, and develops comprehensive insights into the complex role of the sphingosine receptor, S1P₂R, and its mediator, S1P, and sphingosine kinases in renal IR injury. Park *et al.* observed that pharmacological inhibition of S1P₂R provides dose-dependent protection against IR injury. On a genetic level, they could show that gene-targeted mice for the S1P₂R or mice treated with small interfering RNA targeting S1P₂R were protective of IR injury. To confirm that S1P₂R is the most important of the five known S1PRs, the authors determined transcript and protein levels and showed that receptor 2 was the one most upregulated after renal ischemia compared with the other four known receptors.

Published online ahead of print. Publication date available at www.jasn.org.

Correspondence: Dr. Almut Grenz, Mucosal Inflammation Program, Department of Anesthesiology, University of Colorado, 12700 E. 19th Avenue, Anschutz Medical Campus, Room 7121, RC-2 (P15), B112, Aurora, CO 80045. Email: almut.grenz@ucdenver.edu

Copyright © 2012 by the American Society of Nephrology

In a next step, the authors observed that pharmacologic inhibition or genetic deletion of S1P₂R reduces renal tubular necrosis after IR injury. Along this line, proinflammatory cytokines (TNF α and intercellular adhesion molecule 1) were reduced in mice with pharmacological inhibition of S1P₂R. Because S1P₂R is known to activate the Rho GTPase and the Rho-associated, coiled-coil containing protein kinase 1, also known as ROCK1 pathways, the authors suggest a possible role by blocking S1P₂R. They performed an elegant experiment by pretreating the mice with selective Rho and ROCK inhibitors before activating these pathways with a S1P₂R agonist. Both inhibitors attenuated the renal dysfunction induced by an S1P₂R agonist and protected against renal IR injury.

Since it has been shown that a downregulation or deletion of this receptor in mice goes along with an increase of SK1 and SK2, and since an upregulation of SK1 and SK2 is protective against cardiac and renal IR injury,^{33,34} the authors question whether the protective effect they see by inhibition or deletion of the S1P₂R receptor might be caused by an upregulation of SK1 or SK2. Furthermore, the phosphorylation of SKs resulted in the formation of S1P, which could be the mediator of renal injury. Interestingly, S1P₂R deletion is accompanied by a marked upregulation in SK1, and a selective inhibition of SK1 in S1P₂R-deleted mice abolished its protection against IR injury. Furthermore, the protective effect of S1P₂R inhibition was abolished in mice with additional pharmacological blockade of the S1P₂R, suggesting that S1P synthesis is necessary for the protective effect. Thus far, S1P₂R deletion seems to mediate protection by an upregulation of SK1 and an increase of S1P mediated by S1P₁R.

Hypoxia-inducible factors (HIFs) are an important mediator under hypoxic conditions and during inflammatory processes. HIF-1 α is stabilized and induces the expression of genes that are known to be protective in inflammation and organ injury due to ischemia/hypoxia.³⁵ Therefore, the authors hypothesize that the S1P pathways might be stimulated by HIF-1 α . Furthermore, it is known that SK is a hypoxia-regulated gene.³⁶ Interestingly, pretreatment of cells with an HIF-1 α inhibitor before exposure to hypoxia significantly attenuates levels of mRNA encoding SK1. Furthermore, inhibition of S1P₂R increases nuclear HIF-1 α . Moreover, inhibition of HIF-1 α abolishes the protective effect of S1P₂R blockade in IR injury in mice. These data suggest that HIF-1 α plays a critical role in renal protection by S1P₂R inhibition.

In a last experiment, the authors wanted to clarify which renal cell type is involved in that protective mechanism. They could show that SK1 was only induced in isolated tubules and not in isolated endothelial cells from mouse kidneys after IR injury. Thus, their data suggest S1P₂R directly modulates the synthesis of HIF-1 α , which increases SK1 synthesis and thus mediates renal protection in tubular cells.

Lipids play a pivotal role in the resolution process during organ inflammation.^{37–39} The resolution phase is a pretty recently defined period during organ inflammation that has to be separated from active processes of inflammation, particularly the reduction of neutrophil infiltration. The resolution process is a more passive process where various molecules promote the

clearance of inflammatory cells. Members of the lipid family, acting as pro-resolution molecules, can be determinant in attenuating inflammation. The role of the sphingosine lipids, as described by Park *et al.*²² during renal IR injury, fits this resolution scheme of protection from renal IR injury. However, more studies are warranted to clarify the precise mechanism of these sphingosine lipids during renal IR injury.

DISCLOSURES

None.

REFERENCES

- Abuelo JG: Normotensive ischemic acute renal failure. *N Engl J Med* 357: 797–805, 2007
- Chertow GM, Burdick E, Honour M, Bonventre JV, Bates DW: Acute kidney injury, mortality, length of stay, and costs in hospitalized patients. *J Am Soc Nephrol* 16: 3365–3370, 2005
- Day YJ, Huang L, McDuffie MJ, Rosin DL, Ye H, Chen JF, Schwarzschild MA, Fink JS, Linden J, Okusa MD: Renal protection from ischemia mediated by A2A adenosine receptors on bone marrow-derived cells. *J Clin Invest* 112: 883–891, 2003
- Day YJ, Huang L, Ye H, Li L, Linden J, Okusa MD: Renal ischemia-reperfusion injury and adenosine 2A receptor-mediated tissue protection: the role of CD4+ T cells and IFN-gamma. *J Immunol* 176: 3108–3114, 2006
- Molitoris BA, Levin A, Warnock DG, Joannidis M, Mehta RL, Kellum JA, Ronco C, Shah S: Acute Kidney Injury Network: Improving outcomes from acute kidney injury. *J Am Soc Nephrol* 18: 1992–1994, 2007
- Lameire N: The pathophysiology of acute renal failure. *Crit Care Clin* 21: 197–210, 2005
- Schrier RW, Wang W: Acute renal failure and sepsis. *N Engl J Med* 351: 159–169, 2004
- Gelman S: The pathophysiology of aortic cross-clamping and unclamping. *Anesthesiology* 82: 1026–1060, 1995
- Mehta RL: Acute renal failure and cardiac surgery: marching in place or moving ahead? *J Am Soc Nephrol* 16: 12–14, 2005
- Roos A, Rastaldi MP, Calvaresi N, Oortwijn BD, Schlagwein N, van Gijlswijk-Janssen DJ, Stahl GL, Matsushita M, Fujita T, van Kooten C, Daha MR: Glomerular activation of the lectin pathway of complement in IgA nephropathy is associated with more severe renal disease. *J Am Soc Nephrol* 17: 1724–1734, 2006
- Zhou W, Farrar CA, Abe K, Pratt JR, Marsh JE, Wang Y, Stahl GL, Sacks SH: Predominant role for C5b-9 in renal ischemia/reperfusion injury. *J Clin Invest* 105: 1363–1371, 2000
- Grenz A, Dalton JH, Bauerle JD, Badulak A, Ridyard D, Gandjeva A, Aherne CM, Brodsky KS, Kim JH, Tuder RM, Eltzschig HK: Partial netrin-1 deficiency aggravates acute kidney injury. *PLoS ONE* 6: e14812, 2011
- Grenz A, Osswald H, Eckle T, Yang D, Zhang H, Tran ZV, Klingel K, Ravid K, Eltzschig HK: The reno-vascular A2B adenosine receptor protects the kidney from ischemia. *PLoS Med* 5: e137, 2008
- Grenz A, Zhang H, Eckle T, Mittelbronn M, Wehrmann M, Köhle C, Kloor D, Thompson LF, Osswald H, Eltzschig HK: Protective role of ecto-5'-nucleotidase (CD73) in renal ischemia. *J Am Soc Nephrol* 18: 833–845, 2007
- Grenz A, Zhang H, Hermes M, Eckle T, Klingel K, Huang DY, Müller CE, Robson SC, Osswald H, Eltzschig HK: Contribution of E-NTPDase1 (CD39) to renal protection from ischemia-reperfusion injury. *FASEB J* 21: 2863–2873, 2007
- Joo JD, Kim M, Horst P, Kim J, D'Agati VD, Emala CW Sr, Lee HT: Acute and delayed renal protection against renal ischemia and reperfusion injury with A1 adenosine receptors. *Am J Physiol Renal Physiol* 293: F1847–F1857, 2007

17. Lee HT, Emala CW: Protective effects of renal ischemic preconditioning and adenosine pretreatment: role of A(1) and A(3) receptors. *Am J Physiol Renal Physiol* 278: F380–F387, 2000
18. Lee HT, Emala CW: Preconditioning and adenosine protect human proximal tubule cells in an in vitro model of ischemic injury. *J Am Soc Nephrol* 13: 2753–2761, 2002
19. Lee HT, Gallos G, Nasr SH, Emala CW: A1 adenosine receptor activation inhibits inflammation, necrosis, and apoptosis after renal ischemia-reperfusion injury in mice. *J Am Soc Nephrol* 15: 102–111, 2004
20. Lee HT, Kim M, Jan M, Penn RB, Emala CW: Renal tubule necrosis and apoptosis modulation by A1 adenosine receptor expression. *Kidney Int* 71: 1249–1261, 2007
21. Lee HT, Xu H, Nasr SH, Schnermann J, Emala CW: A1 adenosine receptor knockout mice exhibit increased renal injury following ischemia and reperfusion. *Am J Physiol Renal Physiol* 286: F298–F306, 2004
22. Won Park S, Kim M, Brown KM, D'Agati VD, Lee HT: Inhibition of sphingosine 1-phosphate receptor 2 protects against renal ischemia-reperfusion injury. *J Am Soc Nephrol* 23: 266–280, 2012
23. Mandala S, Hajdu R, Bergstrom J, Quackenbush E, Xie J, Milligan J, Thornton R, Shei GJ, Card D, Keohane C, Rosenbach M, Hale J, Lynch CL, Rupprecht K, Parsons W, Rosen H: Alteration of lymphocyte trafficking by sphingosine-1-phosphate receptor agonists. *Science* 296: 346–349, 2002
24. Spiegel S, Milstien S: The outs and the ins of sphingosine-1-phosphate in immunity. *Nat Rev Immunol* 11: 403–415, 2011
25. Means CK, Xiao CY, Li Z, Zhang T, Omens JH, Ishii I, Chun J, Brown JH: Sphingosine 1-phosphate S1P2 and S1P3 receptor-mediated Akt activation protects against in vivo myocardial ischemia-reperfusion injury. *Am J Physiol Heart Circ Physiol* 292: H2944–H2951, 2007
26. Lien YH, Yong KC, Cho C, Igarashi S, Lai LW: S1P(1)-selective agonist, SEW2871, ameliorates ischemic acute renal failure. *Kidney Int* 69: 1601–1608, 2006
27. Shimizu H, Takahashi M, Kaneko T, Murakami T, Hakamata Y, Kudou S, Kishi T, Fukuchi K, Iwanami S, Kuriyama K, Yasue T, Enosawa S, Matsumoto K, Takeyoshi I, Morishita Y, Kobayashi E: KRP-203, a novel synthetic immunosuppressant, prolongs graft survival and attenuates chronic rejection in rat skin and heart allografts. *Circulation* 111: 222–229, 2005
28. Kappos L, Antel J, Comi G, Montalban X, O'Connor P, Polman CH, Haas T, Korn AA, Karlsson G, Radue EW; FTY720 D2201 Study Group: Oral fingolimod (FTY720) for relapsing multiple sclerosis. *N Engl J Med* 355: 1124–1140, 2006
29. Liu Y, Wada R, Yamashita T, Mi Y, Deng CX, Hobson JP, Rosenfeldt HM, Nava VE, Chae SS, Lee MJ, Liu CH, Hla T, Spiegel S, Proia RL: Edg-1, the G protein-coupled receptor for sphingosine-1-phosphate, is essential for vascular maturation. *J Clin Invest* 106: 951–961, 2000
30. Herr DR, Grillet N, Schwander M, Rivera R, Müller U, Chun J: Sphingosine 1-phosphate (S1P) signaling is required for maintenance of hair cells mainly via activation of S1P2. *J Neurosci* 27: 1474–1478, 2007
31. Jo SK, Bajwa A, Awad AS, Lynch KR, Okusa MD: Sphingosine-1-phosphate receptors: Biology and therapeutic potential in kidney disease. *Kidney Int* 73: 1220–1230, 2008
32. Awad AS, Ye H, Huang L, Li L, Foss FW Jr, Macdonald TL, Lynch KR, Okusa MD: Selective sphingosine 1-phosphate 1 receptor activation reduces ischemia-reperfusion injury in mouse kidney. *Am J Physiol Renal Physiol* 290: F1516–F1524, 2006
33. Kim M, Park SW, Kim M, D'Agati VD, Lee HT: Isoflurane activates intestinal sphingosine kinase to protect against bilateral nephrectomy-induced liver and intestine dysfunction. *Am J Physiol Renal Physiol* 300: F167–F176, 2011
34. Vessey DA, Kelley M, Li L, Huang Y, Zhou HZ, Zhu BQ, Karliner JS: Role of sphingosine kinase activity in protection of heart against ischemia reperfusion injury. *Med Sci Monit* 12: BR318–BR324, 2006
35. Eltzschig HK, Carmeliet P: Hypoxia and inflammation. *N Engl J Med* 364: 656–665, 2011
36. Schwalm S, Döll F, Römer I, Bubnova S, Pfeilschifter J, Huwiler A: Sphingosine kinase-1 is a hypoxia-regulated gene that stimulates migration of human endothelial cells. *Biochem Biophys Res Commun* 368: 1020–1025, 2008
37. Serhan CN: Resolution phase of inflammation: novel endogenous anti-inflammatory and proresolving lipid mediators and pathways. *Annu Rev Immunol* 25: 101–137, 2007
38. Serhan CN, Savill J: Resolution of inflammation: The beginning programs the end. *Nat Immunol* 6: 1191–1197, 2005
39. Serhan CN, Chiang N, Van Dyke TE: Resolving inflammation: Dual anti-inflammatory and pro-resolution lipid mediators. *Nat Rev Immunol* 8: 349–361, 2008

See related article, "Inhibition of Sphingosine 1-Phosphate Receptor 2 Protects against Renal Ischemia-Reperfusion Injury," on pages 266–280.

The Crossroad of RAAS Modulation, Inflammation, and Oxidative Stress in Dialysis Patients: Light at the End of the Tunnel?

Joshua J. Zaritsky*[†] and Kamyar Kalantar-Zadeh*^{‡§}

*Harold Simmons Center for Chronic Disease Research and Epidemiology, Los Angeles Biomedical Research Institute at Harbor-UCLA, Torrance, California; and [†]Division of Pediatric Nephrology, [‡]UCLA David Geffen School of Medicine, and [§]Department of Epidemiology, UCLA School of Public Health, Los Angeles, California

J Am Soc Nephrol 23: 189–191, 2012.
doi: 10.1681/ASN.2011121208

After four decades of fully reimbursed chronic dialysis therapy in the United States under the justification of lifesaving treatment, the survival rate of dialysis patients remains worse than many fatal cancers.¹ Most recorded causes of mortality on death certificates are cardiovascular or infectious; however, the true etiology of poor survival is unknown. Some slight improvement in the survival of dialysis patients reported over the past few years may be attributed to increased life expectancy in the background healthy population, although in the interest of fairness, the increased use of cardio-protective and other pharmacologic agents should also be noted as potential contributors.¹ Nevertheless, we have not succeeded in saving lives more significantly than before.

In line with ongoing attempts to uncover the underlying etiology of death in dialysis patients, sporadic attention has been paid to the evil axis of malnutrition, inflammation, and oxidative stress. There are compelling reasons why we cannot afford to ignore this axis: up to two-thirds of long-term dialysis

Published online ahead of print. Publication date available at www.jasn.org.

Correspondence: Dr. Kamyar Kalantar-Zadeh, Harold Simmons Center for Chronic Disease Research and Epidemiology, Los Angeles Biomedical Research Institute at Harbor-UCLA, Torrance, CA 90501. Email: kamkal@ucla.edu

Copyright © 2012 by the American Society of Nephrology