Cyclosporine Inhibits the Renal Response to L-Arginine in Human Kidney Transplant Recipients

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ABSTRACT

To evaluate the association of cyclosporine (CsA)-related nephrotoxicity with nitric oxide (NO) and endothelin, the effects of L-arginine (LA) and branched-chain amino acid (BCAA) infusions on renal hemodynamics in 5 normal volunteers and 12 renal transplant recipients were assessed. In normal humans, LA, but not BCAA, reduced mean arterial pressure and renal vascular resistance while increasing RPF and urinary nitrate (NO\textsuperscript{3-}) excretion. Group 1 included six transplant recipients not on CsA; Group 2 subjects (N = 6) were receiving CsA. In both groups, mean arterial pressure declined during the infusion of LA (116 ± 4 to 109 ± 4 mm Hg; P < 0.001) but not BCAA (116 ± 3 to 115 ± 3; P = not significant). In Group 1, LA increased RPF 33 ± 13% (329 ± 48 to 436 ± 77 ml/min per 1.73 m\textsuperscript{2} ; P = 0.01) and GFR 37 ± 16% (95 ± 7 to 130 ± 18 ml/min per 1.73 m\textsuperscript{2}; P = 0.01); renal vascular resistance declined 27 ± 6%. In Group 2, LA did not affect renal hemodynamics. No changes occurred with BCAA in either group. LA increased urinary NO\textsuperscript{3-} excretion by 27 ± 17% in Group 1 (P < 0.05), but only by 16 ± 13% in Group 2 (P = not significant). Urinary endothelin excretion was higher in Group 2 subjects (10.1 ± 1.3 versus 5.3 ± 0.8 pg/mL of GFR, P < 0.01). LA-induced renal vasodilation is associated with the increased urinary excretion of NO\textsuperscript{3-}. The impaired response noted in the presence of CsA could reflect attenuated NO production and/or its local antagonism by a vasoconstrictor such as endothelin.

Key Words: Hypertension, amino acid, renal hemodynamics, nitric oxide, endothelin

During the past decade, cyclosporine (CsA) has become the cornerstone of immunosuppressive therapy in renal transplantation (1,2). However, the utility of CsA is compromised by hypertension and nephrotoxicity, the pathophysiology of which remain incompletely understood (3,4). CsA is a potent vasoconstricter (5). In humans, CsA decreases RBF, increases urea and sodium reabsorption, and impairs renal functional reserve, all consistent with a mechanism involving preglomerular vasoconstriction (6-10).

Although numerous vasoactive substances have been postulated to mediate CsA-induced vasoconstriction, experimental data obtained from animals and humans are inconsistent (11). Recent studies have implicated a newly described vasoconstrictor, endothelin, in the pathogenesis of CsA nephrotoxicity. Originating in epithelial, mesangial, and vascular endothelial cells, endothelin levels have been found to be elevated in animals receiving CsA (12-14). Conversely, endothelium-dependent relaxation, a vascular change that reflects the local production of nitric oxide (NO), is also impaired by CsA (15).

Vascular endothelial cells require L-arginine (LA) to produce NO via the action of the constitutive cytoplasmic enzyme NO synthase (16,17). In uovo, the resultant NO diffuses into adjacent vascular smooth muscle, and via a cGMP-dependent mechanism, induces vasodilation (18). LA analogs, such as nitro-L-arginine methyl ester and N\textsuperscript{-}methyl-L-arginine, when administered to animals, inhibit NO synthase, resulting in elevated systemic blood pressure and renal vasoconstriction (19,20). Alternatively, the chronic administration of LA stimulates NO production and attenuates salt-sensitive hypertension and renal failure in the Dahl/Rapp rat, perhaps via the activation of a second, inducible NO synthase (21,22). To assess the potential roles of altered endothelin and NO production in CsA nephrotoxicity, we administered LA to human renal allograft recipients, postulating that CsA-based immunosuppression might influence renal hemodynamic responses.

METHODS

Subjects

This study was performed in two phases. Initially, five healthy, normotensive volunteers were recruited to establish the feasibility of the protocol. These subjects consisted of three men and two women, aged 25 to 41 yr. Subsequently, 12 male renal allograft recipients were recruited from the kidney transplant clinic at the University of...
Protocol

The protocol was approved by the University of Alabama at Birmingham Institutional Review Board for Human Use, and written, informed consent was obtained from all patients at the time of recruitment. Subjects were admitted to the GCRC and were placed on a diet containing 150 mgE/day of sodium. Baseline laboratory values and a 24-h urine collection for creatinine clearance were obtained. Antihypertensive medications were discontinued after admission. In four subjects who were also receiving a single daily dose of diuretics (one receiving thiazide and three receiving furosemide), their administration was delayed each day until after amino acid infusion. Group 2 subjects continued to receive CsA at their previously stable dose daily every 24 h.

After a 36-h equilibration period, subjects received, on consecutive days in crossover fashion, an infusion of 4% LA (30 g [360 ± 4 mg/kg or 3 ± 1 per minute] in 750 mL of 5% dextrose) (Kabi Pharmacia, Inc., Clayton, NC) and an equimolar, 4% solution of branched-chain amino acids (BCAA: BranchAmin; Clintel Nutrition, Deerfield, IL) containing leucine (1.38 g/dL), isoleucine (1.38 g/dL), and valine (1.24 g/dL). Six subjects received L-arginine before BCAA; in six the order was reversed. The washout period in each subject between amino acid infusions was 20 h. In Group 2 recipients, infusions occurred 18 to 22 h after the previous CsA dose. After the completion of the second amino acid infusion on Day 3, subjects were observed for 2 to 4 h and then discharged.

**TABLE 1. Characteristics of renal transplant recipients comprising Groups 1 and 2**

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Group 1</th>
<th>Group 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>6</td>
<td>6</td>
</tr>
<tr>
<td>Age (yr)</td>
<td>44 ± 5</td>
<td>40 ± 6</td>
</tr>
<tr>
<td>Race (Black:White)</td>
<td>0.6</td>
<td>3.3</td>
</tr>
<tr>
<td>Donor Source (Cadaver:Living)</td>
<td>1:5</td>
<td>5:1</td>
</tr>
<tr>
<td>Time Since Transplant (months)</td>
<td>94 ± 25</td>
<td>22 ± 5*</td>
</tr>
<tr>
<td>Serum Creatinine (mmol/L)</td>
<td>1.6 ± 0.1</td>
<td>1.8 ± 0.1</td>
</tr>
<tr>
<td>Creatinine Clearance (mL/min)</td>
<td>81 ± 6</td>
<td>68 ± 4</td>
</tr>
</tbody>
</table>

* P < 0.01.

Two hours before the study infusions, subjects were given a 20 mL/kg oral water load. An iv catheter was placed in each arm, one for infusion and the other for blood sampling. Subjects then received iv priming boluses of isothalamate (500 mg) (Conray 30; Mallinckrodt Medical, Inc., St. Louis, MO) and para-aminohippurate (PAH, 8 mg/kg) (Merck Research Laboratories, West Point, PA), followed by the continuous infusion of both isothalamate (100 mg/h) and PAH (700 mg/h) over 5 h. After 60 min of equilibration, urine was collected at 30-min intervals and output was replaced orally with water. In the initial normotensive subjects, after three 30-min periods to establish a baseline, a 90-min (three 30-min intervals) continuous infusion of either LA or BCAA was begun. In order to lengthen the observation period for transplant recipients, the amino acid infusion was continued for an additional 30 min (total, 2 h). During baseline and infusion periods, blood was collected at the midpoint of each 30-min interval. Blood pressure was measured by electronic sphygmomanometer every 10 min throughout the study.

**Laboratory Analysis**

Blood was collected in both heparinized tubes, with and without isobutylmethylxanthine, and calcium-EDTA-treated tubes, with and without aprotinin, and was immediately centrifuged. Plasma and urine were frozen at −70°C and then thawed for analysis.

Clearances of isothalamate and PAH were determined by the use of reverse-phase, high-performance liquid chromatography (HPLC) (Waters Division of Millipore, Milford, MA), as described previously (23,24). Two hundred microliters of acetonitrile (J.T. Baker, Inc., Phillipsburg, NJ) containing 0.035% HPLC-grade phosphoric acid (Fisher Scientific, Atlanta, GA) was added to 200 μL of plasma and urine samples. The samples were vortexed and then centrifuged at 3,200 × g for 10 min. Isothalamate and PAH in these samples were separated and quantitated by the injection of 5 μL of the supernatant onto a 30-cm μBondapak C18 reverse-phase column (particle size, 10 mm) (Waters). Standards prepared from stock solutions of isothalamate and PAH were run simultaneously. The mobile phase consisted of a mixture of HPLC-grade NH₄H₂PO₄ (Fisher Scientific) Atlanta, Georgia), 5 mM, and HPLC-grade acetonitrile (J.T. Baker) (98.5:1.5), pH 2.6 ± 0.01. The flow rate was 1.0 mL/min, resulting in an average pressure of approximately 1,000 psi. The detector was set at 236 nm for the analysis of both plasma and urine samples. All HPLC experiments were performed at 30°C with a column heater (Waters). The output detector and pumps were controlled by computer software (Baseline 810 Chromatography Workstation; Dynamic Solutions, Ventura, CA). Standard curves correlating peak height with the concentration of isothalamate and PAH were used to determine the concentrations in plasma and urine. Samples were run in duplicate, and the results were averaged. Isothalamate and PAH clearances were calculated as follows.

\[
C_{\text{isothalamate}} = \frac{(\text{urine flow rate} \times [\text{isothalamate}]_{\text{urine}})}{[\text{isothalamate}]_{\text{plasma}}}
\]

and

\[
C_{\text{PAH}} = \frac{(\text{urine flow rate} \times [\text{PAH}]_{\text{urine}})}{[\text{PAH}]_{\text{plasma}}}
\]

The mean coefficients of variation of these tests of GFR and RPF averaged 8.6 ± 1.8 and 8.2 ± 0.95%, respectively.

Renal vascular resistance (RVR) was calculated as mean arterial pressure (MAP)/RPF × 100 and is reported in resist-
ance units (millimeters of mercury per milliliter per minute × 100). RPF, GFR, and RVR are reported as values per 1.73 m² body surface area. Urinary sodium concentration was measured by flame photometry. Plasma and urine uric acid concentrations were measured with an autoanalyzer (Ektachem 700; Eastman-Kodak, Rochester, NY).

Urinary endothelin was measured with a commercially available, specific enzyme immunometric assay (Cayman Chemical, Ann Arbor, MI), which recognizes endothelin-1, endothelin-2, and endothelin-3, but not big endothelin.

Urinary samples were prepared by passage over a 1-ml C18 reverse-phase cartridge (Bondapak), followed by elution with a methanol:water-trifluoroacetic acid mixture. The sample was then dried by vacuum centrifugation and reconstituted with enzyme immunometric assay buffer. Absorbance was then assayed in an automated plate reader (Thermo Corporation, Menlo Park, CA) and compared with a standard curve to determine endothelin concentrations.

Urinary nitrite (NO$_3^-$) and nitrate (NO$_2^-$) excretion rates were determined as described previously (25,26). Urine samples and sodium nitrite standards (0 to 200 μM) were simultaneously reduced for 1 h at 37°C by Escherichia coli (ATCC 25922; American Type Culture Collection, Rockville, MD) that had been grown under anaerobic conditions. After centrifugation for 10 min at 1,000 × g, 50 μL of supernatant was added to 50 μL of 1.0% sulfanilamide in 30% acetic acid and 50 μL of 0.1% N-(1-naphthyl)ethylene diamine dihydrochloride in 60% acetic acid (Griess reagent). After mixing, the optical density was read at 540 nm with a microplate reader (Thermo Max). To determine nitrite concentration, the protocol was repeated with the same samples, except the E. coli were not added and sodium nitrite, 0 to 300 μM, served as the standard. As expected, urine nitrite (NO$_2^-$) levels were undetectable. The mean coefficient of variation was 5.2 ± 1.2% for the nitrate assay. Urinary nitrate and endothelin excretion are reported as per 1.73 m² body surface area.

Statistical Analysis

For statistical analysis, the mean value of each parameter during baseline and infusion was determined. Each patient served as his own control. When data for a given parameter were normally distributed, a t test was used to compare means. For other data, the Wilcoxon signed rank test was used to compare baseline with infusion values within groups for both LA and BCAA, as well as to compare mean values during LA infusion with those obtained during BCAA infusion. The Mann-Whitney U test was used for intergroup comparisons. Data are presented as mean ± SE. A P value ≤0.05 was interpreted as statistically significant.

RESULTS

In the five normotensive controls, the protocol was well tolerated and without adverse effects. As noted in Table 2 and Figure 1, MAP declined with LA infusion, but did not change with BCAA. Likewise, LA infusion increased RPF and reduced RVR. No change in renal hemodynamics occurred with BCAA infusion. Mean GFR, reflecting wide variability in the data (declines in three subjects and increases in two) remained constant with both infusions. In all five control subjects, urinary NO$_3^-$ excretion increased with LA relative to BCAA infusion (mean percent change: 41 ± 15 versus -1 ± 7; P < 0.05).

Table 3 summarizes data from renal transplant recipients during L-arginine and BCAA infusions. As noted in Table 1, serum creatinine concentration and creatinine clearances did not differ between groups after admission to the GCRC. However, after water loading, baseline GFR was higher in Group 1 than in Group 2 patients (101 ± 7 versus 78 ± 3 mL/min per 1.73 m²; P < 0.01). Although blood pressure was higher in Group 2 patients at baseline, MAP declined significantly in all subjects with LA infusion; BCAA did not affect systemic pressure.

As depicted in Figure 2a, Group 1 subjects responded to LA infusion in a fashion similar to that noted in normal controls. A drop in MAP was accompanied by increased RPF and reduced RVR. Subjects in Group 1 also demonstrated a 37% increase in GFR during LA infusion. As in the normal controls, BCAA produced no changes in renal hemodynamics. The urinary excretion of NO$_3^-$ was higher in transplanted patients than in normal controls and exhibited marked variability. In Group 1, LA infusion increased NO$_3^-$ excretion in five of six subjects by 27 ± 17% (median Δ = 0.73 μmol/min; P < 0.05). BCAA infusion was associated with a 10% decline in urinary NO$_3^-$ excretion.

In Group 2 patients, the decline in systemic blood pressure with LA infusion was not accompanied by changes in renal hemodynamics (Figure 2b). RPF,

<table>
<thead>
<tr>
<th>Parameter</th>
<th>LA</th>
<th>BCAA</th>
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<tbody>
<tr>
<td></td>
<td>Baseline</td>
<td>Infusion</td>
</tr>
<tr>
<td>MAP (mm Hg)</td>
<td>91 ± 5</td>
<td>81 ± 7</td>
</tr>
<tr>
<td>RPF (mL/min per 1.73 m²)</td>
<td>575 ± 50</td>
<td>733 ± 44</td>
</tr>
<tr>
<td>RVR (mm Hg/mL per min × 100)</td>
<td>16 ± 1</td>
<td>11 ± 1</td>
</tr>
<tr>
<td>GFR (mL/min per 1.73 m²)</td>
<td>131 ± 18</td>
<td>120 ± 15</td>
</tr>
<tr>
<td>Urine flow rate (mL/min)</td>
<td>14 ± 2</td>
<td>18 ± 2</td>
</tr>
<tr>
<td>Urinary NO$_3^-$ excretion (μmol/min per 1.73 m²)</td>
<td>0.43 ± 0.13</td>
<td>0.55 ± 0.15</td>
</tr>
<tr>
<td>Urinary endothelin excretion (pg/mL GFR)</td>
<td>3.5 ± 1.1</td>
<td>3.0 ± 0.6</td>
</tr>
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</table>

* P < 0.05, baseline versus infusion.
These changes were accompanied by a modest increase in urinary urate clearance increased by 60% in Group 1 subjects (6.7 ± 0.3 mg/dL; P < 0.05 versus 0.4 versus 4.7 mg/dL; P = 0.001), although uric acid clearance did not differ between the two groups. Subsequently, urate clearance increased by 60% in Group 1 subjects receiving LA (5.7 ± 0.9 to 9.1 ± 1.8 mL/min per 1.73 m²; P < 0.05), whereas no change occurred in Group 2 subjects or in patients from either group receiving BCAA.

At baseline, there was no difference in urinary sodium excretion between groups. However, LA, but not BCAA, increased sodium excretion in both groups (P < 0.05). Plasma uric acid levels were higher at baseline in Group 2 subjects (6.7 ± 0.4 versus 4.7 ± 0.3 mg/dL; P = 0.001), although uric acid clearance did not differ between the two groups. Subsequently, urate clearance increased by 60% in Group 1 subjects receiving LA (5.7 ± 0.9 to 9.1 ± 1.8 mL/min per 1.73 m²; P < 0.05), whereas no change occurred in Group 2 subjects or in patients from either group receiving BCAA.

**DISCUSSION**

In this controlled study, LA reduced MAP and RVR in both normotensive humans and a group of renal transplant patients not receiving CsA-based therapy. These changes were accompanied by a modest increase in urinary nitrate excretion, an increment rendered more significant when compared with the relative reduction in nitrate excretion noted when another nitrogen source (BCAA) was infused. However, in the presence of CsA-based immunosuppression, renal hemodynamics were unaffected by LA, and the increase in urinary nitrate excretion was attenuated. These data support a role for enhanced NO production in mediating the renal vasodilation that accompanies LA administration, a response that is preserved in renal allograft recipients but blunted in the presence of CsA.

A decided but transient increase in RPF and GFR in response to amino acid infusions is a well-described phenomenon in both animals and humans (27–30). More recently, it has become evident that renal hyperemia and hyperfiltration may not represent a universal response to protein loading, but rather an effect of specific amino acids or their metabolites. Although alanine and BCAA do not elicit such changes (31,32), the administration of either L-glycine or LA alone results in renal hemodynamic alterations identical to those previously described with mixed infusions (33–35). Furthermore, Nakaki et al. (36) have documented a hypotensive response to LA in humans. In the normotensive subjects we studied, LA, but not BCAA, reproduced these findings: reduced systemic blood pressure was accompanied by increased RBF and reduced RVR. Unlike previous studies, GFR did not change in these subjects. This result may be artefactual, reflecting the small number studied and the relatively wide variability in measured GFR responses. Alternatively, the LA solution administered to our subjects was isotonic rather than hypertonic; in addition, it was at a dose (3 mg/kg per minute) that was lower than that used by Hirschberg and Koppel to elicit an increase in GFR (6.7 mg/kg per minute), yet higher than that associated with no response (1.7 mg/kg per minute) (33). Although it is possible that such a "midrange" dose might elicit a selective vasodilatory response, the former explanation seems more likely, especially in light of the GFR response noted in the Group 1 transplanted patients.

Several recent studies have linked amino acid-induced renal hyperemia and hyperfiltration to NO generation. King and coworkers found that N'-methyl-L-arginine abolished the renal response to L-glycine and mixed amino acids in the Munich-Wistar rat, a finding corroborated by Tolins and Raji (37,38). Cernadas et al. reported data confirming the abrogation of the renal response to both LA and L-glycine in the rat by pretreatment with N'-nitro-L-arginine, another inhibitor of NO synthase (34). Those studies support an essential role for NO generation in the renal vasodilation that accompanies amino acid infusion, reflecting either a specific effect of LA as substrate for NO synthesis or the activation of NO synthase by another amino acid (glycine) or group of amino acids.

In this study, LA-stimulated NO production appears to have contributed to the renal vasodilation noted in both normotensive humans and Group 1 transplant recipients. In the salt-sensitive Dahl-Rapp rat, LA abrogates hypertensive renal disease, a response that correlates with enhanced urinary nitrate excretion (22). Hibbs and coworkers have shown increased incorporation of radiolabeled guanidino nitrogen atoms of LA into urinary NO₃ during cancer therapy with interleukin-2 (26). At the same time, the labeled nitrogen did not appear in urea, suggesting that urinary NO₃ was a specific marker for LA-induced NO production. In our normal subjects, baseline urinary NO₃ levels approximated those noted by Hibbs and associates and increased substantially in all five (mean change of 41%) with LA relative to BCAA infusion. LA induced similar increases in five of six transplanted Group 1 subjects, versus a 10% decline in
These findings, as well as the marked increase in RPF vasodilatory response to L-glycine, a response only both may be quantitatively different in the renal and systemic circulations. Together, these data indicate that the response to LA experienced no change in renal hemodynamics. Taken into account, these data do not conclusively link renal vasodilation to NO synthesis occurred. Although these data do not conclusively link renal vasodilation to NO production, they support findings of previously noted animal studies in this regard (34,37,38).

Additional hemodynamic effects of LA infusion occurred. In Group 1 subjects, LA reduced both MAP and RVR, with a relatively greater change in RVR. These findings, as well as the marked increase in RPF that accompanied the reduction in systemic pressure, suggest that renal afferent vasodilation exceeded systemic vasodilation. Group 2 subjects, although demonstrating a similar decline in systemic pressure, experienced no change in systemic hemodynamics. Taken together, these data indicate that the response to LA may be quantitatively different in the renal and systemic circulations and that NO, as its mediator, may play a relatively greater role in modulating renal vascular tone. This mechanism remains intact in transplant recipients but may be locally impaired in the presence of CsA.

CsA attenuates the renal response to amino acid infusion in both animals and humans (9,39–41). In a rat model, pretreatment with CsA abrogated the renal vasodilatory response to L-glycine, a response only partially restored by LA (39). Cairns et al. studied renal hemodynamics in nine CsA-treated renal allograft recipients and nine transplanted controls during a 2-h infusion of mixed amino acids (9). GFR and RPF increased by approximately 20% in the non-CsA group and did not change in the CsA group. These findings were corroborated by Rondeau and coworkers using a similar protocol and by Nunez et al. with oral protein loading (40,41). In this study, the NO precursor LA induced renal vasodilation only in those transplanted patients not receiving CsA (Group 1); the increase in RPF was similar to that noted in normal humans. Alternatively, in CsA-treated patients, neither LA nor BCAA altered renal hemodynamics: both the renal vasodilatory effect of LA and the enhanced urinary nitrate excretion were blunted in the presence of CsA.

CsA-induced renal vasoconstriction has been well documented in previous studies (4,10–12). Pathogenetic roles for angiotensin and thromboxanes and increased sympathetic outflow in mediating CsA-related renal vasoconstriction have been postulated, but experimental data are inconsistent (11,42). Some might suggest that long-term CsA administration results in irreversible renal injury, limiting the ability of the kidney to vasodilate in response to any stimulus (43,44). However, other studies, in both animals and humans, have shown prolonged reversibility of CsA-induced vasoconstriction: the discontinuation of CsA and the administration of calcium antagonists or angiotensin-converting enzyme inhibitors reduce RVR (6,45,46). Our data indicate that, in humans, an aberrant response to endothelial mediators may contribute to clinical CsA nephrotoxicity: the failure of the

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**TABLE 3. Renal and hemodynamic parameters during the infusion of LA and BCAA in renal transplant recipients, by group**

<table>
<thead>
<tr>
<th>Group</th>
<th>LA</th>
<th>BCAA</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Baseline</td>
<td>Infusion</td>
</tr>
<tr>
<td>MAP (mm Hg)</td>
<td>114 ± 6</td>
<td>107 ± 5&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>RPF (mL/min per 1.73 m&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>329 ± 48</td>
<td>436 ± 77&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>RVR (mm Hg/mL per min × 100)</td>
<td>30 ± 6</td>
<td>22 ± 5&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>GFR (mL/min per 1.73 m&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>95 ± 7</td>
<td>130 ± 18&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>Urine flow rate (mL/min)</td>
<td>12.3 ± 1.3</td>
<td>14.8 ± 2.0</td>
</tr>
<tr>
<td>Urinary sodium excretion (mEq/min)</td>
<td>0.18 ± 0.04</td>
<td>0.23 ± 0.05&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Uric acid clearance (mL/min per 1.73 m&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>5.7 ± 0.9</td>
<td>9.1 ± 1.8&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Urinary NO&lt;sub&gt;3&lt;/sub&gt; excretion (mEq/min per 1.73 m&lt;sup&gt;2&lt;/sup&gt;)</td>
<td>1.33 ± 0.28</td>
<td>1.60 ± 0.34&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>Urinary endotoxin excretion (pg/mL GFR)</td>
<td>6.8 ± 1.3</td>
<td>6.3 ± 1.1</td>
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<sup>a</sup>P < 0.05, baseline versus infusion.
<sup>b</sup>P < 0.05, LA infusion versus BCAA infusion.

urinary NO<sub>3</sub> excretion with BCAA. The increased urinary excretion of NO<sub>3</sub> accompanying the infusion of LA, but not another nitrogen source (BCAA), implies that the response to LA was specific and indicates that enhanced NO synthesis occurred. Although these data do not conclusively link renal vasodilation to NO production, they support findings of previously noted animal studies in this regard (34,37,38).
With the work of Dinh-Xuan in transplanted subjects, the inhibition of NO production (consistent with vasodilate in CsA-treated recipients) could reflect increased CsA-induced renal dysfunction and/or endothelin secretion, although not elevated to the extent noted by Perico and colleagues at peak CsA levels, was substantially higher than in transplant recipients not on CsA. Likewise, urinary endothelin excretion, although not elevated to the extent noted by Perico and colleagues at peak CsA levels, was substantially greater in CsA-treated patients. Our data confirm that, even in the presence of nadir CsA concentrations, renal hemodynamics and endothelin excretion remain abnormal. Awazu and associates found CsA to preferentially up-regulate endothelin binding in renal tissue compared with hepatic tissue, providing a potential link between increased endothelin production and renal-specific effects.

Thus, these data are consistent with previous studies indicating a substantial role for NO in mediating the vasodilatory effects of LA. They likewise support the hypothesis that CsA nephrotoxicity in humans is a complex phenomenon that may reflect, at least in part, an adverse imbalance between the effects of the vasodilator NO and the vasoconstrictor endothelin within the renal circulation. Future studies using specific inhibitors of NO synthase and/or endothelin receptor antagonists may further characterize the relative contributions of each of these pathogenetic influences in humans.

ACKNOWLEDGMENTS

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REFERENCES

Cyclosporine and L-Arginine in Humans


Vepacian, one of the great schoolmasters of avarice, which could pick out profit of every thing (yea even of mens urine) taught his Scholars (I mean the whole court of covetous persons) this lesson ensuing:

Lucrifonus odor cace qualibet.
Lucre is sweet, and hath a good savour,
Though it come of urine, dirt or ordure.

So that if there be any Physician so arrogant, that he will take upon him to tell all things alone and will not hear the Patient speak, specially not knowing the party before, neither seeing other signs but only the urine, as I dare boldly pronounce, that such a man is unworthy to be called a Physician.

Robert Record, The Urinal of Physick, printed by Gartrude Dawson, London, 1651. From the collection of the Clendening Library of the History of Medicine, University of Kansas.