Activation of Parenchymal CD47 Promotes Renal Ischemia-Reperfusion Injury

Natasha M. Rogers,*† Angus W. Thomson† and Jeffrey S. Isenberg*

*Division of Pulmonary Allergy and Critical Care Medicine, Vascular Medicine Institute, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania; and †Starzl Transplantation Institute, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania

ABSTRACT

Ischemia-reperfusion injury (IRI) contributes to decreased allograft function and allograft rejection in transplanted kidneys. Thrombospondin-1 is a stress protein typically secreted in response to hypoxia and the ligand activator for the ubiquitously expressed receptor CD47. The function of activated CD47 in IRI remains completely unknown. Here, we found that both CD47 and its ligand thrombospondin-1 were upregulated after renal IRI in mice. CD47-knockout mice were protected against renal dysfunction and tubular damage, suggesting that the development of IRI requires intact CD47 signaling. Chimeric CD47-knockout mice engrafted with wild-type hematopoietic cells had significantly lower serum creatinine and less tubular damage than wild-type controls after IRI, suggesting that CD47 signaling in parenchymal cells predominantly mediates renal damage. Treatment with a CD47-blocking antibody protected mice from renal dysfunction and tubular damage compared with an isotype control. Taken together, these data imply that CD47 on parenchymal cells promotes injury after renal ischemia and reperfusion. Therefore, CD47 blockade may have therapeutic potential to prevent or suppress ischemia-reperfusion-mediated damage.


Renal ischemia-reperfusion injury (IRI) is a major cause of ARF resulting from tubular dysfunction, and contributes to morbidity and mortality. In the context of transplantation, IRI promotes delayed graft function and acute rejection, and subsequently negatively influences long-term allograft outcome.1–3 The cascade of events leading to cellular injury and activation is established by initial oxygen deprivation that leads to ATP depletion and mitochondrial dysfunction (reviewed by Murillo et al.4). After reperfusion, proinflammatory conditions and a coordinated release of cytokines, reactive oxygen species (ROS), and adhesion molecules promote the innate immune response to IRI.

Thrombospondin-1 (TSP1) is a matricellular glycoprotein maintained predominantly within platelets and present in nanomolar concentrations in the blood.5 It is secreted by multiple cell types, including inflammatory and vascular cells, typically in response to hypoxia.6 It is also produced by renal tubular epithelial cells (RTECs) after IRI.7 TSP1, through binding the ubiquitously expressed cell surface receptor CD47, regulates the canonical nitric oxide (NO) pathway via limitation of soluble guanylate cyclase activation.8–10 Organs subjected to IRI demonstrate suppression of NO,11 and therapies that supplement NO have been shown to mitigate IRI.12–15

In this study, we tested the hypothesis that renal tissue–expressed CD47 promotes IRI. We demonstrate for the first time that TSP1 and CD47 are significantly induced in IRI. In null mice, absence of activated CD47 leads to profound cytoprotection from renal IRI with significant abrogation of cell death and ROS production, suppressed cytokine production/secretion, and robust reperfusion, as
assessed by laser Doppler analysis. This protection is independent of inflammatory cell activation because null mice transplanted with wild-type (WT) bone marrow, and hence bearing CD47+/+ inflammatory cells, continue to experience substantial protection from renal IRI. Furthermore, we show that limiting CD47 activation with an antibody that prevents TSP1 binding mitigates complications of renal IRI in mice, providing a potential, clinically relevant, therapeutic intervention.

RESULTS

CD47 Is Expressed by RTECs and the TSP1-CD47 Signaling Axis Is Upregulated in Cells Exposed to Hypoxia-Reoxygenation

A single previous study reported upregulation of TSP1 protein after renal IRI,7 although expression of cellular TSP1 and CD47 in the kidney has not been defined. Staining cultured human RTECs for CD47 demonstrated significant cell surface expression (Figure 1A). RTECs challenged with hypoxia (FiO2 1%) and then 24 hours of reoxygenation (as a mimic of IRI) showed a significant induction of TSP1 protein coincident with persistence of CD47 protein, and persistence of both TSP1 and CD47 mRNA, compared with cells treated with normoxia (Figure 1, B and C).

Renal IRI Upregulates Cell Receptor CD47 and its Ligand TSP1

Extending upon results obtained with cultured RTECs, we found robust induction of both TSP1 mRNA and protein expression in WT mice 24 hours after reperfusion, compared with sham-operated animals (Figure 2A). There are no data on expression of its cognate receptor CD47 in response to acute injury, let alone in renal IRI. We now show for the first time concurrent upregulation of CD47 mRNA and protein expression (Figure 2B) in WT mice after IRI. Interestingly, CD47−/− mice showed decreased TSP1 expression compared with WT mice after IRI (Figure 2A).

CD47 Limits Reperfusion after Renal IRI

CD47 activation by its ligand TSP1 blocks NO and soluble guanylate cyclase signaling in multiple cell types.9,16,17 NO directly promotes tissue perfusion and deletion of TSP1 dramatically increases blood flow after NO challenge.8 Using real-time laser Doppler analysis of blood flow to the left kidney (Figure 3A), WT and CD47−/− mice demonstrated comparable basal blood flow and subsequently underwent renal IRI. Initial reperfusion (within 30 minutes) was similar between both groups of animals; however, blood flow was significantly greater in CD47−/− mice at 24 hours after IRI (Figure 3B).

CD47−/− Mice Experience Less Renal Dysfunction and Acute Tubular Damage in a Model of Severe Renal IRI

Activation of proinflammatory pathways and oxidative stress are major contributors to tissue injury after IRI.18 Because the TSP1-CD47 axis has been documented to inhibit tissue blood flow, we hypothesized that CD47−/− mice would experience less tissue injury post-IRI. To further test the importance of CD47 in renal IRI, we compared serum creatinine levels in CD47−/− versus congenic WT (CD47+/+) mice. Knockout of CD47 resulted in less functional renal impairment after 24 hours of reperfusion as assessed by serum creatinine levels (Figure 4A), indicating parenchymal cytoprotection. Clinically, WT mice were grossly uremic and failed to survive past 36–48 hours after reperfusion (data not shown). In contrast, CD47−/− mice did not demonstrate these findings and survived long term (up to 7 days; n=4 per group). To further confirm the presence of renal injury in our model, we evaluated renal histology. After 24 hours of reperfusion, kidneys of WT mice showed widespread and substantial tubular necrosis, tubular dilation, and cast formation. The injury, although being predominantly corticomedullary, also typically extended to the renal cortex. In contrast, kidneys from CD47−/− mice...
displayed significantly less tubular damage 24 hours after IRI (Figure 4B). Thus, CD47 deficiency had a protective effect on renal function and damage after induction of severe IRI.

**Inflammatory Cell Influx Is Decreased in CD47−/− Mice after IRI**

Leukocyte influx is a hallmark of renal IRI. We found significantly fewer interstitial neutrophils (Figure 5A) and macrophages (Figure 5B) in CD47−/− mice at 24 hours after reperfusion compared with WT mice. The number of infiltrating macrophages (in terms of cell number per high-power field) was also significantly less than reported in other studies.19,20 These data suggest an important role for CD47 in the induction of a cellular inflammatory response during IRI.

**Absence of Activated CD47 Reduces Proinflammatory Cytokine and Chemokine Expression after Renal IRI**

We next investigated whether CD47 deficiency also influenced expression of cytokines and chemokines critical to the inflammatory response after IRI. mRNA transcript profiles of IL-6, TNFα, IL-1β, monocyte chemoattractant protein-1 (CCL2), and macrophage inflammatory protein-1 (CXCL2) were examined in kidneys from WT and CD47−/− mice after sham operation or IRI (Figure 6). mRNA expression in WT mice subjected to renal IRI was uniformly increased compared with sham-operated controls. Overall, cytokine expression increased to a much lesser extent in CD47−/− mice and both TNFα and IL-1β transcript levels were not elevated above baseline values.

**CD47−/− Mice Show Reduced Evidence of Programmed Renal Cell Death after IRI**

Renal IRI is also characterized by RTEC apoptosis.21 CD47 ligation by TSP1 regulates cellular apoptotic machinery by modulating Fas signaling22 and has been linked through Bcl-2 homology 3-only protein 19 kD interacting protein-3 (BNIP3) activation to programmed cell death.23 We assessed tubular cell apoptosis by terminal deoxynucleotidyl transferase-mediated digoxigenin-deoxyuridine nick-end labeling (TUNEL) staining and expression of caspase-3, a transducer of programmed cell death. Compared with sham-operated animals, WT mice subjected to IRI demonstrate significant RTEC apoptosis at 24 hours (Figure 7, A and B). This correlated with detection of activated caspase-3 on Western blot (Figure 7, C and D). Notably, the number of apoptotic cells in kidneys and caspase-3 expression were markedly lower in CD47−/− mice at 24 hours after reperfusion.

**Absence of Activated CD47 Mitigates Renal IRI through a Reduction in ROS**

IRI is associated with increased production of ROS, particularly superoxide (O2−), which is a major mediator of cellular injury.24 To assess whether the cytoprotective effect of a lack of CD47 expression could be attributed to a reduction...
in the production of ROS, intracellular generation of the ROS moiety \( \text{O}_2^- \) was visualized with the fluoroprobe dihydroethidium (DHE). Using confocal microscopy, sections of kidney from WT mice showed a widespread and marked increase in DHE fluorescence compared with sham-operated controls (Figure 8A), and this signal was dramatically abrogated in CD47\(^{-/-}\) mice at 24 hours of reperfusion.

Western blot analysis of kidney tissue indicated the presence of nitrated protein bands at 27 and approximately 47 kD (only bands at 47 kD are shown in Figure 8B) in WT mice after 24-hour reperfusion, which was reduced in CD47\(^{-/-}\) mice. Interestingly, basal tissue nitration trended lower in CD47\(^{-/-}\) kidneys compared with WT. Oxidative stress also induces nitric oxide synthase (iNOS) expression, which aggravates tubular injury by supplying additional NO for ONOO\(^-\) formation in the presence of excess ROS.\(^{26,27}\) mRNA expression for renal iNOS was increased in WT mice but significantly attenuated in CD47\(^{-/-}\) mice (Figure 8C).

**CD47-Mediated Kidney IRI Requires Functional CD47 Signaling on Renal Parenchymal Cells**

CD47 is expressed on all circulating cells, including leukocytes, platelets, and red blood cells.\(^{28}\) Thus, it is not clear if results in CD47\(^{-/-}\) mice are secondary to decreased signaling among tissue parenchymal cells or circulating and interstitial leukocytes. To test this, we used bone marrow (BM) transplantation in irradiated CD47\(^{+/+}\) (WT) and CD47\(^{-/-}\) mice to generate chimeric mice expressing CD47 on renal-resident leukocytes but not on renal parenchymal cells (WTBM-CD47\(^{-/-}\)), as well as appropriate control animals. The inability of CD47\(^{-/-}\) BM to engraft WT mice has been documented previously,\(^{29}\) putatively from augmented recipient phagocytosis of donor BM cells in vivo. Recapitulating this observation, we were not able to generate chimeric mice expressing an absence of leukocyte CD47 (CD47\(^{-/-}\)BM-WT) because all mice (\(n=5\)) died in the early post-transplant period (median survival time 19 days). In other BM chimeras, we determined the expression of CD47 on whole blood by flow cytometric analysis 8 weeks after transplantation, demonstrating complete engraftment in all animals (\(n=7\)) (Figure 9A). WTBM-WT mice showed significant kidney dysfunction and injury 24 hours after IRI, whereas CD47\(^{-/-}\)BM-CD47\(^{-/-}\) chimeric mice were substantially protected from kidney IRI as measured by serum creatinine and tubular damage (Figure 9, B and C, respectively). The difference in serum creatinine levels and tubular injury scores recapitulated the results seen previously (see Figure 3). In addition, WTBM-CD47\(^{-/-}\) chimeras were protected from renal IRI to the same degree as CD47\(^{-/-}\)BM-CD47\(^{-/-}\) mice. These results suggest that functional
CD47 on renal parenchymal cells makes the greatest contribution to tissue damage. Interestingly, TSP1 protein expression was decreased after IRI in WTBM-CD47−/− chimeras and even further suppressed in CD47−/− BM-CD47−/− mice (Figure 9D) suggesting that activated leukocytes are a significant source of tissue TSP1 after injury. Conversely, CD47 protein expression was significantly decreased in CD47−/− BM-CD47−/− mice after IRI (Figure 9E). Renal mRNA levels of TSP1 and CD47 were suppressed in CD47−/− mice after IRI regardless of BM genotype.

**Blockade of CD47 Using mAb In Vivo Abrogates Renal IRI**

We have reported soft tissue protection in mice treated with a CD47 blocking antibody.30 However, it was not clear if systemic treatment results in significant tissue localization of the CD47 antibody. To test this, we treated uninjured animals with a CD47 mAb (0.4 μg/g body weight in sterile saline) via a single intraperitoneal injection. Analysis of tissue sections obtained from animals 90 minutes after treatment demonstrated the presence of CD47 antibody on RTECs (Figure 4).
Extending these studies, we evaluated whether blockade of CD47 activation could influence clinical outcome of renal IRI, treating mice with the same dose of the CD47 blocking antibody as above 90 minutes before induction of IRI. Western blot analysis of renal tissue demonstrates downregulation of both CD47 and TSP1 protein expression in WT mice receiving the CD47 blocking antibody compared with mice that received an isotype control antibody (Figure 10B). Mice treated with a CD47 blocking antibody had markedly improved renal function at 24 hours of reperfusion, as reflected by lower serum creatinine levels and decreased tubular damage by light microscopy assessment of tissue sections (Figure 10, C and D), with results recapitulating those achieved in CD47−/− mice after IRI. Moreover, CD47 blockade resulted in global suppression of inflammation after IRI, with significantly fewer tissue neutrophils observed via immunohistology and markedly lower mRNA levels of renal IL-1β, IL-6, CCL2, CXCL2, and iNOS observed on quantitative PCR (Figure 10, E and F). Interestingly, TNF-α mRNA levels were not decreased in CD47 antibody-treated mice after IRI.

**DISCUSSION**

This study has several major findings. First, we provide evidence of strong expression of cell receptor CD47 on human RTECs. In response to the IRI-like conditions of hypoxia and reoxygenation, RTECs demonstrate persistent upregulation of the TSP1-CD47 signaling axis. Results obtained with cultured RTECs were reproduced by results in animals challenged with renal IRI. In WT mice there is rapid upregulation of activated CD47 after renal IRI, providing the first in vivo evidence of injury-mediated CD47 induction. Further support for an instigating role for CD47 in the pathophysiology of renal IRI is provided by studies in CD47−/− mice that are protected from severe renal IRI. Although we have reported increased tissue survival after injury in CD47 null mice,30–32 the exact mechanism involved has remained unclear. We herein demonstrate that, in renal IRI, cytoprotection is conferred predominantly by absence of CD47 expression on RTECs and identify RTECs as major targets of and responders to CD47-mediated IRI. Finally, in translational studies we show that therapeutic blockade of CD47 activation in the kidney decreases renal dysfunction after IRI. Together, these data suggest an important role for tissue-resident activated CD47 in the pathophysiology of renal IRI.

TSP1 is a secreted glycoprotein made by vascular cells, inflammatory cells, and platelets in response to injury, although it is detectable at low concentrations in plasma of healthy individuals.5 In contrast, cell membrane CD47 is expressed ubiquitously6 and TSP1 binding to and activation of CD47 serves an important physiologic function in terms of regulating cardiovascular responses.33 However, nothing is known about the role of CD47 in renal disease. We are the first to show increased CD47 mRNA and protein expression in kidneys subjected to IRI, which occurs concurrently with upregulation of TSP1. Intriguingly, data from CD47−/− and chimeric mice demonstrated that, in the absence of parenchymal CD47, IRI-mediated gene induction of both TSP1 and CD47 did not occur, and are in agreement with our recent report
endothelial cells. These data suggest an active role of the linked TSP1-CD47 gene regulation in hypoxic pulmonary mice (white bars) and WT mice (black bars). *mice (and WT mice treated with a CD47 blocking antibody) may be in part as a consequence of reduced proinflammatory cytokine and chemokine levels (particularly CXCL2), and may account for the lower degree of renal dysfunction and tissue damage compared with WT controls.

Recruitment of leukocytes, particularly neutrophils and macrophages, is a hallmark of IRI and both cell types are actively involved in tissue damage in ischemia-related ARF. The impaired leukocyte influx seen in kidneys from both CD47–/– mice (and WT mice treated with a CD47 blocking antibody) may be in part as a consequence of reduced proinflammatory cytokine and chemokine levels (particularly CXCL2), and may account for the lower degree of renal dysfunction and tissue damage compared with WT controls.

of linked TSP1-CD47 gene regulation in hypoxic pulmonary endothelial cells. These data suggest an active role of the CD47 receptor in affecting pathophysiology in acute renal injury and the possible existence of a common pathway controlling the gene expression of these proteins.

Experimental data suggest that IRI rapidly activates multiple pathways involving renal hemodynamics, proinflammatory mediators, oxidative stress, and the innate immune system (reviewed in Eltzschig and Eckle and Snoeijs et al.). From our data, it is clear that an absence of CD47, despite the presence of its soluble ligand TSP1, significantly abrogates the induction of many of these pathophysiological pathways. WT mice demonstrate substantial ongoing renal perfusion defects at 24 hours after IRI, whereas CD47–/– mice have enhanced blood flow recovery approaching baseline. Improved blood flow may be part of the reparative repertoire seen in CD47–/– mice and reflect significant normalization of renal dysfunction (measured by serum creatinine) and abrogation of tissue damage (preservation of tubular integrity assessed by light microscopy) compared with WT mice.

Programmed cell death remains an important feature of IRI and both apoptosis, assessed by TUNEL staining, and expression of activated caspase-3 were elevated in WT compared with CD47–/– mice after 24 hours of reperfusion. The difference between WT and knockout mice may have been greater if observed at an earlier time point (3–6 hours of reperfusion) that reflects the early phase development of apoptosis in IRI, but apoptotic figures are typically evident at 24 hours of reperfusion. The role of CD47 in programmed cell death has been defined in T cells through an interaction with BNIP3. The involvement of CD47 in promoting apoptosis in other cell types has not been investigated, but the decreased RTEL apoptosis observed in CD47–/– mice may be explained by the proapoptotic function of CD47.

Arguing against a primary role for inflammatory cells in CD47 promotion of renal IRI are results obtained in chimera mice. Mice with CD47–/– myeloid cells after renal IRI. Recruitment of leukocytes, particularly neutrophils and macrophages, is a hallmark of IRI and both cell types are actively involved in tissue damage in ischemia-related ARF. The impaired leukocyte influx seen in kidneys from both CD47–/– mice (and WT mice treated with a CD47 blocking antibody) may be in part as a consequence of reduced proinflammatory cytokine and chemokine levels (particularly CXCL2), and may account for the lower degree of renal dysfunction and tissue damage compared with WT controls.

Lastly, to assess the clinical implications of these findings, we tested whether CD47 blockade could mimic the improved outcome seen in null mice after renal IRI. Immunofluorescent histology provided preliminary evidence that treating via systemic injection with CD47 mAb results in renal cellular and tissue-specific localization of the antibody. These data are important in that the results were obtained in the absence of renal IRI, and suggest the potential to obtain adequate receptor...
targeting as a pretreatment to renal transplant. WT mice treated with a CD47 antibody that blocks its activation demonstrated downregulation of tissue expression of both cell receptor CD47 and ligand TSP1, concurrent with decreased renal damage and pan-suppression of inflammatory cell response and cytokine mRNA after IRI. Results for a single intraperitoneal dose of antibody recapitulated the null phenotypic response after renal IRI and suggest that this therapeutic approach may be efficacious as a mitigant for transplant IRI.

In summary, our results document a new and important role for activated CD47 in mediating RTEC injury in IRI and that inhibition of this pathway is central to cytoprotection from inflammatory and oxidative stress. Significantly, renal IRI appears to be mediated predominantly by tissue-expressed CD47. Elimination of activated CD47 signaling, whether in knockout animals or via antibody blockade, provided robust protection from renal IRI, suggesting a role for CD47 blockade strategies to improve outcomes after procedures and disease processes that effect organ perfusion.

**CONCISE METHODS**

A detailed methods section can be found in the Supplemental Material.

**Animals**

Male C57BL/6 WT and CD47<sup>−/−</sup> mice were obtained from The Jackson Laboratory (Bar Harbor, ME). All studies were performed using protocols approved by the Institutional Animal Care and Use Committee at the University of Pittsburgh and in accordance with National Institutes of Health guidelines.

**Cell-Based Experiments**

Human RTECs (Lonza, Basel, Switzerland) were grown in appropriate media. Cells were serum starved at 80% confluence, subjected to 30-minute hypoxia (FiO₂ 1%) and then 24-hour reoxygenation and collected for RNA or protein (as described below). For immunofluorescence, cells were subjected to cytospin, stained with a CD47 antibody (clone B6H12; Santa Cruz Biotechnology, Santa Cruz, CA) or IgG1 isotype control, and then AlexaFluor 555 (Molecular Probes, Eugene, OR) and 4′,6-diamidino-2-phenylindole (Molecular Probes).

**IRI**

Mice were anesthetized using isoflurane and oxygen titrated to effect, and body temperature maintained at 37°C. Microaneurysm clips were placed to occlude both renal pedicles for 22 minutes. The abdomen was closed with 5/0 monofilament suture. Mice were euthanized 24 hours after reperfusion; blood was collected and kidney tissue snap frozen, placed in RNA later, embedded in optimal cutting temperature compound (OCT; Sakura Finetek, Torrance, CA), or fixed in 10% neutral buffered formalin.

**Figure 8.** Renal oxidative stress is reduced in CD47<sup>−/−</sup> mice after IRI. (A) Representative renal tissue sections from IRI and sham-operated mice (n=6–8 per group) stained with DHE and visualized by confocal microscopy. Intensity of staining was quantified using Image J. *P<0.001 CD47<sup>−/−</sup> after IRI versus WT after IRI. (B) Kidney tissue lysate was prepared from WT or CD47<sup>−/−</sup> mice subjected to sham operation or IRI, resolved by SDS-PAGE, and probed for 3-nitrotyrosine (3NT). Densitometry represents mean ± SD of n=4 samples per group. *P<0.001 CD47<sup>−/−</sup> after IRI versus WT after IRI. (C) mRNA expression profile of iNOS in renal tissue in WT and CD47<sup>−/−</sup> mice subjected to IRI or sham operation. Results have been normalized to the housekeeping gene (HPRT1) and WT sham-operated animals used as the reference group. Data shown are mean ± SD, n=6–10 per group. *P<0.001 CD47<sup>−/−</sup> after IRI versus WT after IRI. Original magnification, ×200.
intraperitoneally in 100 μl sterile PBS) or an IgG2a isotype-matched control antibody (Santa Cruz Biotechnology) injected 90 minutes before surgery. Measurement of creatinine and histology are as described below. Control mice received antibody treatment but did not undergo IRI, and kidneys were embedded in optimal cutting temperature compound. Sections were fixed in ethanol and subsequently stained with AF-555 and 4’,6-diamidino-2-phenylindole.

**Assessment of Renal Function after IRI**

Renal function was assessed by measurement of serum creatinine at 24 hours after IRI using a Jaffe creatinine picric acid reaction (OSR 6178; Beckman Coulter, Brea, CA) analyzed on an Olympus AU640 analyzer (Beckman Coulter).

**Kidney Histology**

Kidneys embedded in paraffin were sectioned at 3 μm and stained with periodic acid–Schiff stain by standard methods. Markers of tubular damage (tubular dilation, cell necrosis, infarction, and cast formation) were scored by calculation of the percentage of tubules in the corticomedullary junction that displayed such features. The scores were as follows: 0, none; 1, 1%–10%; 2, 11%–25%; 3, 26%–45%; 4, 46%–75%; and 5, >75%. Histologic examination was performed blinded on six randomly selected corticomedullary fields (magnification, ×200).

**Immunostaining**

Formalin-fixed, paraffin-embedded sections of 4 μm thickness were deparaffinized and boiled for 30 minutes in 10 mM sodium citrate buffer (pH 6.0). Immunohistochemistry was performed using the following primary antibodies: rat anti-mouse neutrophil (clone Ly6B.2) and macrophage (clone F4/80, both AbDSerotec, Oxford, UK). Sections were exposed to 3% H2O2 in methanol to quench endogenous peroxidases and blocked with 10% normal goat serum and Avidin/Biotin block (Vector Laboratories, Burlingame, CA). The primary antibody was incubated for 2 hours at room temperature, sections were washed with PBS, and then biotinylated goat anti-rat secondary antibody was added in combination with a Vectastain ABC kit (Vector Laboratories) and a metal enhanced 3,3’ diaminobenzidine substrate kit (Thermo Scientific, Waltham, MA). Sections were counterstained for hematoxylin, dehydrated, and covered. Quantification of the cellular infiltrate was performed in a blinded manner by assessing 20 consecutive high-power fields (magnification, ×400).

### Figure 9.

Expression of parenchymal activated CD47 determines kidney damage after IRI. (A) Donor recipient chimerism of the hematopoietic system 8 weeks after BM transplantation in wild-type and CD47−/− mice. Whole blood collected from CD47−/− mice transplanted with WT BM were subjected to FACS analysis of CD47 expression and compared with relevant control groups. Analysis of (B) serum creatinine and (C) tubular damage in respective BM chimeric mice at 24 hours reperfusion after bilateral renal ischemia. Data are mean ± SEM, n=6–7 per group. *P<0.001 WTBM→WT versus WTBM→CD47−/− and CD47−/−BM→CD47−/−. Kidney tissue lysate was prepared from chimeric mice and (D) TSP1 and (E) CD47 protein and mRNA assessed. Data are mean ± SEM, n=6 per group. **P<0.001 WTBM→WT versus WTBM→CD47−/−; ***P=0.001 WTBM→CD47−/− versus CD47−/−BM→CD47−/−; #P<0.0001 compared with WTBM→WT; #P=0.45 WTBM→WT versus WTBM→CD47−/−.

**Renal Doppler Blood Flow Analysis**

Renal perfusion was measured using laser Doppler imaging (MoorLDI-2%; Moor Instruments, Devon, UK). Briefly, animals were anesthetized and core temperature maintained at 37°C. Renal blood flow was assessed at baseline, in response to ischemia and reperfusion at 30 minutes and 24 hours. Results are expressed as the percent change from baseline control of the region of interest.

**CD47 Antibody Treatment**

WT C57BL/6 mice were randomized to receive either anti-mouse CD47 mAb (clone 301, Santa Cruz Biotechnology; 0.4 μg/g intraperitoneally in 100 μl sterile PBS) or an IgG2a isotype-matched control antibody (Santa Cruz Biotechnology) injected 90 minutes before surgery. Measurement of creatinine and histology are as described below. Control mice received antibody treatment but did not undergo IRI, and kidneys were embedded in optimal cutting temperature compound. Sections were fixed in ethanol and subsequently stained with AF-555 and 4’,6-diamidino-2-phenylindole.
Renal Parenchymal Cellular Apoptosis
TUNEL staining was used to detect apoptosis with a commercially available in situ cell death detection kit (Roche, Basel, Switzerland), performed according to the manufacturer’s instructions. Thermal cycling conditions were 50°C for 2 minutes, 95°C for 2 minutes, followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 minute. Data were analyzed using the \( \Delta \Delta Ct \) method with expression normalized to the housekeeping gene and WT sham-operated animals used as the referent control.

Renal Oxidative Stress
DHE (Molecular Probes) was used to evaluate in situ production of superoxide. DHE (10 \( \mu \)M) was applied to unfixed frozen sections, incubated in a light-protected humidified chamber at 37°C for 30 minutes, washed with PBS, and mounted with fluorescent mounting medium (DAKO). Images were quantified using Image J software (http://rsbweb.nih.gov/ij/).

RNA Extraction and Quantiﬁcation by Real-Time PCR
Total RNA was extracted using Qiagen RNeasy Mini Kits (Qiagen, Hilden, Germany) as per the manufacturer’s instructions. RNA was quantiﬁed using the Take3 Gen5 spectrophotometer (BioTek, Winooski, VT). One microgram of RNA was treated with DNase I (ampliﬁcation grade; Invitrogen) and then reverse-transcribed using the Superscript III First Strand Synthesis SuperMix (Invitrogen). cDNA was ampliﬁed using Platinum Quantitative PCR SuperMix-UDG (Invitrogen) in 20-\( \mu \)l volumes in quadruplicate with gene-speciﬁc primers and probe on the ABI Prism 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA), according to the manufacturer’s instructions. Thirty micrograms of total protein was resolved by SDS-PAGE and transferred onto nitrocellulose (BioRad). In blots for CD47, nonreducing Laemmli buffer was used with 8% SDS-PAGE. Blots were probed with primary antibody to the respective proteins and visualized on an Odyssey Imaging System (Lecor, Lincoln, NE). The intensity of the bands was quantiﬁed using Image J.

Western Blotting
Kidney tissue was homogenized in ice-cold lysis buffer. Supernatants were collected and lysates quantiﬁed using a Bradford assay (BioRad, Hercules, CA). Six hours after irradiation, recipient mice were given 1 \( \times \)107 BM cells retro-orbitally. The animals were allowed to recover for 8 weeks to ensure stable engraftment before being subjected to renal IRI. Full chimerism of each mouse was conﬁrmed by ﬂow cytometry of whole blood.

Statistical Analyses
The data are presented as mean ± SD unless otherwise stated. Data were analyzed with the \( t \) test (parametric variables) or Mann–Whitney U test (nonparametric variables) for means between two groups or ANOVA between multiple (>2) groups using STATA software (version 11.0; STATA Corporation, College Station, TX). In all cases, \( P<0.05 \) was deemed signiﬁcant.

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DISCLOSURES

J.S.I. is chair of the scientific advisory board of Vasculox Inc. (St. Louis, MO) and Radiation Control Technologies Inc. (Rockville, MD).

REFERENCES


and (D) quantitative analysis of tubular damage with representative renal tissue sections of corticomedullia from treatment groups 24 hours after ischemia-reperfusion are shown. Data are mean ± SD, n=6 per group. *P<0.001 and **P<0.0001 WT + αCD47 antibody after IRI versus WT + CTRL antibody after IRI. (E) Representative photomicrographs and quantitative analysis of kidney tissue sections stained by immunohistochemistry for neutrophils (total cell number per 10 hpf). Data shown are mean ± SD, n=7 per group and six independent fields assessed. (F) mRNA expression of proinflammatory cytokines IL-6, TNFa, and IL-1β, chemokines CCL2 and CXCL2, and iNOS in kidneys from WT mice pretreated with CTRL or αCD47 antibody and subjected to IRI. Results have been normalized to the housekeeping gene (HPRT1) and WT CTRL antibody animals used as the referent control. Data shown are mean ± SD, n=6 per group. "P<0.0001; *P<0.01; **P=0.002; ***P=0.02. Original magnification, ×400 in A and E; ×200 in C.


18. Rogers NM, Matthews TJ, Kauffman YJ, Kitching AR, Coates PT: Review article: Kidney dendritic cells: Their role in homeostasis, inflammation and transplantation. Nephrology (Carlton) 14: 625–635, 2009


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Methods

Animals. Ten week old male C57BL/6 wild type (WT) and CD47−/− mice (stock numbers 000664 and 003173 respectively) were obtained from The Jackson Laboratory (Bar Harbor, ME). All studies were performed using protocols approved by the Institutional Animal Care and Use Committee (IACUC) at the University of Pittsburgh and in accordance with NIH guidelines.

Cell-based experiments. Human renal tubular epithelial cells (Lonza, Basel, SWI) were grown in appropriate media. Cells were serum starved at 80% confluence, subjected to 30 min hypoxia (FiO2 1%) and then 24 h re-oxygenation and collected for RNA or protein (as described below). For immunofluorescence, cells were subjected to cytopsin (5x10^5 cells/ml, 400rpm, 5 min), fixed in ethanol, stained with αCD47 antibody (clone B6H12, 1:500 dilution, Santa Cruz Biotechnology, Santa Cruz, CA) or IgG1 isotype control (Santa Cruz), then secondary antibody AlexaFluor 555 (Molecular Probes, Carlsbad, CA) and 4′,6-diamidino-2-phenylindole (DAPI, Molecular Probes).

Ischemia-reperfusion injury. Mice (n=10-12 per group) were anaesthetized using isoflurane and oxygen titrated to effect, and body temperature maintained at 37°C. A midline laparotomy incision was made and microaneurysm clips were placed to occlude both renal pedicles for 22 min. After removal of the clips, the kidneys were inspected for restoration of blood flow. The abdomen was closed with 5/0 monofilament suture. Animals received 600µl 0.9% saline and 0.4mg/kg buprenorphine subcutaneously at the end of the procedure. Mice were euthanized 24 h after reperfusion; blood was collected and kidney tissue snap frozen, placed in RNAlater, embedded in OCT compound (Sakura FineTek, Torrance, CA), or fixed in 10% neutral buffered formalin. Sham-operated mice
(n=6) were subjected to anaesthesia and midline laparotomy only.

**Laser Doppler blood flow analysis.** Renal perfusion was measured using laser Doppler imaging (MoorLDI-2%, Moor Instruments, Devon, UK). Briefly, animals were anesthetized and core temperature maintained at 37°C. Renal blood flow was assessed at baseline, in response to ischemia and reperfusion at 30 min and 24 h. Results are expressed as the percent change from baseline control of the region of interest.

**CD47 antibody treatment.** WT C57BL/6 mice were randomized to receive either rat anti-mouse CD47 monoclonal antibody (clone 301, Santa Cruz Biotechnology, 0.4µg/g IP in 100µl sterile PBS) or an IgG2a isotype-matched control antibody (Santa Cruz Biotechnology) injected 90 min before surgery. Measurement of creatinine and histology are as described below. WT mice receiving either antibody not subjected to IRI were euthanized and kidneys embedded in OCT compound. Sections were fixed in ethanol and subsequently stained with AF-555 and DAPI.

**Assessment of renal function after IRI.** Renal function was assessed by measurement of serum creatinine at 24 h after IRI using a Jaffe creatinine picric acid reaction (OSR 6178, Beckman Coulter, Brea, CA) analyzed on an Olympus AU640 analyzer respectively (Beckman Coulter).

**Kidney histology.** Kidneys embedded in paraffin were sectioned at 3µm and stained with Periodic Acid Schiff (PAS) stain by standard methods. Markers of tubular damage (tubular dilatation, cell necrosis, infarction and cast formation) were scored by calculation of the percentage of tubules in the corticomedullary junction that displayed such features: 0, none; 1, 1-10%; 2, 11-25%; 3, 26-45%; 4, 46-75%; 5>75%. Histological examination was performed blinded on 6 randomly selected corticomedullary fields.
Immunostaining. Formalin-fixed, paraffin embedded sections of 4µm thickness were deparaffinized and boiled for 30 min in 10mM sodium citrate buffer (pH 6.0). Immunohistochemistry was performed using the following primary antibodies: rat anti-mouse neutrophil (clone Ly6B.2) and macrophage (clone F4/80, both AbDSerotec, Oxford, UK). Sections were exposed to 3% H2O2 in methanol to quench endogenous peroxidases and blocked with 10% normal goat serum (Sigma Aldrich, St. Louis, MO) and Avidin/Biotin block (Vector Laboratories, Burlingame, CA). The primary antibody was incubated for 2 h at room temperature, sections were washed with PBS, and then biotinylated goat anti-rat secondary antibody was added in combination with a Vectastain ABC kit (Vector Laboratories) and a metal enhanced 3,3’ Diaminobenzidine (DAB) substrate kit (Thermo Scientific, Waltham, MA). Sections were counterstained for hematoxylin (Vector Laboratories), dehydrated and covered. Quantification of the cellular infiltrate was performed in a blinded manner by assessing 20 consecutive hpfs (magnification, x400).

Renal parenchymal cellular apoptosis. TUNEL staining was used to detect apoptosis with a commercially available in situ cell death detection kit (Roche, Basel, SWI), performed according to manufacturer’s instructions. Tissue sections were subsequently counterstained with DAPI, fixed with 4% paraformaldehyde, mounted with mounting medium (DAKO, Glostrup, Denmark) and imaged.

Renal oxidative stress. Dihydroethidium (DHE, Molecular Probes) was used to evaluate in situ production of superoxide. DHE (10µM) was applied to unfixed frozen sections, incubated in a light-protected humidified chamber at 37°C for 30 min, washed with PBS
RNA extraction and quantification by real-time PCR. Total RNA was extracted using Qiagen RNeasy® Mini Kits (Qiagen, Hilden, Germany) as per the manufacturer’s instructions. RNA was quantified using the Take3 Gen5 spectrophotometer (BioTek, Winooski, VT). One microgram (1µg) of RNA was treated with DNase I (amplification grade, Invitrogen) and then reverse-transcribed using the Superscript III First Strand Synthesis Supermix (Invitrogen). cDNA was amplified using Platinum® Quantitative PCR SuperMix-UDG (Invitrogen) in 20µl volumes in quadruplicate with gene specific primers and probe on the ABI Prism 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA), according to manufacturer’s instructions. Thermal cycling conditions were 50°C for 2 min, 95°C for 2 min, followed by 40 cycles of 95°C for 15 sec and 60°C for 1 min. Data were analyzed using the ΔΔCt method with expression normalized to the housekeeping gene and WT-sham-operated animals used as the referent control.

Western blotting. Kidney tissue was homogenized in ice-cold lysis buffer containing NP-40, protease inhibitor cocktail (Sigma Aldrich), 1mM sodium fluoride, 1mM sodium orthovanadate and PhosStop phosphatase inhibitor (Roche), and centrifuged at 12000rpm for 20 min at 4°C. Supernatants were collected and lysates quantified using a Bradford assay (BioRad, Hercules, CA). Thirty micrograms of total protein was resolved by SDS-page and transferred onto nitrocellulose (BioRad). In blots for CD47, non-reducing Laemmlli buffer was used with 8% SDS-PAGE. Blots were probed with primary antibody to the respective proteins and visualized on an Odyssey Imaging System (Licor, Lincoln,
The following antibodies were employed – mouse anti-TSP1 (Abcam, Cambridge, UK, 1:500 dilution); goat anti-CD47 C-18 (Santa Cruz, 1:500 dilution), rabbit anti-caspase-3 (Cell Signaling Technology, Danvers, MA, 1:1000 dilution), mouse anti-3-nitrotyrosine (Abcam, 1:1400 dilution) and rabbit anti-β-actin (Cell Signaling, dilution 1:5000). The intensity of the bands was quantified using Image J.

**Generation of bone marrow (BM) chimeric mice.** BM was collected from WT or CD47⁻/⁻ mice by flushing femurs and tibiae with Hank’s balanced salt solution. Recipient mice were lethally irradiated with 2 doses of 5Gy from a cesium¹³⁷ source using a Mark1 irradiator (JL Shepherd & Associates, San Fernando, CA). Six hours after irradiation, recipient mice received 1x10⁶ BM cells retro-orbitally. The animals were allowed to recover for 8 weeks to ensure stable engraftment before being subjected to renal IRI. Full chimerism of each mouse was confirmed by flow cytometry of whole blood stained for CD47 (clone 301 FITC, BD Pharmingen, Franklin Lakes, NJ) run on a FACSCalibur machine and analyzed using CellQuest software (BD Pharmingen).

**Statistical analysis.** The data are presented as mean ± SD unless otherwise stated. Data were analysed with Student’s t-test (parametric variables) or Mann-Whitney U-test (non-parametric variables) for means between two groups or analysis of variance (ANOVA) between multiple (>2) groups using STATA v11.0 (STATA Corporation, College Station, TX). In all cases, p < 0.05 was deemed significant.