CD73-Dependent Generation of Adenosine and Endothelial Adora2b Signaling Attenuate Diabetic Nephropathy

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ABSTRACT

Nucleotide phosphohydrolysis by the ecto-5′-nucleotidase (CD73) is the main source for extracellular generation of adenosine. Extracellular adenosine subsequently signals through four distinct adenosine A receptors (Adora1, Adora2a, Adora2b, or Adora3). Here, we hypothesized a functional role for CD73-dependent generation and concomitant signaling of extracellular adenosine during diabetic nephropathy. CD73 transcript and protein levels were elevated in the kidneys of diabetic mice. Genetic deletion of CD73 was associated with more severe diabetic nephropathy, whereas treatment with soluble nucleotidase was therapeutic. Transcript levels of renal adenosine receptors showed a selective induction of Adora2b during diabetic nephropathy. In a transgenic reporter mouse, Adora2b expression localized to the vasculature and increased after treatment with streptozotocin. Adora2b/−/− mice experienced more severe diabetic nephropathy, and studies in mice with tissue-specific deletion of Adora2b in tubular epithelia or vascular endothelia implicated endothelial Adora2b signaling in protection from diabetic nephropathy. Finally, treatment with a selective Adora2b agonist (BAY 60–6583) conveyed potent protection from diabetes-associated kidney disease. Taken together, these findings implicate CD73-dependent production of extracellular adenosine and endothelial Adora2b signaling in kidney protection during diabetic nephropathy.


During conditions of inflammation and tissue injury, nucleotides in the form of ATP or ADP are released from multiple cell types.1–4 Nucleotide release pathways include the spillover of ATP from necrotic cells,5 its controlled release from apoptotic cells via pannexin hemichannels6 or from inflammatory or endothelial cells via connexin hemichannels.7,8 and vesicular nucleotide release from platelets.9–12 Once released into the extracellular compartment, ATP and ADP are rapidly converted to adenosine via enzymatically-controlled

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Figure 1. CD73 is induced by diabetic nephropathy. Age-, sex-, and weight-matched mice were subjected to STZ-induced diabetes for 16 weeks until organs were removed for analysis. (A) Wild-type mice show increased renal adenosine concentrations after 16 weeks of diabetes compared with wild-type controls without diabetes (n=3–4). Cd73 mRNA and CD73 protein renal tissue content (quantification by densitometry [n=3] was induced during diabetic nephropathy in (B–D) the STZ-induced diabetic model in wild-type mice, (E–G)
pathways. The pacemaker reaction for the extracellular generation of adenosine is catalyzed by the ecto-5'-nucleotidase CD73, a glycosylphosphatidylinositol-anchored ecto-enzyme that converts AMP into adenosine. Once generated into the extracellular compartment, adenosine can signal through four distinct adenosine receptors (Adora1, Adora2a, Adora2b, or Adora3). Previous studies had implicated CD73-dependent generation of extracellular adenosine and concomitant adenosine signaling in tissue protection during conditions of acute tissue injury, such as AKI, hepatic or intestinal ischemia, or myocardial infarction. In contrast, the functional role of extracellular adenosine generation and signaling during chronic disease states is less clear. For example, extracellular adenosine generation and signaling appears to be protective during the acute phase of lung injury, but has been implicated in promoting a fibrotic response during chronic forms of lung disease.

Diabetic nephropathy is among the leading causes of morbidity and mortality in patients with diabetes mellitus. It belongs to the group of CKDs and is characterized by its progressive nature, eventually leading to nephrotic syndrome and diffuse glomerulosclerosis. It typically occurs in the context of longstanding diabetes mellitus and is among the primary reasons patients require dialysis or kidney transplantation. For example, patients with type 1 diabetes carry a 20%–50% risk of developing ESRD requiring dialysis or renal transplantation. Although effective interventions to slow the progression of diabetes-induced CKD have been described, enhanced therapeutic approaches are urgently needed. Therefore, studies to identify novel therapeutic interventions to prevent diabetic nephropathy or slow its progression are an area of intense investigation and would significantly affect the disease course of diabetes mellitus.

In the present study, we hypothesized a functional role for extracellular adenosine generation and signaling during diabetic nephropathy. Because adenosine responses can differ during acute or chronic disease stages, we pursued these studies without anticipating what the outcome would be. However, we were surprised that combinations of pharmacologic and genetic studies provided a very clear read-out. Indeed, we found that CD73-dependent adenosine generation and signaling through endothelial Adora2b receptors represent an endogenous pathway that protects during diabetic nephropathy and can be targeted for the treatment of kidney diseases caused by chronic elevation of serum glucose levels.

RESULTS

CD73 Transcript and Protein Levels Are Elevated during Diabetic Nephropathy

To study the role of extracellular adenosine generation and signaling during diabetic nephropathy, we induced diabetes in mice at 8 weeks of age by streptozotocin (STZ) treatment. Sixteen weeks after STZ treatment, we performed studies to address the role of extracellular adenosine generation and signaling during diabetic nephropathy. Initial studies in C57BL6 mice showed that renal adenosine levels were significantly elevated 16 weeks after STZ treatment compared with vehicle-treated controls matched by age, sex, and weight (Figure 1A). Because adenosine is generated in the extracellular compartment by enzymatic conversion of AMP to adenosine catalyzed by CD73, we subsequently measured renal CD73 transcript and protein levels. Indeed, we observed that renal CD73 transcript (Figure 1B) and protein levels (Figure 1, C and D) were significantly elevated in mice with STZ-induced diabetic nephropathy. We could confirm an up-regulation of CD73 on a transcript and protein level also in the genetic Akita (Ins2+/−) model (Figure 1, E–G), the STZ-induced eNOS−/− model (Figure 1, H–J), and the db/db model (Figure 1, K–M). These findings demonstrate elevations of renal adenosine levels and concomitant induction of CD73 transcript and protein levels in different models of diabetic nephropathy.

Genetic and Pharmacologic Studies Implicate CD73-Dependent Adenosine Generation in Renal Protection from Diabetic Nephropathy

To demonstrate a functional role for CD73 during diabetic nephropathy, we subsequently performed studies in previously described Cd73−/− mice by inducing diabetes mellitus with STZ treatment and performed measurements 16 weeks afterward. We observed that systolic BP and blood glucose levels were similar in all treatment groups (Supplemental Figure 1), indicating that STZ–diabetes is similar in both groups. Next, we measured renal adenosine levels. We found that baseline levels of renal adenosine between wild-type and Cd73−/− mice were similar (Figure 2A), which is consistent with the notion that CD73-dependent production of extracellular adenosine predominantly occurs during injurious conditions wherein increased levels of precursor nucleotides are being liberated. Importantly, we found that elevations of renal adenosine levels during diabetic nephropathy were almost completely abolished. Moreover, Cd73−/− mice showed a more profound loss of body weight (Figure 2B),
Figure 2. Diabetic nephropathy is increased in Cd73−/− mice. Cd73−/− mice and age-, weight-, and sex-matched wild-type mice were subjected to STZ-induced diabetes for 16 weeks until measurement of (A) renal adenosine content, (B) body weight, (C) kidney weight, (D) urine volume, (E) drink volume, (F) GFR, (G) albumin excretion, (H) renal nephrin transcript, (I) renal VEGF transcript, (J) and urine MCP-1 (n=6–8 in each group). (K) Extracellular matrix deposition as determined by PAS staining. PAS staining was increased in diabetic
whereas their kidney weights were elevated (Figure 2C) in conjunction with elevated urine and drinking volumes (Figure 2, D and E). Moreover, Cd73−/− mice exhibited a more severe degree of renal dysfunction as assessed by measurements of GFR (Figure 2F) and albuminuria (Figure 2G). Similarly, transcript levels of renal nephrin—a protein necessary for the proper functioning of the renal filtration barrier, previously implicated in kidney protection during diabetic nephropathy—was more repressed in Cd73−/− mice, indicating a more severe degree of renal dysfunction. Furthermore, transcript levels of vascular endothelial growth factor (VEGF) and urinary monocyte chemoattractant protein-13 (MCP-1)—both implicated in the pathogenesis of diabetic nephropathy—were more profoundly elevated in Cd73−/− mice than in controls (Figure 2, I and J). In addition, elevations of inflammatory markers (TNF-α and MCP-1) in Cd73−/− mice were specific for the kidneys because there were no differences in the liver or the lungs of diabetic Cd73−/− mice or wild-type littermate controls (Supplemental Figure 2).

Finally, histologic tissue injury in Cd73−/− mice was more severe during diabetic nephropathy, including more profound increases in glomerular size and a higher percentage of glomeruli with mesangial expansion (Figure 2, K–N). Taken together, these findings indicate a protective role for extracellular adenosine production during diabetic nephropathy.

**Pharmacologic Studies Implicate CD73 Function in Renal Protection from Diabetic Nephropathy**

As proof of principle for the assertion that CD73 plays an important role in diabetes, we subsequently reconstituted Cd73−/− mice with soluble 5′-nucleotidase (5-NT). For this purpose, we implanted osmotic pumps (Alzet pump 2006) for continuous subcutaneous 5-NT treatment over 12 weeks. Therefore, the Alzet pumps were implanted at weeks 4 and 10 after the induction of diabetes (STZ, 50 mg/kg on 5 successive days), and kidney function was examined 16 weeks after STZ injection. Systolic BP and blood glucose levels were similar in Cd73 gene-targeted mice with nucleotidase or without nucleotidase treatment (Supplemental Figure 3), indicating that STZ-induced diabetes is similar in both groups. Interestingly, renal adenosine levels in Cd73−/− mice treated with 5-NT returned to higher renal adenosine levels during diabetes, as observed in diabetic wild-type mice (Figures 2A and 3A). Indeed, 5-NT treatment (4 U per mouse per day) of Cd73−/− mice was associated with the reconstitution of a normal phenotype, as shown for their body weight, kidney weight, urine volume, drinking volume, GFR, and albuminuria (Figure 3, B–G). Furthermore, the decrease of renal nephrin transcript in Cd73−/− mice was attenuated and VEGF expression and urinary MCP-1 excretion were decreased (Figure 3, I and J). These findings could also be confirmed on a histologic level (Figure 3, K–N). Taken together, these pharmacologic studies confirm our findings in Cd73−/− mice and implicate CD73-dependent production of extracellular adenosine in kidney protection during diabetic nephropathy.

**Selective Induction of the Adora2b Adenosine Receptor**

On the basis of the above genetic and pharmacologic studies implicating extracellular adenosine generation in kidney protection from diabetic nephropathy, we subsequently examined transcriptional responses of extracellular adenosine receptors during diabetic nephropathy. Indeed, we observed a selective and robust induction of the Adora2b (Figure 4, A–D) in mice with diabetic nephropathy. According to these findings, we next examined Adora2b protein expression using a previously described Adora2b gene reporter mouse. Consistent with previous studies,23,24 we observed that Adora2b protein expression at baseline was predominantly associated with vascular sites (Figure 4E). Furthermore, Adora2b induction at 16 weeks after STZ treatment was dramatically increased at vascular sites (Figure 4E). Moreover, we could show an upregulation of Adora2b protein by Western blot analysis in the STZ-induced diabetic wild-type and eNOS−/− model and in Ins2−/− and db/db mice (Figure 4, F–M). Taken together, these findings indicate vascular induction of the Adora2b during diabetic nephropathy.

**Diabetic Nephropathy Is More Severe in Adora2b−/− Mice**

Based on the above studies showing a selective induction of the renal Adora2b adenosine receptor in mice with diabetes, we subsequently pursued studies in genetic models to address the functional role of the Adora2b. As first step, we induced diabetic nephropathy in previously described Adora2b−/− mice.35,51 Similar to the preceding studies in Cd73−/− mice, we observed a more severe degree of diabetic nephropathy in Adora2b−/− mice without differences regarding systolic BP and blood glucose levels (Supplemental Figure 4, A and B), but a more severe degree of body weight loss (Figure 5A), increased kidney weight (Figure 5B), and elevated urine (Figure 5C) and drinking (Figure 5D) volumes. Determination of the GFR showed a more severe hyperfiltration in Adora2b−/− mice compared with diabetic control mice (Figure 5E). In addition, urinary albumin excretion was significantly increased in diabetic Adora2b−/− mice compared with the wild-type (WT) mice and in particular in Cd73−/− mice (arrows; original magnification, ×400). (L) Histologic score of PAS staining sections.

(M) Glomerular size in all groups. (N) Mesangial expansion. M and N show the result of 30 glomeruli in sections of six mice per group. DM, diabetes mellitus; n.s., not significant. All data are shown as mean ± SD.
Figure 3. Treatment with soluble 5′-NT attenuates diabetic nephropathy in Cd73−/− mice. Cd73−/− mice with or without 5′-NT treatment (from Crotalus atrox venom via Alzet pump, 4 U per mouse per day) and age-, weight-, and sex-matched Cd73−/− mice without 5′-NT treatment were subjected to STZ-induced diabetes for 16 weeks until measurement of (A) renal adenosine content, (B) body weight, (C) kidney weight, (D) urine volume, (E) drink volume, (F) GFR, (G) albumin excretion, (H) renal nephrin transcript, (I) renal
respective diabetic control mice (Figure 5F). Renal nephrin expression was significantly reduced in diabetic Adora2b<sup>−/−</sup> mice compared with the diabetic control group, whereas renal VEGF expression and urinary MCP-1 excretion were significantly increased (Figure 5, G–I). Consistent with previous studies implicating transcriptional mechanisms in the control of hypoxia-inducible factor-α (HIF1-α),<sup>52</sup> we found that HIF1-α mRNA levels were elevated in the kidneys of diabetic Adora2b<sup>−/−</sup> mice. Because HIF1-α is a key transcriptional regulator of VEGF,<sup>1</sup> these findings explain elevated VEGF levels in diabetic Adora2b<sup>−/−</sup> mice (Figure 5J).

Accordingly, histologic characterization of diabetic nephropathy was more severe in Adora2b<sup>−/−</sup> mice (Figure 5, K–N). Furthermore, we could confirm the increase of VEGF transcript in diabetic Adora2<sup>−/−</sup> mice on a protein level showing increased VEGF protein in glomerular epithelia and tubular cells of diabetic Adora2<sup>−/−</sup> mice compared with diabetic wild-type mice (Supplemental Figure 4C). To exclude potential non-specific tissue inflammatory effects that might occur in the STZ model of type 1 diabetes, we used genetic insulin-deficient Akita mice. Therefore, we crossed male Akita mice with female Adora2b<sup>−/−</sup> mice. Ins2<sup>−/−/Adora2b<sup>−/−</sup></sup> mice had a severe decrease in body weight and showed an increase in kidney weight, and drink and urine volumes and albuminuria compared with Ins2<sup>−/−</sup> mice (Supplemental Figure 5, A–G). Taken together, these studies implicate Adora2b signaling in kidney protection during diabetic nephropathy in a drug-induced and genetic model.

**Adora2b-Mediated Kidney Protection during Diabetic Nephropathy Involves Endothelial Adora2b Signaling**

To address the tissue-specific role for Adora2b signaling during diabetic nephropathy, we subsequently examined previously described mice with deletion of the Adora2b in tubular epithelia or vascular endothelial cells.<sup>23</sup> Consistent with the above studies localizing Adora2b induction during diabetic nephropathy to the vasculature, we failed to observe a phenotype in mice with tubular epithelial Adora2b deletion (Adora2b<sup>loxP/loxP</sup>PEPCK-Cre<sup>−</sup>) during diabetic nephropathy (Figure 6, A–H). In contrast, mice with deletion of the Adora2b on vascular endothelia (Adora2b<sup>loxP/loxP</sup>VE-cadherin-Cre<sup>−</sup>) showed a more severe degree of diabetic nephropathy. Their renal phenotype was characterized by a more severe degree of albuminuria (Figure 7A), a more severe degree of vascular and inflammatory parameters (Figure 7, B–D), and a more severe degree of histologic signs for diabetic nephropathy (Figure 7, E–H). The VE-cadherin mouse line has been widely used to investigate endothelial-specific gene deletion in different experimental settings, although Cre expression in this mouse line exists to a lesser extent in hematopoietic cells.<sup>33–35</sup> In conjunction with our studies using Adora2b reporter mice showing almost exclusive expression of the Adora2b in the vasculature of diabetic kidneys, we believe that the most likely explanation of our findings is vascular Adora2b signaling as a mediator of kidney protection during diabetic nephropathy.

**Adora2b Agonist Treatment (BAY 60–6583) Provides Potent Kidney Protection during Diabetic Nephropathy**

On the basis of these genetic findings implicating endothelial Adora2b signaling in kidney protection during diabetic nephropathy, we subsequently pursued pharmacologic studies with the selective Adora2b agonist BAY 60–6583. We had shown in several previous studies that this compound has pharmacologic effects in wild-type or Adora2a<sup>−/−</sup> mice, but not in Adora2b<sup>−/−</sup> mice.<sup>24,31,50,51</sup> For the purpose of our studies, we administered BAY 60–6583 via osmotic pump (0.3 µg per mouse per hour). Indeed, we observed that continuous BAY 60–6583 treatment had no effect on systolic BP or blood glucose levels (Supplemental Figure 6, A and B) but was associated with an attenuated wasting syndrome after STZ treatment (Figure 8, A–D). Hyperfiltration was attenuated in diabetic mice with BAY 60–6583 treatment (Figure 7E). Moreover, changes in albuminuria, renal nephrin and VEGF expression, and urinary MCP-1 excretion, as well as histologic signs of diabetic nephropathy, were attenuated in the mice treated with BAY 60–6583 (Figure 7, G–M). Furthermore, we tested the Adora2b agonist treatment in the genetic diabetic Akita model (Ins2<sup>−/−</sup>). Akita mice treated with the Adora2b agonist BAY 60 6583 showed a less severe phenotype of diabetic nephropathy compared with untreated mice at age 6 months (Supplemental Figure 7, A–G). Systolic BP and blood glucose levels were similar in all groups (Supplemental Figure 7, A and B). Taken together, these data indicate that treatment with a selective Adora2b agonist is associated with significant kidney protection from diabetic nephropathy in the STZ-induced diabetic and the Akita model.

**DISCUSSION**

In the present studies we combined genetic and pharmacologic approaches to investigate the functional roles of extracellular adenosine production and signaling during diabetic nephropathy. Initial findings suggested that renal CD73 transcript and

VEGF transcript, and (J) urine MCP-1 (n=6–8 in each group). (K) Extracellular matrix deposition as determined by PAS staining. PAS staining was attenuated in Cd73<sup>−/−</sup> mice with 5′-NT treatment compared with untreated mice (arrows; original magnification, ×400). (L) Histologic score of PAS staining sections. (M) Glomerular size in all groups. (N) Mesangial expansion. M and N show the result of 30 glomeruli in sections of six mice per group. DM, diabetes mellitus; n.s., not significant. All data are shown as mean±SD.
Figure 4. Adora2b expression is induced during diabetic nephropathy. Age-, sex-, and weight-matched mice were subjected to STZ-induced diabetes for 16 weeks until kidneys were removed for analysis. (A–D) Adenosine receptors (Adora1, Adora2a, Adora2b, and Adora3) transcript levels in kidneys with or without diabetic nephropathy assessed by real-time RT-PCR relative to housekeeping gene β-actin (n=4). (E) Renal tissue of Adora2b reporter mice (Adora2b-KO/β-gal–knockin mice) with or without diabetes were stained for β-galactosidase as an
protein levels are induced during diabetic nephropathy. These findings coincided with elevations of renal adenosine levels that were abolished in gene-targeted mice for CD73, thereby implicating CD73 to function as an endogenous regulator for extracellular adenosine levels during diabetic nephropathy. Indeed, functional studies using *Cd73*−/− mice or treatment with soluble nucleotidase pointed to CD73-dendent adenosine production as an endogenous protective pathway during diabetic nephropathy. Subsequent studies demonstrated a selective induction of Adora2b during diabetic nephropathy, and studies in mice with global or tissue-specific Adora2b deletion identified a protective role for endothelial Adora2b signaling during diabetic nephropathy. Importantly, continuous treatment *via* an osmotic pump delivery system of a selective Adora2b agonist (BAY 60–6583) provided potent protection from diabetic nephropathy, thereby implicating the Adora2b as a therapeutic target.

The results from the present study are consistent with previous findings implicating extracellular adenosine signaling in kidney protection from diabetic nephropathy. For example, a previous study provided compelling evidence that signaling through Adora2a mediates protection from kidney dysfunction during STZ-induced diabetes. These studies were carried out in Sprague-Dawley rats that were treated with the selective Adora2a agonist ATL146e. Indeed, a serious of previous investigations from the laboratory of Dr. Okusa had shown that activation of Adora2a receptors on T cells provides protection during acute ischemic kidney disease. As such, it is conceivable that Adora2a receptors expressed in inflammatory cells and vascular Adora2b receptors function together to protect the kidneys during diabetic nephropathy.

The Adora2b adenosine receptor is the most adenosine “insensitive” adenosine receptor, typically requiring at least micromolar adenosine concentrations for signaling to occur. However, in the context of a chronic inflammatory disease, such increases of extracellular adenosine levels may occur—particularly in a specific microenvironment. Our findings of elevated renal adenosine levels and transcriptional induction of the Adora2b further support this notion. Indeed, previous studies had shown that the Adora2b is highly regulated on a transcriptional level. For example, the stabilization of the transcription factor HIF,1 the stabilization of the transcription factor HIF,1 previous studies had shown that the Adora2b is a classic HIF target gene, and conditions of ambient hypoxia, ischemia, or inflammation are associated with HIF-dependent induction of the Adora2b.30,59,60

The downstream mechanisms of adenosine signaling in the development of chronic forms or renal disease are somewhat controversial. VEGF has been discussed as key factor in the progression of diabetic nephropathy. Abnormal angiogenesis has been shown in human diabetic nephropathy and has been linked to increased VEGF expression.46 VEGF also mediates renal hypertrophy, increases in GFR, and urinary protein excretion in early diabetic nephropathy. The functional role of Adora2b signaling on VEGF release has been controversial, with some studies showing VEGF promotion and other studies showing VEGF reduction.62,63 To our knowledge, the present studies provide the first genetic evidence for a functional role of Adora2b signaling in attenuating renal VEGF levels. Indeed, we found that renal VEGF expression was markedly increased in Adora2b gene-targeted mice after 16 weeks of diabetes compared with wild-type mice. VEGF is mainly produced by podocytes and has a paracrine role to mediate angiogenesis beyond development *via* VEGF receptors on endothelial cells. These findings are consistent with our studies to address the tissue-specific functions of Adora2b signaling. Indeed, we found in an Adora2b reporter mouse model that Adora2b expression is predominantly on the vasculature. Moreover, we found that tissue-specific deletion of Adora2b signaling on vascular endothelial cells conveys increased disease susceptibility during diabetic nephropathy. In fact, Adora2b*loxP/loxP* VE-cadherin-Cre+ mice show dramatically increased levels of VEGF transcript levels during diabetic nephropathy. Together, these findings suggest a potential role for endothelial Adora2b signaling in attenuating VEGF production during diabetic nephropathy as a potential mechanism for adenosine-mediated kidney protection during diabetic nephropathy.

Another possible downstream mechanism could be the known potency of adenosine in mediating the resolution of inflammation and wound healing *via* promoting regulatory T cells. For example, a recent study demonstrated that *Cd73*−/− mice experience a dramatic failure to resolve lipopolysaccharide-induced inflammation due to a lack of regulatory T cells. Similarly, Adora2b signaling has been implicated in promoting regulatory T cells and inflammation resolution. Because T cell activation plays an important role in the pathophysiology of diabetic organ injury, future studies toward a possible role of adenosine in regulating T-cell activation could be of great clinical relevance.
Figure 5. Diabetic nephropathy is increased in Adora2b−/− mice. Adora2b−/− mice and age-, weight-, and sex-matched wild-type (WT) mice were subjected to STZ-induced diabetes for 16 weeks before blood and organs were taken for (A) body weight, (B) kidney weight, (C) urine volume, (D) drink volume, (E) GFR, (F) albumin excretion, (G) renal nephrin transcript, (H) renal VEGF transcript, (I) urine MCP-1, and (J) renal HIF1-α transcript (n=6 in each group). (K) Extracellular matrix deposition as determined by PAS staining. PAS
Taken together, the present studies implicate the CD73-dependent generation of extracellular adenosine and signaling through endothelial-expressed Adora2b receptors in kidney protection during diabetic nephropathy. Future challenges will include the translation of the present findings from bench to bedside. Particularly, it will be critical to move forward with Adora2b agonist treatment in human disease conditions. At present, an Adora2b agonist has never been used in a clinical setting to treat patients.

**CONCISE METHODS**

**Gene-Targeted Mouse Strains**

The animal protocol was approved by the Institutional Animal Care and Use Committee of the University of Colorado Denver and is in accordance with the National Institutes of Health guidelines for use of live animals. Previously described Cdh7−/−, Adora2b−/−, Adora2b−/−/Adora2b−/−/PEPCK-Cre+, Adora2b−/−/VE-cadherin-Cre+ and Adora2b reporter mice (Adora2b–KO/β-gal–knock-in mice) mice were on the C57BL/6 strain or the respective littermate controls, matched for age, sex, and weight, were used.

**Mouse Models of Diabetes**

For the purpose of studying the role of adenosine generation and signaling during diabetic nephropathy, we used four murine models of diabetes (type 1 and 2). In most of our functional studies we used the low-dose STZ model. For this purpose, mice at 8 weeks of age received daily STZ injections intraperitoneally (50 mg/kg, made fresh in 0.1 M citrate buffer, pH 4.5) for 5 consecutive days. Vehicle-injected mice served as controls. Development of diabetes (defined by blood glucose >400 mg/dl) was verified 2 weeks after the first STZ injection.

Another type 1 diabetes mellitus mouse model with more progressive renal vascular injury is the STZ-induced diabetes in eNOS−/− mice (The Jackson Laboratory, Bar Harbor, ME). C57BL/6J mice (C57BL/6) and C57BL/6J-Nos3tm1Unc (eNOS−/− mice; The Jackson Laboratory) that were 8 weeks of age were rendered diabetic by using an established protocol (STZ, 100 mg/kg per day for 2 consecutive days; freshly dissolved in 0.01 M citrate buffer, pH 4.5). Development of diabetes (defined by blood glucose >400 mg/dl) was verified 2 weeks after the first STZ injection. We had an approximately 10% loss of diabetic mice in all STZ-induced diabetes groups during the 16 weeks of investigation.

To exclude potential nonspecific tissue inflammatory effects that might occur in the STZ model of type 1 diabetes, we used genetic insulin-deficient Akita mice. Insulin deficiency is the consequence of a severe pancreatic cell decline resulting from a mutation of the insulin 2 gene and the proteotoxic effect of misfolded insulin.

Male Akita mice (Ins2+/− C57BL/6 background) from The Jackson Laboratories were crossed with female C57BL/6 WT mice. For generation of Ins2+/−/Adora2b−/− double mutants, female Adora2b−/− mice (C57BL/6 background) were crossed with male mice that were heterozygous for both the Ins2 (Akita) and Adora2b mutations. All experiments were performed in male mice at 6 months of age.

To investigate CD73 and Adora receptor regulation also in a type 2 diabetes mouse model, we used the db/db mouse, which has been investigated intensively and exhibits many features similar to human diabetic nephropathy. Db/db mice and their respective controls (db/m) were purchased from The Jackson Laboratories.

**Measurement of Blood Glucose Levels**

Fasting blood glucose levels were examined weekly with a One Touch Ultra Smart blood glucose meter (detection range, 20–600 mg/dl) (LifeScan, Inc.). For glucose measurements in Akita mice, we used a glucose kit from Sigma-Aldrich, GAGO20, to detect values above 600 mg/dl (detection range unlimited depending on prior dilution). To validate the phenotype in our diabetic STZ model, glycated hemoglobin levels were determined in diabetic C57BL/6J mice 16 weeks after STZ treatment. These studies showed hemoglobin A1c concentrations similar to those in previous studies in the STZ model (Supplemental Figure 8).

**Alzet Pump Implantation**

For continuous application of soluble 5-NT (4 U per mouse per day) or the Adora2b agonist BAY 60–6583 (0.3 μg per mouse per hour), Alzet pumps (2006, delivery rate: 0.15 μl/h for 6 weeks) were subcutaneously implanted under isoﬂurane anesthesia. Implantation was performed at week 4 after STZ injections, and Alzet pumps were replaced at week 10 to ensure continuous compound delivery. For BAY 60–6583 (0.3 μg per mouse per hour) treatment in Akita mice, Alzet pumps were implanted at 6 weeks of age and were replaced three times to ensure a continuous treatment for the duration of 18 weeks until mice were euthanized after 6 months of age.

**Determination of GFR**

Inulin clearance was measured 16 weeks after STZ-induced diabetes as described previously. Briefly, mice were anesthetized using pentobarbital, 50 mg/kg intraperitoneally. Animals were then placed on a temperature-controlled operating table to keep rectal temperature at 37°C. The right jugular vein was cannulated for continuous infusion. Blood samples were taken via retroorbital vein plexus puncture. A catheter was placed in the urinary bladder for timed urine collection after removal of the right kidney. After surgery, mice received a bolus of 0.45% sodium chloride solution in an amount equal to 20% body weight. Continuous infusion was maintained at a rate of 800 μl/h per 25 g body weight, and FITC-labeled inulin (0.75 g/100 ml; staining was increased in diabetic wild-type mice and in particular in Adora2b−/− mice (arrows; original magnification, ×400). (L) Histologic score of PAS staining sections. (M) Glomerular size in all groups. (N) Mesangial expansion. M and N show the result of 30 glomeruli in sections of six mice per group. DM, diabetes mellitus; n.s., not significant. All data are shown as mean ± SD.
Sigma-Aldrich) was added to the infusion for evaluation of whole kidney GFR. After stabilization of the animals for 20 minutes, 20-minute timed urine collections were performed. Blood was obtained in the middle of every urine collection period for measurement of FITC-inulin. Concentration of inulin in plasma and urine were determined by measurement of wavelength using a spectrophotometer (Biotek Synergy 2), and GFR was calculated by standard formulas.

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Metabolic Cage Investigation

Experimental mice were placed in metabolic cages (Tecniplast, Italy) for urine collection after 16 weeks of STZ-induced diabetes or at 6 months of age (Ins+/−), respectively. Urine was collected and drinking volume measured. The urinary excretion amount per day and drinking volume per day were calculated. Urinary albumin and MCP-1 excretion were determined. Blood and organs were harvested after metabolic cage investigations and were stored at −80°C until further analysis.

Figure 6. Renal epithelial-specific deletion of the Adora2b has no impact on the severity of diabetic nephropathy. Mice with deletion of the Adora2b in tubular epithelia of the kidneys were generated using the Cre-flox system (Adora2bloxP/loxP PEPCK-Cre+). Adora2bloxP/loxP PEPCK-Cre+ mice and age-, weight-, and sex-matched PEPCK-Cre+ mice were subjected to STZ-induced diabetes for 16 weeks before blood, urine, and organs were taken for measurement of (A) albumin excretion, (B) renal nephrin transcript, (C) renal VEGF transcript, and (D) urine MCP-1 (n=6–8 in each group). (E) Extracellular matrix deposition as determined by PAS staining. PAS staining was increased in diabetic PEPCK-Cre+ mice and in Adora2bloxP/loxP PEPCK-Cre+ mice (arrows; original magnification, ×400). (F) Histologic score of PAS staining sections. (G) Glomerular size in all groups. (H) Mesangial expansion. G and H show the result of 30 glomeruli in sections of six mice per group. DM, diabetes mellitus; n.s., not significant. All data are shown as mean ± SD.
Determination of Albuminuria
Urine was collected over 24 hours via metabolic cage investigations, and mouse albumin ELISA was used by following the manufacturer’s instructions (Exocell).

ELISA Measurements
Blood hemoglobin A1c was measured using an ELISA (A1Cnow) from Bayer AG (Germany). Urine MCP-1 concentrations were determined by ELISA (Pharmingen, San Diego, CA).

Adenosine Measurement via HPLC-Ultraviolet
Whole kidneys from mice with and without diabetes were removed and immediately snap-frozen, and tissue adenosine levels were determined as previously described.23

Transcriptional Studies
We used real-time RT-PCR (iCycler; Bio-Rad Laboratories, Inc.) to examine cd73 and Adora1, Adora2a, Adora2b, and Adora3 expression in renal tissue as previously described.23 Furthermore, we determined...
Figure 8. Adora2b agonist treatment (BAY 60–6583) attenuates diabetic nephropathy. Wild-type mice were subjected to STZ-induced diabetes for 16 weeks. Four weeks after STZ injection and confirmation of diabetes (glucose levels > 400 mg/dl), mice were treated with BAY 60–6583 or vehicle via Alzet pump (0.3 μg per mouse per hour). After 16 weeks of diabetes with or without BAY 60–6583 treatment, blood, urine, and organs were taken for measurement of (A) body weight, (B) kidney weight, (C) urine volume, (D) drink volume, (E) GFR, (F) albumin excretion, (G) renal nephrin transcript, (H) renal VEGF transcript, and (I) urine MCP-1 (n = 6–8 in each group). (J) Extracellular matrix deposition as determined by PAS staining. PAS staining was increased in diabetic wild-type mice and attenuated in BAY 60–6583 treated mice (arrows; original magnification, ×400). (K) Histologic score of PAS staining sections. (L) Glomerular size in all groups. (M) Mesangial expansion. L and M show the result of 30 glomeruli in sections of six mice per group. DM, diabetes mellitus; n.s., not significant. All data are shown as mean ± SD.
Renal transcript of MCP-1, VEGF, nephrin (Applied Biosystems), and HIF1-α (Qiagen; catalog number QT01039542).55,75,76

**Immunoblotting Experiments**

Renal tissues from different diabetic mouse strains with or without diabetes were blotted using polyclonal goat anti-CD73 or anti-Adora2b (Santa Cruz Biotechnology).24,25 Tissues were homogenized and lysed for 10 minutes in ice-cold lysis buffer (150 mM NaCl, 25 mM Tris [pH 8.0], 5 mM EDTA, 2% Triton X-100, and 10% mammalian tissue protease inhibitor cocktail; Sigma-Aldrich), and further processed as previously described.23 To control for protein loading, blots were stripped in stripping buffer for 30 minutes and washed for 10 minutes with Tris-buffered saline and Tween 20, and membranes were blocked for 1 hour at room temperature in PBS-Tween and 4% BSA. Thereafter, the membrane was incubated with β-actin (Abcam, Inc.).

**Renal Histology and Quantification of Morphology**

Kidneys were excised and harvested as previously described.24 Extracellular matrix deposition in glomeruli was assessed by periodic acid–Schiff (PAS) staining. An investigator scored sections in a blinded fashion, according to an established scoring system (range, 0–4: 0, no extracellular matrix deposition; 4, extracellular matrix deposition in all sections of the glomeruli).77 All quantifications were performed in a masked manner. With use of coronal sections of the kidney, 30 consecutive glomeruli per mouse, six mice per group were examined for histologic evaluation. The glomerular area was traced along the outline of the capillary loop using AxioVision image analyzer (Carl Zeiss, Thornwood, NY). The extent of the mesangial expansion was determined by assessing the PAS-positive and nuclei-free area in the mesangium on 30 glomeruli of six mice in each group using ScanScope image analyzer (Aperio Technologies, Vista, CA).78

**Immunohistochemistry**

Formalin-fixed tissues were deparaffinized, rehydrated, and processed for antigen retrieval using 10 mM citrate buffer (pH 6.0) for VEGF (Millipore, Billerica, MA). The sections were incubated with primary antibody overnight at 4°C, followed by treatment with peroxidase-coupled secondary antibody. Color development was achieved using 3,3′-diaminobenzidine.

**Analysis of β-Gal Expression**

To localize the Adora2b in renal tissues, we analyzed β-gal expression in renal sections in Adora2b-KO/β-gal–knockin mice. Kidneys were harvested and fixed as previously described.23

**Statistical Analyses**

Data are presented as mean ± SD from six to eight animals per condition. One-way ANOVA followed by Bonferroni correction was used to compare more than two groups. For comparison between two groups, the unpaired two-tailed t test was performed. All tests were performed using the software program GraphPad Prism (GraphPad Software, San Diego, CA). P < 0.05 was considered to represent a statistically significant difference.

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**DISCLOSURES**

None.
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