Reduced Production of Creatinine Limits Its Use as Marker of Kidney Injury in Sepsis

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ABSTRACT

Although diagnosis and staging of acute kidney injury uses serum creatinine, acute changes in creatinine lag behind both renal injury and recovery. The risk for mortality increases when acute kidney injury accompanies sepsis; therefore, we sought to explore the limitations of serum creatinine in this setting. In mice, induction of sepsis by cecal ligation and puncture in bilaterally nephrectomized mice increased markers of nonrenal organ injury and serum TNF-α. Serum creatinine, however, was significantly lower in septic animals than in animals subjected to bilateral nephrectomy and sham cecal ligation and puncture. Under these conditions treatment with chloroquine decreased nonrenal organ injury markers but paradoxically increased serum creatinine. Sepsis dramatically decreased production of creatinine in nephrectomized mice, without changes in body weight, hematocrit, or extracellular fluid volume. In conclusion, sepsis reduces production of creatinine, which blunts the increase in serum creatinine after sepsis, potentially limiting the early detection of acute kidney injury. This may partially explain why small absolute increases in serum creatinine levels are associated with poor clinical outcomes. These data support the need for new biomarkers that provide better measures of renal injury, especially in patients with sepsis.

First, we evaluated sepsis in bilaterally nephrectomized mice. We induced sepsis by CLP surgery with 8-mm cecal ligation, which causes modest sublethal sepsis in normal outbred CD-1 mice. All animals survived until they were killed at 18 h after surgery. CLP surgery in non-nephrectomized mice caused a numerically small, but not significant increase of serum creatinine (shamBNx/CLP group). As expected, we found large increases of serum creatinine in the BNx/shamCLP group; however, induction of sepsis at the time of BNx significantly decreased serum creatinine (BNx+CLP group) compared with nonseptic BNx alone (BNx/shamCLP group; Figure 1A), raising doubt about whether serum creatinine accurately reflects impaired kidney function during sepsis. In contrast, nonrenal organ injury markers (aspartate aminotransferase [AST], alanine aminotransferase [ALT], and lactate dehydrogenase [LDH]) and serum TNF-α were higher in the BNx+CLP group than in the BNx+shamCLP group, confirming the presence of severe sepsis (Figure 1, B through E).
We next examined whether treatment of sepsis would affect this observation by using chloroquine, which decreases mortality and improves organ function in a model of CLP-induced sepsis. Chloroquine treatment of septic bilaterally nephrectomized mice (BNx+CLP+CQ) decreased nonrenal organ injury markers (AST, ALT, and LDH) and TNF-α compared with vehicle-treated mice (BNx+CLP); however, serum creatinine was paradoxically higher in treated animals (Figure 2). As a control, chloroquine did not cause any changes in the BNx+sham group. We reported recently that chloroquine reduced mortality and sepsis-induced AKI, including renal pathologic damage. At first glance, these results seem contradictory: Serum creatinine is reduced by chloroquine when kidneys are intact yet increased by chloroquine in the absence of any kidneys. In the septic state, chloroquine alters nonrenal metabolism of creatinine, which is confirmed by completely removing urinary creatinine excretion by BNx; therefore, in both untreated and treated septic mice, serum creatinine changes in the opposite direction as markers of injury from other organs, challenging the underlying assumptions behind the use of serum creatinine as a biomarker for AKI.

To determine whether changes in fluid compartments could account for these observations, we measured body weight changes and hematocrit, as well as volume of distribution (Vd) for FITC-inulin and creatinine. There was no significant difference in body weight change or hematocrit (body weight change: BNx+sham <sub>CLP</sub> 2.3 ± 1.0% [n = 5], BNx+CLP 1.7 ± 0.6% [n = 5]; hematocrit: BNx+sham <sub>CLP</sub> 36.0 ± 0.9% [n = 6], BNx+CLP 34.6 ± 1.3% [n = 6]). Creatinine space, a marker of total body water, was not different between the BNx+CLP and the BNx+sham <sub>CLP</sub> groups (Figure 3A). FITC-inulin space, a marker of extracellular fluid volume, was only modestly decreased in the BNx+CLP group (Figure 3A). We then estimated creatinine production from the increase in serum creatinine between time 0 and 14 h and the creatinine Vd. We found that sepsis significantly decreased estimated creatinine production by 29.7 ± 4.3% (Figure 3B). Direct measurement of creatinine production would be necessary to confirm our result.

Creatinine is the end product of creatine metabolism. Alteration in creatinine production can alter serum creatinine levels. For example, N-acetylcysteine, which has been shown to protect from radiocontrast medium–induced nephropathy, decreases serum creatinine levels of healthy humans without changing serum cystatin C levels. This suggests that N-acetylcysteine directly reduces creatinine production. We suspected that creatinine production was reduced, on the basis of the small increase in serum creatinine after CLP sepsis versus renal ischemia-reperfusion. In this study, we found that sepsis reduced creatinine production, which would largely account for the slow rise in serum creatinine. Serum creatinine could fall as a result of reduced production or renal or extrarenal clearance (creatinine degradation or gastrointestinal excretion). Creatine is synthesized from guanidinoacetate acid primarily in the liver, enters skeletal muscle via a membrane transporter, and accumulates there because it is phosphorylated by creatine kinase. Creatine is converted into creatinine by a nonenzymatic cyclization throughout the body but especially in skeletal muscle as a result of the high abundance of creatine. Creatinine production can fall because of reductions in lean body mass, dietary intake of creatine, or liver disease. Indeed, the BNx+CLP group showed significantly higher liver enzymes and pathologic changes such as extensive loss of hepatocyte glycogen stores and bland cytoplasm (data not shown). Intensive care unit patients have a progressive decline in creatinine production as a result of a loss of muscle mass worsened by subclinical hepatic injury.

Sepsis reduces energy production and metabolic rate because of hormonal and inflammatory mediators, which could reduce muscle production of creatinine. As we previously reported, CLP induced severe septic shock with hypodynamic circulation failure and reduced micro-
vascular capillary perfusion, which could reduce muscle creatinine release, liver creatine-to-creatinine conversion, and/or release into the circulation. In this study, the numerical decrease in creatinine space in the BNx/CLP group is consistent with reduced cellular perfusion in sepsis. Because our CLP model has a prolonged period of hypodynamic shock, our results may not translate to patients with hyperdynamic shock. Examination of creatinine production in hyperdynamic animal sepsis models would be needed to determine the extent that microvascular perfusion may contribute to the decrease in creatinine production during hyperdynamic sepsis.

Sepsis-induced hypothermia may also decrease nonenzymatic conversion of creatine to creatinine. CLP sepsis causes profound hypothermia, which is a marker of reduced metabolism. In this study, we found that the severe hypothermia in the BNx/CLP animals was attenuated by chloroquine treatment (data not shown). Creatinine is converted from creatine and creatine phosphate nonenzymatically, and the rate of nonenzymatic conversion of creatine to creatinine depends on the pH and temperature; a 3°C decrease would reduce this conversion by 15 to 20%. In addition, decreasing metabolism by systemic injection of drugs such as 5’-adenosine monophosphate (AMP) or 2-deoxy-D-glucose (2-DG) instead of sepsis to bilaterally nephrectomized mice also caused hypothermia and decreases of serum creatinine 18 h after surgery (Supplemental Figure 1).

Our findings may have direct clinical significance. As a result of decreased creatinine production, serum creatinine is an even poorer indicator of renal damage in sepsis because its reduced production further magnifies the kinetic disparity between apparent and actual renal function; serum creatinine underestimates renal damage to a greater extent in sepsis than in other forms of AKI. The reduced creatinine production may also explain why such small increases in serum creatinine are associated with dramatic increases in morbidity and mortality of human patients. Higher serum creatinine was paradoxically associated with better survival in AKI in several clinical studies. Underestimation of renal function changes by “inappropriately low” serum creatinine will delay the early diagnosis of AKI, impede recognition of an additional organ injury for prognostic purposes, and suppress entry into clinical trials. Although the timing of renal replacement therapy initiation (including prophylactic dialysis) is controversial, renal replacement therapy may need to be started at relatively lower serum creatinine than other types of AKI.

The paradoxic effect of chloroquine on creatinine in bilaterally nephrectomized animals also has implications for clinical trial design. Chloroquine im-
proved sepsis and sepsis-induced AKI by survival analysis and pathologic examination. We hypothesize that chloroquine restores creatinine metabolism indirectly by reducing the severity of sepsis, perhaps by improvement in muscle metabolism and/or liver function. Although serum creatinine is widely used as an end point for AKI clinical studies, the effects of treatment of sepsis and sepsis-induced AKI might be incorrectly ascertained if kidney damage in sepsis is evaluated only by serum creatinine. Confirmation with other end points such as injury biomarkers and mortality and morbidity rate should be required, especially for clinical studies on sepsis.

In conclusion, we demonstrated that sepsis reduced creatinine production, thereby blunting the expected increases of serum creatinine in bilaterally nephrectomized mice. Moreover, treatment with chloroquine reduced improved sepsis but paradoxically increased serum creatinine in bilaterally nephrectomized mice, suggesting a normalization of creatinine production. Our data indicate that evaluation of kidney injury by serum creatinine alone would cause a severe underestimation of renal injury, serious failure of early diagnosis of sepsis-induced AKI, and incorrect ascertainment of drug effects. Newer biomarkers that more accurately measure renal injury are needed, especially in patients with sepsis.

**Concise Methods**

**BNx and Subsequent CLP**

All animal experiments were conducted in accordance with an animal study protocol approved by the National Institute of Diabetes and Digestive and Kidney Diseases animal care and use committee. Eight-week-old male CD-1 mice (Charles River Laboratories, Wilmington, MA) weighing 30 to 35 g were allowed food and water *ad libitum*. For each experiment, all of the animals received BNx or its sham operation (shamBNx), and CLP or its sham operation (shamCLP). Under isofluorane anesthesia, the kidneys were removed by flank approach, or kidneys were isolated in sham surgeries. Immediately after BNx, sepsis was induced by CLP surgery. A 4-0 silk ligature was placed 8 mm from the cecal tip, and the cecum was punctured twice with a 21-G needle and gently squeezed. At the end of surgery, 1 ml of prewarmed normal saline was injected intraperitoneally, and mice were subsequently kept at 29°C. Mice were treated with fluid and antibiotic at 6 h after surgery by subcutaneous injection of imipenem/cilastatin (14 mg/kg) in 1 ml of normal saline and killed at 18 h after for collecting specimens. Mice were not allowed food and water after the surgery.

**Chloroquine, AMP, and 2-DG Administration**

Chloroquine (Sigma-Aldrich, St. Louis, MO) was dissolved in normal saline at the dosage of 50 mg/kg was administered orally at 3 h before surgery. In other animals, AMP (Sigma-Aldrich) at the dosage of 1.5 mmol/kg or 2-DG (Sigma-Aldrich) at the dosage of 400 mg/kg dissolved in 0.3 ml of normal saline was injected intraperitoneally at 0, 3, and 6 h after surgery. All of the animals in the other group received the same amount of vehicle at the indicated times.

**Measurements of Blood Chemistries, Hematocrit, and Serum Cytokine**

Serum creatinine was measured by HPLC, which has a coefficient of variation of 1.6 to 5.1%. AST, ALT, and LDH were measured using an autoanalyzer (Hitachi 917; Boehringer-Mannheim, Indianapolis, IN). Hematocrit was analyzed by an automated hematocrit analyzer (Abbott Cell-Dyn 3500; GMI, Ramsey, MN). Serum TNF-α was measured by ELISA (R&D Systems, Minneapolis, MN).

**Measurement of Vd and Creatinine Production**

Inulin-labeled FITC preparation and measurement of plasma concentration was performed as described previously. Inulin-labeled FITC was injected intravenously at 30 min after surgery, and its Vd was calculated from the amount injected and the plasma concentration at 18 h after injection. Creatinine Vd and production were measured using injection of a known amount of unlabeled creatinine, as previously reported in bilaterally nephrectomized dogs. Creatinine (30 mg/kg) was injected intravenously at 14 h after surgery, and blood samples were collected before injection and 4 h later. Vd of creatinine in nephrectomized mice was calculated from the 4-h increase in serum creatinine in the vehicle-injected mice (n = 6 to 8 per group). The creatinine production in each mouse was estimated from the time 0 to 14 h increase in serum creatinine multiplied by the creatinine Vd determined in that mouse. Creatinine at 0 h was assumed to be distributed in a Vd of 60% of body weight, which is approximately the same of total body water. Because the normal mouse creatinine is so low compared with the creatinine after BNx, the calculated creatinine production is not sensitive to the time 0 Vd. Nevertheless, a more precise value could be directly measured after equilibrating the mouse creatine compartment with radiolabel.

**Statistical Analysis**

Results are expressed as means ± SEM. Differences between groups were analyzed by ANOVA followed by Fisher least significant difference test. These calculations were done using SigmaStat 3.10 (Systat Software, Richmond, CA). The null hypothesis was rejected at P < 0.05.

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**DISCLOSURES**

None.

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